

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
22 March 2001 (22.03.2001)

PCT

(10) International Publication Number
WO 01/20002 A1

(51) International Patent Classification⁷: C12N 15/53, 15/10, 9/02

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(21) International Application Number: PCT/US99/30925

(81) Designated States (national): AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.

(22) International Filing Date: 22 December 1999 (22.12.1999)

(25) Filing Language: English

(26) Publication Language: English

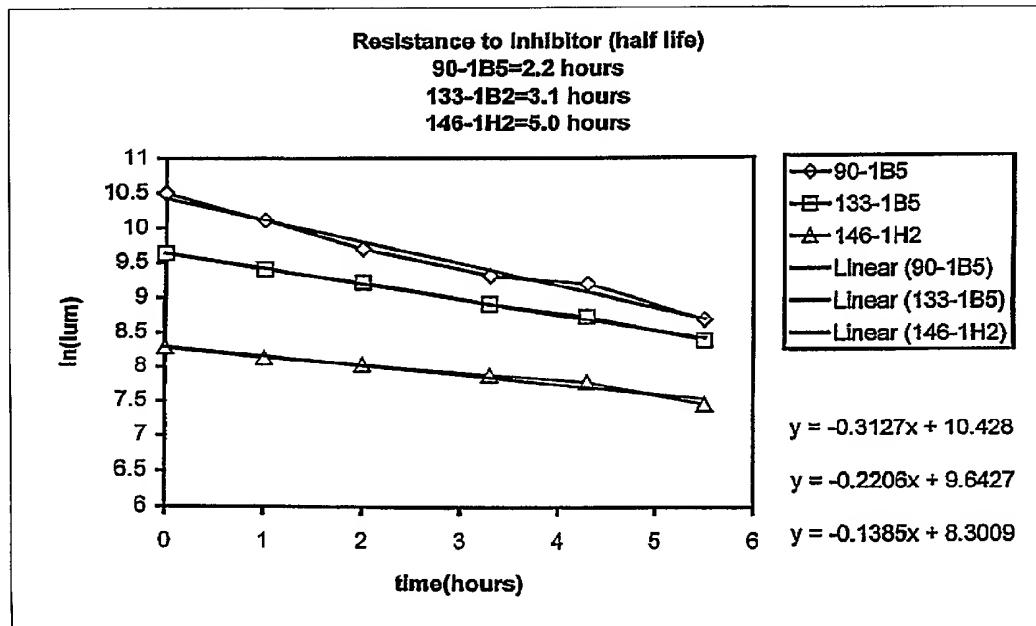
(30) Priority Data: 09/396,154 15 September 1999 (15.09.1999) US

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(54) Title: THERMOSTABLE LUCIFERASES FROM PHOTURIS PENNSYLVANICA AND PYROPHORUS PLAGIOPHTHALAMUS AND METHODS OF PRODUCTION



WO 01/20002 A1

(57) Abstract: Luciferase enzymes with greatly increased thermostability, e.g., at least half lives of 2 hours at 50 °C, cDNAs encoding the novel luciferases, and hosts transformed to express the luciferases, are disclosed. Methods of producing the luciferases include recursive mutagenesis. The luciferases are used in conventional methods, some employing kits.

**THERMOSTABLE LUCIFERASES FROM PHOTURIS PENNSYLVANICA AND PYROPHORUS
PLAGIOPHTHALAMUS AND METHODS OF PRODUCTION**

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Statement of Government Rights

The invention was made with grants from the Government of the United States of America (grants 1R43 GM506 23-01 and 2R44 GM506 23-02 from the National Institutes of Health and grants ISI-9160613 and III-9301865 from the National Science Foundation). The Government may have certain rights to the 10 invention.

Field of the Invention

The invention is directed to mutant luciferase enzymes having greatly increased thermostability compared to natural luciferases or to luciferases from which they are derived as measured, e.g., by half-lives of at least 2 hours at 50°C 15 in aqueous solution. The invention includes mutant luciferase enzymes that are resistant to inhibition by a substrate inhibitor, e.g., a substrate analog. The invention is also drawn to polynucleotides encoding the novel luciferases, and to hosts transformed to express the luciferases. The invention is further drawn to methods of producing luciferases with increased thermostability and the use of 20 these luciferases in any method in which previously known luciferases are conventionally employed. Some of the uses employ kits. The invention also provides a method of producing a polynucleotide sequence encoding an enzyme that is resistant to inhibition by an inhibitor, and a method which yields a 25 polynucleotide sequence encoding an enzyme having enhanced enzymological properties.

Background of the Invention

Luciferases are defined by their ability to produce luminescence. Beetle luciferases form a distinct class with unique evolutionary origins and chemical mechanisms (Wood, 1995).

30 Although the enzymes known as beetle luciferases are widely recognized for their use in highly sensitive luminescent assays, their general utility has been limited due to low thermostability. Beetle luciferases having amino acid sequences encoded by cDNA sequences cloned from luminous beetles are not stable even at moderate temperatures. For example, even the most stable of the

luciferases, *LucPpe2*, obtained from a firefly has very little stability at the moderate temperature of 37°C. Firefly luciferases are a sub-group of the beetle luciferases. Historically, the term “firefly luciferase” referred to the enzyme *LucPpy* from a single species *Photinus pyralis* (Luc + is a mutant version of 5 *LucPpy*, see U.S. Patent No. 5,670,356).

Attempts have been reported to mutate natural cDNA sequences encoding luciferase and to select mutants for improved thermostability (White et al., 1994; from *P. pyralis*, and Kajiyama and Nekano, 1993; from *Luciola lateralis*.) However, there is still a need to improve the characteristics and 10 versatility of this important class of enzymes.

Summary of the Invention

The invention is drawn to novel and remarkably thermostable luciferases, including luciferase enzymes with half-lives of at least 2 hours at 50°C, or at least 5 hours at 50°C, in an aqueous solution. As described hereinbelow, after 2 15 hours at 50°C in an aqueous solution, a thermostable luciferase of the invention lost less than 5% luminescence activity. The mutant luciferases of the present invention display remarkable and heretofore unrealized thermostability at 22°C in an aqueous solution and at temperatures at least as high as 60°C in an aqueous solution. For example, the luciferases of the invention are thermostable for at 20 least 10 hours at 50°C; for at least 2 hours, preferably at least 5 hours, more preferably at least 10 hours, and even more preferably at least 24 hours, at 60°C; and/or for at least 100 days, preferably at least 200 days, more preferably at least 500 days, and even more preferably at least 800 days, at 22°C, in aqueous solution. For example, after 30 days at 22°C in an aqueous solution, a 25 thermostable luciferase of the invention lost less than 5% luminescence activity. Preferably, the thermostable luciferases of the invention have enhanced luminescence intensity, enhanced signal stability, enhanced substrate utilization, and/or decreased Km, relative to a reference, e.g., a native wild-type, luciferase. The invention is further directed to the mutant luciferase genes (e.g., cDNA or 30 RNA) which encode the novel luciferase enzymes. The terminology used herein is, e.g., for the mutants isolated in experiment 90, plate number 1, well B5, the *E. coli* strain is 90-1B5, the mutant gene is *luc90-1B5*, and the mutated luciferase is *Luc90-1B5*.

As defined herein, a "thermostable" enzyme, e.g., a luciferase, or an enzyme which has "thermostability", is an enzyme which under certain conditions, e.g., at certain temperature, in aqueous solution and/or for certain periods of time, has an increased retention of activity relative to a reference 5 enzyme. For example, for a thermostable luciferase, a reference luciferase may be native wild-type luciferase or recombinant wild-type luciferase. Preferably, for beetle luciferases, the activity is luminescence under conditions of saturation with luciferin and ATP. One measure of thermostability of an enzyme is the half-life of the enzyme in an aqueous solution (the time over which 50% of the 10 activity is lost) at a stated temperature.

The invention further encompasses expression vectors and other genetic constructs containing the mutant luciferases, as well as hosts, bacterial and otherwise, transformed to express the mutant luciferases. The invention is also drawn to compositions and kits which contain the novel luciferases, and use of 15 these luciferases in any methodology where luciferases are employed.

Various means of random mutagenesis were applied to a luciferase gene (nucleotide sequence), most particularly gene synthesis using an error-prone polymerase, to create libraries of modified luciferase genes. This library was expressed in colonies of *E. coli* and visually screened for efficient luminescence 20 to select a subset library of modified luciferases. Lysates of these *E. coli* strains were then made, and quantitatively measured for luciferase activity and thermostability. From this, a smaller subset of modified luciferases was chosen, and the selected mutations were combined to make composite modified luciferases. New libraries were made from the composite modified luciferases 25 by random mutagenesis and the process was repeated. The luciferases with the best overall performance were selected after several cycles of this process.

Methods of producing improved luciferases include directed evolution using a polynucleotide sequence encoding a first beetle luciferase as a starting (parent) sequence, to produce a polynucleotide sequence encoding a second 30 luciferase with increased thermostability, compared to the first luciferase, while maintaining other characteristics of the enzymes. A cDNA designated *lucPpe2* encodes a firefly luciferase derived from *Photuris pennsylvanica* that displays increased thermostability as compared to the widely utilized luciferase

designated *LucPpy* from *Photinus pyralis*. The cDNA encoding *LucPpe2* was isolated, sequenced and cloned (see Leach et al., 1997). A mutant of this gene encodes a first luciferase *LucPpe2* [T249M]. However, the methods of the invention are not limited to use with a polynucleotide sequence encoding a beetle luciferase, i.e., the methods of the invention may be employed with a polynucleotide sequence encoding other enzymes.

In an embodiment of a mutant luciferase, the amino acid sequence is that of *LucPpe2* shown in Figure 45 with the exception that at residue 249 there is a M (designated T249M) rather than the T reported by Leach et al. The underlined residue (249) shows mutation from T to M. This enzyme produced approximately 5-fold more light *in vivo* when expressed in *E. coli*.

Diluted extracts of recombinant *E. coli* that expressed mutant luciferases made by the methods of the invention were simultaneously screened for a plurality of characteristics including light intensity, signal stability, substrate utilization (K_m), and thermostability. A fully automated robotic system was used to screen large numbers of mutants in each generation of the evolution. After several cycles of mutagenesis and screening, thereby creating mutant libraries of luciferases, an increased thermostability compared to *LucPpe2* [T249M] of about 35°C was achieved for clone Luc90-1B5 which also essentially maintained enzymatic activity (there was only negligible loss in activity of 5%) when kept in aqueous solution over 2 hours at 50°C, 5 hours at 65°C, or over 6 weeks at 22°C.

Mutant luciferases of the present invention display increased thermostability for at least 2 hours at 50°C, preferably at least 5 hours at 50°C, and in the range of at least 2 hours, preferably at least 24 hours, and more preferably at least 50 hours, at temperatures including 50°C, 60°C, and/or at temperatures up to 65°C. In particular, the present invention comprises thermostable mutant luciferases which, when solubilized in a suitable aqueous solution, have a thermostability greater than about 2 hours at about 50°C, more preferably greater than about 10 hours at 50°C, and more preferably still greater than 5 hours at 50°C. The present invention also comprises mutant luciferases which, when solubilized in a suitable aqueous solution, have a thermostability greater than about 2 hours, more preferably at least 5 hours, even more preferably greater than about 10 hours, and even more preferably still greater

than about 24 hours, at about 60°C. The present invention further comprises mutant luciferases which when solubilized in a suitable aqueous solution have a thermostability greater than about 3 months at about 22°C, and more preferably a thermostability of at least 6 months at 22°C. An embodiment of the invention is

5 a luciferase mutant having thermostability at 65°C, wherein a loss of activity of about 5-6% was found after 6 hours (equivalent to a half-life of 2 days). The half-lives of enzymes from the most stable clones of the present invention, extrapolated from data showing small relative changes, is greater than 2 days at 65°C (corresponding to 6% loss over 6 hours), and about 2 years at 22°C

10 (corresponding to 5% loss over 9 weeks).

In particular, the invention comprises luciferase enzymes with embodiments of amino acid sequences disclosed herein (e.g., mutant luciferases designated Luc49-7C6, Luc78-0B10; Luc90-1B5, Luc133-1B2, and Luc146-1H2, as well as all other beetle luciferases that have thermostability as measured

15 in half-lives of at least 2 hours at 50°C. The invention also comprises mutated polynucleotide sequences encoding luciferase enzymes containing any single mutation or any combination of mutations of the type which convert an amino acid of the reference beetle luciferase into a consensus amino acid. Conserved amino acids are defined as those that occur at a particular position in all

20 sequences in a given set of related enzymes. Consensus amino acids are defined as those that occur at a particular position in more than 50% of the sequences in a given set of enzymes. An example is the set of beetle luciferase sequences shown in Figure 19, excluding *LucPpe2*.

Nucleotide sequences encoding beetle luciferases are aligned in Figure

25 19. Eleven sequences found in nature in various genera, and species within genera, are aligned, including *lucPpe2*. There are at least three mutations present in each mutant luciferase that show increased thermostability. In general, mutations are not of a conserved amino acid residue. The mutations in the mutant luciferases are indicated in Figures 22-47 by underlining.

30 The invention also provides methods to prepare enzymes having one or more desired properties, e.g., resistance to inhibition by a substrate analog of the enzyme or enhanced enzymological properties. The method comprises selecting at least one isolated polynucleotide sequence encoding an enzyme with the

desired property, e.g., an enzymological property, from a first population of mutated polynucleotide sequences. The selected, isolated polynucleotide sequence is then mutated to yield a second population of mutated polynucleotide sequences. Preferably, a mixture of selected isolated polynucleotide sequences 5 are mutated to yield a second population of mutated polynucleotide sequences. The process may be repeated until a further polynucleotide sequence is obtained, e.g., selected and/or isolated, which further polynucleotide sequence encodes an enzyme which has at least one of the desired properties. As used herein, the terms "isolated and/or "purified" refer to *in vitro* isolation of a RNA, DNA or 10 polypeptide molecule from its natural cellular environment, and from association with other components of the cell, such as nucleic acid or polypeptide, e.g., so that it can be sequenced, replicated, and/or expressed.

Brief Description of the Drawings

Figure 1 is a graphical representation of thermostability at 37°C of 15 LucPpe2[T249M]; Luc39-5B10; and Luc49-7C6, normalized to t = 0 [the X-axis is time in minutes; the Y-axis is % remaining activity; and "t" is time].

Figure 2 is a graphical representation of the remaining activity of Luc49- 7C6 and Luc78-0B10 at 50°C normalized to a t = 0 reading [the X-axis is time in hours; the Y-axis is % remaining activity; and t is time].

20 Figure 3 is a graphical representation of the luminescence produced by Luc49-7C6 and Luc78-0B10 at 60°C normalized to t = 0 [the X-axis is time in hours; the Y-axis is % remaining activity; and t is time].

Figure 4 is a graphical representation of thermostability of luciferases, 25 LucPpe2[T249M]; Luc49-7C6; and Luc78-0B10 thermostability at 22°C [the X-axis is time in days; the Y-axis is normalized light units].

Figure 5 is a graphical representation of the observed log luminescence produced by (Y) Luc78-0B10 compared to log luminescence predicted by the regression equation $Y = 0.0043X + 10.91$; the half life of the enzyme is calculated as 144 hours (6 days) [the X-axis is time in hours; the Y-axis is log 30 luminescence].

Figure 6 is a graphical representation of the observed log luminescence produced by Luc78-0B10 at 60°C compared to the log luminescence calculated by the regression equation $Y = 0.154X + 10.86$; the half life of the enzyme is

calculated as 38 hours (1.58 days) [the X-axis is time in hours; the Y-axis is log luminescence].

Figure 7 is a graphical representation of the observed log luminescence produced by Luc49-7C6 at 50°C compared to log luminescence predicted by the 5 regression equation $Y = -0.0059X + 8.757$; the half-life of the enzyme is calculated as 100.5 hours (4.2 days) [the X-axis is time in hours; the Y-axis is log luminescence].

Figure 8 is a graphical representation of the observed log luminescence produced by Luc49-7C6 at 60°C compared to the log luminescence calculated by 10 the regression equation $Y = -0.169X + 8.647$; the calculated half-life of the enzymes is 2.9 hours (the X-axis is time in hours; the Y-axis is log luminescence).

Figure 9 is a graphical representation of the observed log luminescence produced by Luc78-0B10 at 22°C compared to a predicted log luminescence, the 15 half-life of the enzyme is 109 days [the X-axis is time in days; the Y-axis is log luminescence].

Figure 10 is a graphical representation of the observed luciferase log luminescence produced by Luc49-7C6 at 22°C compared to a predicted log luminescence; the half-life of the enzyme is 64 days [the X-axis is time in days; 20 the Y-axis is log luminescence].

Figure 11 is a graphical representation of the observed log luminescence produced by luciferase Luc49-7C6 at 37°C compared to predicted log luminescence [the X-axis is time in minutes; the Y-axis is log luminescence].

Figure 12 is a graphical representation of the observed log luminescence produced by luciferase LucPpe2 [T249M] at 22°C compared to predicted log 25 luminescence [the X-axis is time in days; the Y-axis is log luminescence].

Figure 13 is a graphical representation of the observed log luminescence produced by luciferase LucPpe2 [T249M] at 37°C compared to predicted log luminescence [X-axis is time in minutes; Y-axis is log luminescence].

30 Figure 14 is a flow chart showing steps for an assay of *in vivo* and *in vitro* luciferase luminescence (Li); enzyme stability (τ); assay kinetics (S); and substrate binding (K_m).

Figure 15 is a schematic representation of a table top layout robot.

Figure 16A is a graphical representation of luciferase mutant Luc90-1B5 luminescence measured at 65°C, pH 6.5 (the X-axis is time in hours; the Y-axis is % luminescence).

Figure 16B is a graphical representation of luciferase mutant Luc90-1B5 5 luminescence at 22°C, pH 6.5 (the X-axis is time in days; the Y-axis is % luminescence).

Figure 17 is a diagram showing the evolutionary relationships among beetle luciferases based on amino acid sequences.

Figure 18A is a representation of the secondary structures of beetle 10 luciferase enzymes (helices are symbolized by cylinders, sheets by collections of arrows, loops connect helices with sheets).

Figure 18B shows the amino acids (tertiary structures) of the *LucPpe2* luciferase, wherein small spirals correspond to cylinders of Figure 18A.

Figure 18C shows that the general beetle architecture matches (is 15 superimposed on) that of Luc90-1B5.

Figure 19A presents alignment of the amino acid sequence (SEQ ID Nos:27-37) for luciferases from various beetle species (Lcr, Lla, Lmi, Pmi, Ppy, Lno, Ppe1, Phg, GR, YG, Ppe2, respectively) and luciferases of the present invention (Luc49-7C6; Luc78-0B10; Luc90-1B5, Luc133-1B2; and Luc146-20 1H2, SEQ ID Nos. 14, 19, 24, 44, and 45, respectively); the sequences are aligned, spaces where sequences cannot be aligned are shown by dots (e.g., . . .); only amino acids that differ in the luciferases of the present invention from those of some beetle species are shown, not the full sequences. “Cons” is a sequence showing conserved amino acids by single letters, and indicates non-conserved 25 amino acids by “-”.

Figure 19B presents alignment of the amino acid sequence (SEQ ID Nos:27-37) for luciferases from various beetle species (Lcr, Lla, Lmi, Pmi, Ppy, Lno, Ppe1, Phg, GR, YG, Ppe2) and luciferases of the present invention (Luc30-4B02 and Luc81-6G01, SEQ ID Nos. 47 and 26, respectively); the sequences are 30 aligned, spaces where sequences cannot be aligned are shown by dots (e.g., . . .); amino acids that differ in the luciferases of the present invention from those of some beetle species are shown in bold.

Figure 19C presents alignment of the amino acid sequence (SEQ ID NOs:27-34 and 36-37) for luciferases from various beetle species (Lcr, Lla, Lmi, Pmi, Ppy, Lno, Ppc1, Phg, YG, Ppe2, Ppl); the sequences are aligned, spaces where sequences cannot be aligned are shown by dots (e.g., . . .); in the line 5 beneath YG, X indicates positions in YG where mutations could yield a consensus amino acid; O indicates positions in YG where mutations could not yield a consensus amino acid.

Figure 20 is the 7216 bp *Ppe2* vector map in a pRAM backbone.

Figure 21 is a bar graph comparing luminescence as expressed in 10 recombinant colonies of *E. coli*; the colonies differ in the identity of the luciferase encoding vector (Luc+; Luc90-1B5; Luc78-1B10; Luc49-7C6; Luc*Ppe2* [T249M] and Luc*Ppe2*); in the recombinant colony shown in the Y-axis [the X-axis is normalized light units].

Figure 22 is a nucleotide (DNA) sequence (SEQ ID NO:1) encoding 15 mutant luciferase enzyme Luc49-7C6; mutations are indicated by underlining.

Figure 23 is a nucleotide (DNA) sequence (SEQ ID NO:2) encoding mutant luciferase enzyme Luc49-6C10, mutations are indicated by underlining.

Figure 24 is a nucleotide (DNA) sequence (SEQ ID NO:3) encoding a mutant luciferase enzyme Luc49-0G12; mutations are indicated by underlining.

20 Figure 25 is a nucleotide (DNA) sequence (SEQ ID NO:4) encoding a mutant luciferase enzyme Luc49-7A5; mutations are indicated by underlining.

Figure 26 is a nucleotide (DNA) sequence (SEQ ID NO:5) encoding a mutant luciferase enzyme Luc49-4G11; mutations are indicated by underlining.

25 Figure 27 is an amino acid sequence (SEQ ID NO:14) of the mutant luciferase designated Luc49-7C6; mutations are indicated by underlining.

Figure 28 is an amino acid sequence (SEQ ID NO:15) of mutant luciferase enzyme Luc49-6C10; mutations are indicated by underlining.

Figure 29 is an amino acid sequence (SEQ ID NO:16) of mutant luciferase enzyme Luc49-0G12; mutations are indicated by underlining.

30 Figure 30 is an amino acid sequence (SEQ ID NO:17) of mutant luciferase enzyme Luc49-7A5; mutations are indicated by underlining.

Figure 31 is an amino acid sequence (SEQ ID NO:18) of mutant luciferase enzyme Luc49-4G11; mutations are indicated by underlining.

Figure 32 is a nucleotide (DNA) sequence (SEQ ID NO:6) encoding mutant luciferase enzyme Luc78-0B10; mutations are indicated by underlining.

Figure 33 is a nucleotide (DNA) sequence (SEQ ID NO:7) encoding mutant luciferase enzyme Luc78-0G8; mutations are indicated by underlining;

5 X's signify unknown identities of nucleotides at certain positions.

Figure 34 is a nucleotide (DNA) sequence (SEQ ID NO:8) encoding mutant luciferase enzyme Luc78-1E1; mutations are by underlining; X's signify that the identity of a nucleotide at a position is unknown.

Figure 35 is a nucleotide (DNA) sequence (SEQ ID NO:9) encoding a 10 mutant luciferase Luc78-2B4; underlined nucleotides are mutations; X's signify unknown identities of nucleotides at certain positions.

Figure 36 is an amino acid sequence (SEQ ID NO:19) of the mutant luciferase Luc78-0B10; underlined amino acids are mutations.

Figure 37 is an amino acid sequence (SEQ ID NO:20) of the mutant 15 luciferase enzyme Luc78-0G8; underlined amino acids are mutations; X's signify unknown amino acids at a position.

Figure 38 is an amino acid sequence (SEQ ID NO:21) for mutant luciferase enzyme Luc78-1E1; underlined amino acids are mutations; X's signify an unknown amino acid at a position.

20 Figure 39 is an amino acid sequence (SEQ ID NO:22) for mutant luciferase enzyme Luc78-2B4; underlined amino acids are mutations; X's signify an unknown amino acid at a position.

Figure 40 is a nucleotide (DNA) sequence (SEQ ID NO:10) for encoding a mutant luciferase enzyme Luc85-4F12; underlined nucleotides are mutations; 25 X's signify an unknown amino acid at that position.

Figure 41 is an amino acid listing (SEQ ID NO:23) for a mutant luciferase enzyme Luc85-4F12; underlined amino acids are mutations; X's signify an unknown amino acid at that position.

Figure 42 is a nucleotide (DNA) sequence (SEQ ID NO:11) encoding 30 mutant luciferase enzyme Luc90-1B5; underlined nucleotides are mutations.

Figure 43 is an amino acid sequence (SEQ ID NO:24) for the mutant luciferase designated Luc90-1B5; underlined amino acids are mutated positions.

Figure 44 is a nucleotide (DNA) sequence (SEQ ID NO:12) encoding luciferase enzyme *LucPpe2* [T249M].

Figure 45 is an amino acid sequence (SEQ ID NO:25) for *LucPpe2* [T249M]; the underlined amino acid is a mutation from Thr to Met at residue 5 249.

Figure 46 is an amino acid sequence (SEQ ID NO:26) for luciferase enzyme *LucPp181-6G1*; underlined amino acids are mutations from a starting sequence; X shows ambiguity.

Figure 47 is a nucleotide (DNA) sequence (SEQ ID NO:13) encoding 10 luciferase enzyme *Luc81-6G1*; underlined nucleotides are mutations.

Figure 48 is a graphical representation of mutant luciferases *Luc49-7C6* and *Luc78-0B10* luminescence at 60°C normalized to t = 0 [the X-axis is time in hours, the Y-axis is log normalized luminescence].

Figure 49 is a graphical representation of luciferases *LucPpe2* [T249M], 15 *Luc49-7C6*, and *Luc78-0B10*, thermostability at 4°C, normalized to initial values [the X-axis is time in days; Y is log normalized light units].

Figure 50 is a graphical representation of mutant luciferases *Luc49-7C6* and *Luc78-0B10* luminescence at 50°C normalized to t = 0 [the X-axis is time in hours; the Y-axis is log luminescence].

20 Figure 51 is a graphical representation of mutant luciferases *Luc49-7C6* and *Luc78-0B10* luminescence at 50°C normalized at t = 0.

Figure 52 is a graphical representation of mutant luciferases *Luc49-7C6* and *Luc78-0B10* luminescence at 60°C normalized to t = 0 [the X-axis is time in hours; the Y-axis is luminescence].

25 Figure 53 is a graphical representation of luciferases *LucPpe2* [T249M], *Luc49-7C6*, and *Luc78-0B10* thermostability at 22°C [the X-axis is time in days; the Y-axis is log luminescence].

Figure 54A is a graphical representation of luminescence of *Luc90-1B5*; *Luc133-1B2*; and *Luc146-1H2*, at pH 4.5 and 48°C, normalized to t = 0.

30 Figure 54B is a graphical representation of the half-life of *Luc90-1B5*; *Luc133-1B2*; and *Luc146-1H2*, at pH 4.5 and 48°C. The half-life of *Luc90-1B5* under these conditions is about 3 minutes, *Luc133-1B2* about 20 minutes, and *Luc146-1H2* about 62 minutes.

Figure 55 is a nucleotide (DNA) sequence (SEQ ID NO:42) encoding a luciferase enzyme Luc133-1B2; mutations are indicated by underlining.

Figure 56 is a nucleotide (DNA) sequence (SEQ ID NO:43) encoding a luciferase enzyme Luc146-1H2; mutations are indicated by underlining.

5 Figure 57 is an amino acid sequence (SEQ ID NO:44) of mutant luciferase Luc133-1B2; mutations are indicated by underlining.

Figure 58 is an amino acid sequence (SEQ ID NO:45) of mutant luciferase Luc146-1H2; mutations are indicated by underlining.

10 Figure 59 is a graphical representation of the signal kinetics of clones Luc49-7C6; Luc78-0B10; Luc90-1B5; Luc133-1B2; and Luc146-1H2 at pH 7.8 at room temperature.

Figure 60 is a graphical representation of the normalized luminescence at 50°C pH 7.8 of Luc49-7C6; Luc78-0B10; Luc90-1B5; Luc133-1B2; and Luc146-1H2; from t = 0 to about 8 hours.

15 Figure 61 is a graphical representation of the resistance of selected luciferases to a substrate inhibitor. The data is presented as the log of the luminescence versus time for Luc90-1B5; Luc133-1B5; and Luc146-1H2.

Figure 62 is a graphical representation of the log of luminescence over time at 22°C, pH 6.5 for Luc90-1B5 and LucPpe2[T249M].

20 Figure 63 is a graphical representation of thermostability of selected mutant luciferases and LucPpIYG at room temperature in aqueous solution containing 1% Triton X-100.

Figures 64 is a graphical representations of the sustained luminescence activity (expressed as luminescence/O.D.) over time for certain luciferases.

25 Figure 65 is a nucleotide (DNA) sequence (SEQ ID NO:46) encoding a luciferase enzyme Luc81-0B11; mutations are indicated by underlining.

Figure 66 is an amino acid sequence of mutant luciferase Luc81-0B11; mutations are indicated by underlining.

Detailed Description of the Invention

The invention relates to enzymes, e.g., beetle luciferases, that are created by mutations made in the encoding genes, generally by recursive mutagenesis, which mutated enzymes have one or more desired properties, for example,

5 increases thermostability, increased resistance to inhibitors, and/or enhanced enzymological properties, relative to a reference enzyme, e.g., the wild-type enzyme. The polynucleotide sequence which encodes an enzyme of the invention comprises mutations that encode a plurality of amino acid substitutions relative to the polynucleotide sequence encoding the enzyme from which the

10 enzyme of the invention was derived. For example, the invention relates to enzymes, e.g., luciferases, that are thermostable. The increased thermostability allows storage of enzymes such as luciferases without altering its activity, and improves reproducibility and accuracy of assays using the mutated luciferases. Thus, one embodiment of the invention comprises isolated polynucleotide

15 sequences (cDNAs) which encode mutant luciferases with increased thermostability, vectors containing the polynucleotide sequences, and hosts transformed to express the polynucleotide sequences. Table 1 shows results of about 250 clones and characteristics of the luciferases from the clones including thermostability. The invention also encompasses the use of the mutant

20 luciferases in any application where luciferases are conventionally utilized, and kits useful for some of the applications.

Unexpectedly, beetle luciferases with the sought after improved thermostability were achieved in the present invention through a process of recursive mutagenesis and selection (sometimes referred to as "directed evolution"). A strategy of recursive mutagenesis and selection is an aspect of the present invention, in particular the use of multi-parameter automated screens. Thus, instead of screening for only a single attribute such as thermostability, simultaneous screening was done for additional characteristics of enzyme activity and efficiency. By this method, one property is less likely to "evolve" at 25 the expense of another, resulting in increased thermostability, but decreased activity, for example.

Table 1 presents examples of parameter values (Li, Tau, K_m and S, see below) derived from experiments using different luciferases as starting (parent)

sequences. The subtitles refer to designations of the temperature at which the enzyme stability was measured and the starting luciferase, e.g., Luc39-5B10 at 51°C and so forth. All parameters in each experiment are recorded as relative values to the respective starting sequence, e.g., the parameter values for the 5 starting sequence in any experiment equals “1.” (See Example 2 herein for definitions.)

Thermostability has evolved in nature for various enzymes, as evidenced by thermostable isozymes found in thermophilic bacteria. Natural evolution works by a process of random mutagenesis (base substitutions, gene deletions, 10 gene insertions), followed by selection of those mutants with improved characteristics. The process is recursive over time. Although the existence of thermostable enzymes in nature suggests that thermostability can be achieved through mutagenesis on an evolutionary scale, the feasibility of achieving a given level of thermostability for a particular class of enzymes by using short 15 term laboratory methods was unpredictable. The natural process of evolution, which generally involves extremely large populations and many millions of generations and genes, by mutation and selection cannot be used to predict the capabilities of a modern laboratory to produce improved genes by directed evolution until such mutants are produced.

20 After such success, because the overall three-dimensional structure of all beetle luciferases are quite similar, having shown it possible for one member of this class makes it predictable that high thermostability can be achieved for other beetle luciferases by similar methods. Figure 17 shows an evolutionary relationship among beetles luciferases, all of which have a similar overall 25 architecture. The structural class to which the beetle luciferases belong is determined by the secondary structure (e.g. helices are symbolized by cylinders, sheets by collections of arrows, loops connect helices with sheets (Figure 18A). Figure 18B shows the amino acids of the *LucPpe2* luciferase wherein small spirals correspond to cylinders of Figure 18A; Figure 18C shows that the general 30 beetle architecture matches (is superimposed on) that of *LucPpe2*. This is support for the expectation that the methods of the present invention can be generalized to all beetles luciferases.

Enzymes belong to different structural classes based on the three-dimensional arrangement of secondary elements such as helices, sheets, and loops. Thermostability is determined by how efficiently the secondary elements are packed together into a three-dimensional structure. For each structural class, 5 there also exists a theoretical limit for thermostability. All beetle luciferases belong to a common structural class as evident by their common ancestry (Figure 17), homologous amino acid sequences, and common catalytic mechanisms.

The application of a limited number of amino acid substitutions by mutagenesis is unlikely to significantly affect the overall three-dimensional 10 architecture (i.e., the structural class for mutant luciferases is not expected to change.) Because the theoretical limit for thermostability for any structural class is not known, the potential thermostability of beetle luciferases was not known until demonstrations of the present invention.

A priori difficulties in achieving the goals of the present invention 15 included: 1. The types of mutations which can be made by laboratory methods are limited.

- i) By random point mutation (e.g. by error-prone PCR), more than one base change per codon is rare. Thus, most potential amino acid changes are rare.
- 20 ii) Other types of random genetic changes are difficult to achieve for areas greater than 100 bp (e.g., random gene deletions or insertions).
- 2. The number of possible luciferase mutants that can be screened is limited.
- 25 i) Based on sequence comparisons of natural luciferases, ignoring deletions and insertions, more than 10^{189} functional enzyme sequences may be possible.
- ii) If 100,000 clones could be screened per day, it would require more than 10^{179} centuries to screen all possible mutants assuming same mutant was never screened twice (actual screening rate for 30 the present invention was less than 5000 per day).

3. The probability of finding functional improvement requiring cooperative mutations is rare (the probability of finding a specific cooperative pair is 1 out of 10⁸ clones).

5 Thus, even if the theoretical limits of thermostability were known, because only a very small number of the possible luciferase mutants can be screened, the *a priori* probability of finding such a thermostable enzyme was low.

However, the present invention now shows that it is possible and feasible to create novel beetle luciferases having high thermostability.

10 a) The approximately 250 mutants produced by methods of the present invention wherein the initial sequence was from *lucPpe2* or *lucPplYG* demonstrate that it is possible and feasible for at least one member of this enzyme class to achieve high thermostability.

15 b) Any beetle luciferase can be improved by similar means because the luciferases belong to the same structural class.

i) Because all beetle luciferases belong to the same structural class, they also share in the same pool of potentially stabilizing mutations (this conclusion is supported by observation that a high percentage of the stabilizing mutations found in the clones of the present invention were conversions to “consensus amino acids” in other beetle luciferases that is, amino acids that appear in the majority of beetle luciferase sequences (see Figure 19).

20 ii) Similar results were achieved using beetle luciferase, consisting largely of a different amino acid sequence, from the luminous beetle *Pyrophorus plagiophthalmus* (*LucPplYG*). The wild-type *lucPplYG* has 48% nucleotide sequence identity to the wild type *lucPpe2*.

25 The *LucPplYG* mutants were subjected to fewer cycles of directed evolution than the *LucPpe2* mutants described herein. Also, in some instances, mutants were selected with less emphasis placed on their relative thermostability.

30

The most stable clone resulting from this evolution (Luc80-5E5) has a half-life of roughly 3.8 hours at 50°C in solution.

To compensate for a statistical effect caused by the large number of deleterious random mutations expected relative to the beneficial mutations, methods were employed to maximize assay precision and to re-screen previously selected mutations in new permutations. Among the methods for maximizing assay precision were closely controlling culture conditions by using specialized media, reducing growth rates, controlling heat transfer, and analyzing parameters from mid-logarithmic phase growth of the culture. The robotic processes maximized for precision include controlling mixing, heat transfers, and evaporation of samples in the robotic screening process; and normalizing data to spatially distributed control samples. New permutations of the selected mutations were created by a method of DNA shuffling using proof-reading polymerases.

The difficulty in predicting the outcome of the recursive process is exemplified by the variable success with the other characteristics of luciferase that were also selected for. Although the primary focus was on the enzyme thermostability, selection for mutants producing brighter luminescence, with more efficient substrate utilization, and an extended luminescence signal was also attempted. The definitions are given by equations herewith. The selection process was determined by changes relative to the parent clones for each iteration of the recursive process. The amount of the change was whatever was observed during the screening process. The expression of luciferase in *E. coli* was relatively inefficient, for *LucPpe2*, compared to *Luc +*. Other luciferases varied (see Figure 21).

To improve the overall efficiency of substrate utilization, reduction in the composite apparent utilization constant (i.e., $K_m[\text{ATP} + \text{luciferin}]$) for both luciferin and ATP was sought. Although there was an unexpected systematic change in each utilization constant ($K_m[\text{ATP}]$, $K_m[\text{luciferin}]$), there was little overall change. Finally, the luminescence signal could only be moderately affected without substantially reducing enzyme efficiency. Thus, although the

enzyme thermostability was greatly increased by methods of the present invention, other characteristics of the enzyme were much less affected.

Figures 1-13, 16, 48-53, 60 and 62 present measurements of thermostability of mutant luciferases. Figures 48-53 present other results of the 5 mutant luciferases. Compositions of the invention include luciferases having greater than the natural level of thermostability. Each mutant luciferase is novel, because its individual characteristics have not been reported. Specific luciferases are known by both their protein and gene sequences. Many other luciferases were isolated that have increased, high thermostability, but whose sequences are 10 not known. These luciferases were identified during the directed evolution process, and were recognized as distinct by their enzymological characteristics. The mutant luciferases of the present invention, e.g., Luc90-1B5, can display remarkable and heretofore unrealized thermostability at temperatures ranging from 22°C to at least as high as 60°C.

15 Other aspects of the invention include methods that incorporate the thermostable luciferases, specifically beetle luciferases having high thermostability, as well as methods to prepare an enzyme, including a luciferase, having one or more desired properties, e.g., resistance to inhibition by a substrate inhibitor, or enhanced enzymological properties. Thus, the invention also 20 provides a method to prepare an enzyme which has at least one enhanced enzymological property. From a population of polynucleotide sequences encoding the enzyme which is derived from a first polynucleotide sequence encoding the enzyme which is subject to mutation, at least one polynucleotide sequence encoding an enzyme which has the enhanced enzymological activity is 25 selected and isolated. In one embodiment, oligonucleotide-mediated mutagenesis is then employed to introduce at least one codon which encodes a consensus amino acid to at least one of the selected, isolated polynucleotide sequences encoding the enzymes to yield a further polynucleotide sequence encoding the enzyme and having the codon which encodes the consensus amino 30 acid, wherein the codon which is introduced is not present in the first polynucleotide sequence.

Production of Luciferases of the Present Invention

The method of making luciferases with increased thermostability is recursive mutagenesis followed by selection. Embodiments of the highly thermostable mutant luciferases of the invention were generated by a reiterative 5 process of random point mutations beginning with a source nucleotide sequence, e.g., the *lucPpe2* [T249M] cDNA. Recombination mutagenesis is a part of the mutagenesis process, along with point mutagenesis. Both recombination mutagenesis and point mutagenesis are performed recursively. Because the mutation process causes recombination of individual mutants in a fashion similar 10 to the recombination of genetic elements during sexual reproduction, the process is sometimes referred to as the sexual polymerase chain reaction (sPCR). See, for instance, Stemmer, U.S. Patent No. 5,605,793, issued February 25, 1997.

Taking the *lucPpe2* cDNA sequence as a starting point, the gene was mutated to yield mutant luciferases which are far more thermostable. A single 15 point mutation to the *lucPpe2* sequence yielded the luciferase whose sequence is depicted as T249M. This mutant is approximately 5 times brighter *in vivo* than that of *lucPpe2*, it was utilized as a template for further mutation. It was also used a baseline for measuring the thermostability of the other mutant luciferases described herein.

20 Embodiments Of Sequences Of Luciferases Of The Present Invention

Figure 45 shows the amino acid sequence of the *LucPpe2* luciferase (T249M). The sequence contains a single mutation at position 249 from T to M (underlined) which distinguishes it from the sequence reported by Leach et al. (1997). This luciferase has a spectral maximum of 552 nm, which is yellow 25 shifted from that of the luciferase of Leach et al. This mutant was selected for use as an original template in some of the Examples because it is approximately 5 times brighter *in vivo*, than the form reported by Leach et al. which allowed for more efficient screening by the assay. These sequences show changes from the starting sequence (T249M) by underlining. Note that "x" in the sequence 30 denotes an ambiguity in the sequence.

Directed Evolution, A Recursive Process

Directed evolution is a recursive process of creating diversity through mutagenesis and screening for desired changes. For enzymological properties that result from the cumulative action of multiple amino acids, directed evolution 5 provides a means to alter these properties. Each step of the process typically produces small changes in enzyme function, but the cumulative effect of many rounds of this process can lead to substantial overall change.

The characteristic, "thermostability" is a candidate for directed evolution because it is determined by the combined action of many of the amino acids 10 making up the enzyme structure. Luminescence output and efficiency of substrate binding of the modified luciferase were also screened. This was to ensure that changes in thermostability did not also produce undesirable changes in other important enzymological properties.

Because the frequency of deleterious mutations is much greater than 15 useful mutations, it is likely that undesirable clones are selected in each screen within the precision limits of the present invention. To compensate for this, the screening strategy incorporated multiple re-screens of the initially selected mutations. However, before re-screening, the selected mutations were "shuffled" to create a library of random intragenetic recombinations. This process allows 20 beneficial mutations among different clones to be recombined together into fewer common coding sequences, and unlinks deleterious mutations to be segregated and omitted. Thus, although essentially the same set of selected mutations was screened again, they were screened under different permutations as a result of the recombination or shuffling.

25 Although results of each step of the evolutionary process were assayed by quantitative measurements, these measurements were mutually made in cell lysates rather than in purified enzymes. Furthermore, each step only measured changes in enzyme performance relative to the prior step, so global changes in enzyme function were difficult to judge.

30 Table 1 summarizes the characteristics of various clones obtained using the methods of the invention.

Table 1

Control is Luc39-5B10 at 51°C

Experiment	Clone ID	Luminescence (Li)	Enzyme stability (tau)	Substrate binding (Km)	Signal stability (S)	
5	40	0a7	1.04	4.5	0.78	
	40	5h4	1.29	1.61	1.16	
	40	0c2	1.13	1.54	0.91	
	40	5g4	1	1.4	0.85	
	40	6d3	1.02	1.37	0.79	
	40	1g4	1.06	1.28	0.77	
	40	1d4	1.69	1.23	0.73	
	40	0h9	1.26	1.21	0.63	
	40	2f6	3	1.07	0.49	
	40	7d6	3.09	1.058	1.09	
10	40	5a7	4.3	1.025	0.93	
	40	4c8	1	1	0.33	
					1.004	
15	Experiment	Clone ID	Li	tau	Km	S
	40	7h7	0.73	2.4	2.1	0.995
	40	5a5	0.77	1.93	2.7	1.002
	40	2c12	1.06	1.7	0.91	1.003
	40	6e5-	1.16	1.62	1.53	0.997
	40	4e5-	1.08	1.37	1.4	1.004
	40	6g7	1.3	1.27	1.39	0.999
	40	1h4	1.36	1.24	0.56	0.994
	40	0c11	4.1	1.23	1.24	0.996
	40	2h9	5.3	1.01	0.83	0.986
20	42	6b10	0.97	3.6	0.97	0.997
	42	1c3	0.91	2.1	0.6	0.998
	42	7h9	0.8	1.8	0.8	0.982
	42	6b2	0.77	1.72	0.8	0.978
	42	6d6	0.83	1.7	0.733	0.975
	42	4e10-	0.77	1.63	1.8	0.954
25	42	1b5	0.83	1.41	1.05	0.955
	42	6e6-	0.71	1.16	0.89	0.955
30	42					
	42					
	42					
	42					
	42					
35	42					
	42					
	42					

Experiment	Clone ID	Li	tau	Km	S
20	41	7h7	0.73	2.4	2.1
	41	5a5	0.77	1.93	2.7
	41	2c12	1.06	1.7	0.91
	41	6e5-	1.16	1.62	1.53
	41	4e5-	1.08	1.37	1.4
	41	6g7	1.3	1.27	1.39
	41	1h4	1.36	1.24	0.56
	41	0c11	4.1	1.23	1.24
	41	2h9	5.3	1.01	0.83
	42	6b10	0.97	3.6	0.97
25	42	1c3	0.91	2.1	0.6
	42	7h9	0.8	1.8	0.8
	42	6b2	0.77	1.72	0.8
	42	6d6	0.83	1.7	0.733
	42	4e10-	0.77	1.63	1.8
	42	1b5	0.83	1.41	1.05
30	42	6e6-	0.71	1.16	0.89
	42				
	42				
	42				
	42				
35	42				
	42				
	42				

42	3a9	0.85	1.3	0.86	0.997
42	6b6	2.7	1.3	0.91	1.02
42	6e9-	1.5	1.27	0.98	1.01
42	3h11	1.73	1.21	0.63	0.985
42	1a2	1.11	1.17	0.77	1.005
42	3f7	0.49	1.16	1.13	0.944
42	1a4	2	1.01	0.76	0.996

10

Control is Luc40-0A7 at 54°C

5

Experiment	Clone ID	Li	tau	Km	S
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46	2h3	0.86	6.4	0.37	0.96
46	4a9	0.67	5.7	0.66	0.997
46	2g4	0.65	5.3	0.78	0.96
46	5d12	0.94	4.9	0.94	1.002
46	1h11	1.02	4.8	0.84	0.998
46	5a10	1.23	4.4	0.81	0.9842
46	0a8	1.35	4.3	0.89	1
46	4d3	0.51	3.6	0.65	0.975
46	2a3	1.17	2.9	0.57	0.988
46	3b11	1.39	2.5	0.63	1.02
46	7g12	1.49	2.5	0.91	1.02
46	0g9	1.86	2.25	0.5	0.998
46	7h8	1.07	1.36	0.52	0.99
46	1g8	0.3	1.31	0.72	0.92
46	1d3	1.74	1.13	1.02	1.001
46	0c3	1.68	1.01	0.74	1.01
46	5c11	0.82	1.01	0.6	0.95

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Control is Luc46-2H3 at 54°C

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Experiment	Clone ID	Li	tau	Km	S
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49	6c10	0.57	2.2	0.98	1
49	7c6	1.12	1.9	0.93	1.01
49	0g12	1	1.58	0.69	1.08
49	7a5	1.08	1.44	1.1	0.99

49	1f6	0.66	1.13	1.04	1.006
49	0b5	0.76	1.07	1.03	0.98
49	4a3	0.94	1.06	0.77	1

5

Control is Luc49-7C6 at 56°C

Experiment	Clone ID	Li	tau	Km	S
56	2d12	0.97	2.9	0.29	1.006
56	5g10	1.01	2.77	0.64	1.007
56	3d5	1.32	2.25	1.85	1.0

Experiment	Clone ID	Li	tau	Km	S
57	3d1	1.06	2.9	1.05	1.02
57	6g12	1	2.7	0.87	1.004
57	4c1	0.79	2.6	0.93	1.014
57	5f10	0.72	1.9	0.64	1.03
57	1e6-	0.84	1.49	0.984	0.9871
57	1h2	0.94	1.43	0.68	0.991
57	2a6	1.08	1.08	0.89	0.9976

Experiment	Clone ID	Li	tau	Km	S
58	1g6	1.57	8.9	1.78	1.02
58	0a5	1.53	8.5	1.56	1.05
58	1b1	0.84	8.5	0.6	1.04
58	3g1	1	7.34	0.62	1.006
58	0f3	1.31	6.9	0.57	0.98
58	3e12-	1.06	6.3	0.47	0.996
58	0c7	1.9	4	0.64	1.06
58	0d1	1.03	3.76	0.49	1.03
58	3c7	1.49	3.4	0.55	1.04
58	2a2	1.4	2.2	0.5	1.05
58	2a8	3.2	2	0.81	1.05
58	0f2	2.2	1.92	0.45	1.04

58	1b4	5.1	1.87	1.08	1.09
58	2b3	2.7	1.55	0.57	1.04
58	4g1	4.9	1.2	0.72	1.06

5

Control is Luc58-0A5 at 58°C

	Experiment	Clone ID	Li	tau	Km	S
10	61	4e9-	1.03	1.84	0.76	1.01
	61	1f1	1.02	1.43	0.7	1
	61	2e12-	1.56	1.34	0.48	1.003
	61	2f2	1.5	1.3	0.32	1.01
	61	6b4	1.2	1.26	0.88	0.98
	61	4c10	1.46	1.12	1.06	0.99
15	61	4g11	1.31	1.03	1.43	1.03
	61	2f1	1.41	1.02	0.79	0.995
	61	2g1	1.3	1	1.17	1

	Experiment	Clone ID	Li	tau	Km	S
20	65	6g12	0.87	2.3	0.73	0.9605
	65	1h6	0.84	2.2	1.62	0.9598
	65	7f5	1.2	1.56	2.07	1.0087
	65	5g5	2.3	1.49	0.45	0.9985
	65	7h2	1.56	1.27	0.91	1.0658
	65	7b2	1.98	1.16	0.6	0.9289
25	65	0g9	1.36	1.09	1.46	0.9927
	65	6c7	1.48	1.06	0.86	0.9967
	65	1e12-	1.59	1.05	1.03	0.9582
	65	4e2-	1.21	1.05	1.11	0.943
	65	6a10	1.7	1.04	0.93	0.992
	65	4b9	1.48	1.04	1.61	1.0009
30	65	6c1	1.36	1.02	0.72	0.9978

35	Experiment	Clone ID	Li	tau	Km	S

68	2g6	1.39	3.9	1.17	0.9955
68	4g3	2	2.5	0.27	0.9927
68	5a3	1.04	1.64	0.65	0.8984
68	2b7	1.04	1.64	5.2	0.9237
68	5d10	2.75	1.36	0.73	1.0078
68	7d12	1.85	1.32	0.66	1.0084
68	7b9	1.8	1.19	0.56	1.0052
68	7b3	1.2	1.16	0.55	0.9951
68	1g10	1.48	1.05	1.22	1.0025

10

Experiment	Clone ID	Li	tau	Km	S
70	2a7	1.94	4.6	0.7	1.0015
70	3d6	3.5	4.2	0.18	1.03
70	4f8	1.87	4.2	0.69	0.9979
70	7h5	2.4	2.6	0.18	1
70	5h6	3.1	2.3	0.6	0.999
70	7d6	3	2.2	2.29	0.9989
70	5a3	3.1	1.5	0.18	1.0058
70	7d2	2.5	1.4	0.66	1.0126
70	3h7	3.2	1.22	0.23	1.002
70	0h5	2.5	1.15	0.36	0.9992
70	0d7	1.86	1	1.83	0.993
70	1g12	2.42	1	0.26	0.965

25

Experiment	Clone ID	Li	tau	Km	S
71	1d10	1.6	4.5	1.06	1.0065
71	6f11	1.8	4.3	0.98	0.953
71	7h4	3.4	3.6	0.56	1.0045
71	4h3	3.1	3.1	0.42	1.0171
71	1h5	1.31	3.01	1.31	0.9421
71	5e4-	5.4	2.3	0.35	0.994
71	5c1	2.2	2.3	0.89	0.9746
71	0h7	3.6	1.8	0.59	1.0197

5	71	6h9	23.7	1.71	0.91	1.0064
	71	7e3-	5.3	1.7	0.7	1.0028
	71	5d4	11.1	1.48	0.35	1.0213
	71	2e3-	4	1.47	0.45	0.9654
	71	6h11	17.7	1.15	2.8	1.0064
	71	2e10-	3	1.1	0.66	0.9588
	71	2g2	4.4	1.01	0.44	1.0046

10 Control is Luc71-5D4 at 60°C

	Experiment	Clone ID	Li	tau	Km	S
15	72	2g6	0.38	3.1	1.58	1.0052
	72	5f12	0.81	1.53	1.02	0.9678
	72	0d7	0.76	1.44	1.4	0.9838
	72	5c12	0.87	1.43	1.04	0.9718
	72	1e1-	1.04	1.41	1.15	0.9956
	72	5b12	0.83	1.41	1.02	0.9731
	72	0b7	1.11	1.04	0.91	1.0049
	72	3b4	0.49	1.03	2.2	0.9581

20

	Experiment	Clone ID	Li	tau	Km	S
25	73	2h8	0.85	1.9	1.08	1.0123
	73	4e6-	0.95	1.76	0.94	0.9939
	73	3g8	0.86	1.53	1.04	1
	73	1g3	1.7	1.14	0.97	0.9921

	Experiment	Clone ID	Li	tau	Km	S
30	74	2a9	0.96	1.77	0.86	0.999
	74	4e10-	0.8	1.36	1.33	0.09897
	74	0d5	1.69	1.28	0.61	0.9927
	74	6g7	1.75	1.07	1.33	1.0022
	74	5d8	0.46	1.06	0.95	0.899
	74	5e7-	1.22	1.05	0.87	0.9977

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74	6e1-	1.19	1.02	0.96	0.999
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	Experiment	Clone ID	Li	tau	Km	S
5	76	6c3	2.3	6.4	1.2	0.9865
	76	2a9	0.93	4.7	1.08	0.999
	76	3h9	1.26	2.6	1.02	0.9973
	76	0b10	1.52	2.4	1.4	0.992
	76	0h9	1.71	1.44	1.05	1.018
10	76	2e9-	0.44	1.15	1.2	0.9318
	76	0e10-	1.67	1.1	1.02	1.014
	76	0c10	1.13	1.05	1	0.9974
	76	3e8-	1.35	1.03	1.1	0.9894
	76	0d12	0.69	1	0.92	0.932
15	76	0f10	0.62	1	1.2	0.9478

	Experiment	Clone ID	Li	tau	Km	S
20	78	1e1-	0.54	8.9	1.15	0.9877
	78	0h7	1.4	5	0.97	1.014
	78	0a6	1	4.3	1.5	0.9967
	78	0b10	1.93	2	1	0.9926
	78	0f11	1.6	2	0.91	0.9905
25	78	3f1	2.4	1.7	1.09	0.9936
	78	2b4	1.97	1.36	0.98	1.0094
	78	5b3	3.2	1.19	1.03	0.9735
	78	2g12	2.5	1.03	1	1.0134
	78	0h2	1.6	1	1.15	1.0168

30

Control is Luc78-0B10 at 62°C

	Experiment	Clone ID	Li	tau	Km	S
35	82	2g12	0.9811	2.09	0.8851	0.9939
	82	4b9	1.0845	1.8419	0.8439	1.0078
	82	0d11	0.7622	1.5171	1.11	0.9998

82	3g1	0.8805	1.504	0.9629	0.9927
82	1d1	0.9741	1.4497	0.8936	0.9986
82	1e8-	0.8206	1.4433	0.9876	0.9968
82	0h9	1.1355	1.3626	0.9171	1.0094
5	82	2c6	1.0931	1.3402	0.9482
	82	3g9	1.0364	1.251	0.968
	82	4h8	0.8816	1.1667	0.9165
	82	0a10	1.0535	1.1128	1.0413
	82	4g1	1.4305	1.0862	1.1734
	82				1.0059

10

Experiment	Clone ID	Li	tau	Km	S
84(121)	6h7	0.3755	29.3639	2.3636	0.8905
84(121)	2h9	0.4264	28.7958	1.819	0.904
15	84(121)	3f7	0.4161	25.3058	1.8079
	84(121)	2h10	0.9667	14.4658	0.8073
	84(121)	3a2	0.3329	12.6	2.5444
	84(121)	3a6	1.2299	7.2384	0.7866
	84(121)	5b12	1.0535	6.0315	0.7824
20	84(121)	5a7	1.0413	4.9054	0.8864
	84(121)	3d2	0.2032	4.8	2.4623
	84(121)	2a9	1.0847	4.7486	0.7746
	84(121)	5e11-	1.1918	4.0988	0.872
25	84(121)	7h2	0.9115	3.9929	0.909
	84(121)	3b5	1.2014	3.8251	0.7509
	84(121)	1f8	1.07	3.06	0.8276
	84(121)	2e2-	1.4356	1.9315	0.7863
	84(121)				1.0175

30

Control is Luc84-3A6 at 64°C

Experiment	Clone ID	Li	tau	Km	S
85(86)	2a2	0.2266	12.9013	3.326	0.8705
85(86)	4f12	1.1167	4.7851	0.7439	1.0092
	85(86)	4e9-	1.0869	4.4953	0.8539
35	85(86)	1f11	0.6994	4.0976	0.842
	85(86)				1.0124

85(86)	5a4	1.2273	4.09	0.9683	1.0098
85(86)	3e10-	0.8902	3.5342	0.8106	1.0069
85(86)	3e12-	1.0512	3.4883	0.853	1.0054
85(86)	5e4-	0.9562	3.3886	1.0328	1.0069
5	85(86)	0e6-	0.1494	3.0145	3.6293
	85(86)	6b1	0.7615	2.5712	0.8695
	85(86)	6h7	1.0285	2.5401	0.8963
	85(86)	4b11	0.9816	2.3899	0.7927
10	85(86)	6d7	1.1087	2.0607	0.9042
	85(86)	2e10-	0.3028	2.0603	1.9649
	85(86)	2a9	1.448	1.1819	0.9722
					1.0046

Control is Luc85-4F12 at 65°C

15	Experiment	Clone ID	Li	tau	Km	S
	88	3c1	1.4439	2.0938	0.9874	0.9976
	88	6g1	1.0184	1.2665	1.2184	1.0019
	88	3e4-	1.331	1.0996	1.0669	0.9983

20	Experiment	Clone ID	Li	tau	Km	S
	89	1a4	1.2565	2.4796	1.0338	0.997
	89	3b1	0.7337	1.9976	0.9628	1.0001
	89	2b12	1.0505	1.8496	1.0069	1.0012
25	89	0b5	1.5671	1.1362	1.0912	0.9995
	89	1f1	1.378	1.1018	0.9804	0.996
	89	2f1	1.4637	1.0894	0.9189	0.9992

30	Experiment	Clone ID	Li	tau	Km	S
	90	0f1	1.4081	1.3632	1.027	0.9987
	90	1b5	1.4743	1.1154	1.0812	1.0011
	90	6g5	1.2756	1.0605	1.0462	1.0012
35	90	5e6-	1.0556	1.0569	1.1037	1.0011
	90	4e3-	1.2934	1.0291	1.0733	1.0002

To evaluate the impact of directed evolution on enzyme function, clones from the beginning, middle and end of the process (Table 2) were purified and analyzed. The clones selected for this analysis were Luc[T249M], Luc49-7C6, and Luc78-0B10. Another clone, Luc90-1B5, created by a subsequent strategy 5 of oligonucleotide-directed mutagenesis and screening was also purified for analysis.

Table 2: Thermostability Of Luciferase Activity At Different Temperatures (Half-Life In Hours)

10

	<i>Room Temperature*</i>	37°C	50°C	60°C
Luc[T249M]	110	0.59	0.01	
Luc49-7C6	430	68	31	6.3
Luc78-0B10	3000	220	47	15

15 * about 25°C

The effect of directed evolution on thermostability was dramatic. At high temperatures, where the parent clone was inactivated almost instantaneously, the mutant enzymes from the related clones showed thermostability over several 20 hours (see also Table 1, and Figures 1-3, 5-8, 11, 13, 50-52 and 60). Even at room temperature, these mutants are several fold more thermostable than the parent enzyme (see also Figures 4, 9-10, 12, 53, and 62). Subsequent analysis of Luc90-1B5 showed this enzyme to be even more thermostable, having a half-life of 27 hours at 65°C when tested under the same buffer conditions (Figure 16A). 25 With some optimization of buffer conditions, this enzyme showed very little activity loss at 65°C over several hours (citrate buffer at pH 6.5; Figure 16A). This luciferase was stable at 22°C over several weeks when incubated at pH 6.5 (Figure 16B). At times over 100 days at 4°C, the mutant enzymes had increased thermostability. At times of less than 15 days at 4°C, the thermostabilities of the 30 mutants Luc49-7C6 and Luc78-0B10 were not distinguishable from the parent enzyme (Figure 49).

Kajiyama and Nakano (1993) showed that a single amino acid substitution of A at position 217; to either I, L, or V, in the firefly luciferase from *Luciola*

lateralis, resulted in a luciferase having increased thermostability. Substitution with leucine produced a luciferase that maintained 70% of its activity after incubation for 1 hour at 50°C. All of the enzymes of the present invention created through directed evolution, are much more stable than this *L. lateralis* mutant. One clone, Luc90-1B5, maintains 75% activity after 120 hours (5 days) incubation under similar conditions (50°C, 25 mol/L citrate pH 6.5, 150 mmol/L NaCl, 1 mg/mL BSA, 0.1 mmol/L EDTA, 5% glycerol). Interestingly, the LucPpe2 reported by Leach et al. already contains isoleucine at the homologous position described for the *L. lateralis* mutant.

10 Although thermostability was the characteristic of interest, clones were selected based on the other enzymological parameters in the screens. By selecting clones having greater luminescence expression, mutants were found that yielded greater luminescence intensity in colonies of *E. coli*. However, the process showed little ability to alter the kinetic profile of luminescence by the 15 enzymes. This failure suggests that the ability to support steady-state luminescence is integral to the catalytic mechanism, and is not readily influenced by a cumulative effect of many amino acids.

Substrate binding was screened by measuring an apparent composite K_m (see Example 2) for luciferin and ATP. Although the apparent composite K_m 20 remained relatively constant, later analysis showed that the individual K_m 's systematically changed. The K_m for luciferin rose while the K_m for ATP declined (Table 3). The reason for this change is unknown, although it can be speculated that more efficient release of oxyluciferin or luciferin inhibitors could lead to more rapid enzyme turnover.

25 Each point mutation, on its own, increases (to a greater or lesser extent) the thermostability of the mutant enzyme relative to the wild-type luciferase. The cumulative effect of combining individual point mutations yields mutant luciferases whose thermostability is greatly increased from the wild-type, often on the order of a magnitude or more.

Table 3: Michaelis-Menten Constants for Mutants Created by Directed Evolution

		K_m -luciferin	K_m -ATP
5	Luc[T249M]	0.32 μ M	18 μ M
	Luc49-7C6	0.99 μ M	14 μ M
	Luc78-0B10	1.6 μ M	3.4 μ M
	Luc90-1B5	2.2 μ M	3.0 μ M

The following examples illustrate the methods and compositions
10 of the present invention and their embodiments.

EXAMPLE 1

Producing Thermostable Luciferases Of The Present Invention

Mutagenesis Method.

15 An illustrative mutagenesis strategy is as follows: From the “best” wild-type luciferase clone, that is a clone with increased thermostability and not appreciably diminished values for other parameters, random mutagenesis was performed by three variations of error-prone PCR. From each cycle of random mutagenesis, 18 of the best clones were selected. DNA was prepared from these 20 clones yielding a total of 54 clones. These clones represent new genetic diversity.

These 54 clones were combined and recombination mutagenesis was performed. The 18 best clones from this population were selected.

25 These 18 clones were combined with the 18 clones of the previous population and recombination mutagenesis was performed. From this screening, a new luciferase population of 18 clones was selected representing 6 groups of functional properties.

30 In this screening the new mutations of the selected 54 clones, either in their original sequence configurations or in recombinants thereof, were screened a second time. Each mutation was analyzed on the average about 10 times. Of the 90 clones used in the recombination mutagenesis, it was likely that at least 10 were functionally equivalent to the best clone. Thus, the best clone or recombinants thereof should be screened at least 100 times. Since this was greater than the number of clones used in the recombination, there was

significant likelihood of finding productive recombination of the best clone with other clones.

Robotic Processing Methods.

Heat transfers were controlled in the robot process by using thick aluminum at many positions where the 96-well plates were placed by the robotic arm. For example, all shelves in the incubators or refrigerator were constructed from $\frac{1}{4}$ inch aluminum. One position in particular, located at room temperature, was constructed from a block of aluminum of dimensions 4.5 x 7 x 6.5 inches.

When any 96-well plate was moved from a high temperature (e.g., incubators) or low temperature (e.g., refrigerator) to a device at room temperature, it was first placed on the large aluminum block for temperature equilibration. By this means, the entire plate would rapidly reach the new temperature, thus minimizing unequal evaporation for the various wells in the plate due to temperature differences. Heat transfers in a stack of 96-well plates placed in an incubator (e.g., for overnight growth of *E. coli*) were controlled by placing 1 mm thick sheets of aluminum between the plates. This allowed for more efficient heat transfer from the edges of the stack to the center. Mixing in the robotic process was controlled by having the plate placed on a shaker for several second after each reagent addition.

Please refer to Figure 14 for a schematic of the order in which the plates are analyzed and to Figure 15 for a robotic apparatus which can be programmed to perform the following functions:

1. Culture Dilution Method.

A plate with lid (Falcon 3075) containing cells (*E. coli* JM109) is placed on a shaker and mixed for 3-5 minutes.

A plate (with lid) is obtained from a carousel and placed in the reagent dispenser. 180 μ l of media (M9 minimal media) is added after removing the lid and placing on the locator near the pipetter. The plate is then placed in the pipetter.

The plate on the shaker is placed in the pipetter, and the lid removed and placed on the locator. Cells are transferred to the new plate using pipetting procedure (see "Dilution of Cells into New Cell Plate").

The lids are replaced onto both plates. The new plate is placed in the refrigerator and the old plate is returned to the carousel.

2. Luminescence Assay Method.

A plate containing cells is retrieved from the carousel and placed on the shaker for 3-5 minutes to fully mix the cells. The cells tend to settle from solution upon standing.

To measure Optical Density (O.D.), the plate is moved from the shaker to the locator near the luminometer; the lid is removed and the plate placed into the luminometer. The O.D. is measured using a 620 nm filter.

10 When it is finished, the plate is then placed in the refrigerator for storage.

The above steps are completed for all plates before proceeding with subsequent processing.

To prepare a cell lysate, the plate of cells is first retrieved from the refrigerator and mixed on the shaker to resuspend the cells. A new plate from the carousel without a lid is placed in the reagent dispenser and 20 μ l of Buffer A is added to each well. This is placed in the pipetting station.

The plate of cells in the shaker is placed in the pipetting station. A daughter plate is prepared using pipetting procedure (see "Pipetting Cells into the Lysis Plate") to prepare a daughter plate of cells.

20 After pipetting, the new daughter plate is placed on the shaker for mixing.

After mixing, the Lysate Plate is placed into a solid CO₂ freezing station to freeze the samples. The plate is then moved to the thaw block to thaw for 10 minutes.

25 The plate is then moved to the reagent dispenser to add 175 μ l of Buffer B, and then mixed on the shaker for about 15 minutes or more. The combination of the freeze/thaw and Buffer B will cause the cells to lyse.

30 A new plate with a lid from the carousel is used to prepare the dilution plate from which all assays will be derived. The plate is placed in the reagent dispenser and the lid removed to the locator near the pipetter. 285 μ l of Buffer C is added to each well with the reagent dispenser, then the plate is placed in the pipetting station.

The Lysate Plate in the shaker is moved to the pipetting station and pipetting procedure (see “Dilution from Lysis Plate to Incubation Plate”) is used. After pipetting, the new daughter plate is placed on the shaker for mixing. The Lysate Plate is discarded.

- 5 Two white assay plates (Labsystems #9502887) are obtained from the plate feeder and placed in the pipetter. The incubation plate from the shaker is placed in the pipetter, and the lid removed and placed on the nearby locator. Two daughter plates are made using the pipetting procedure (see “Create Pair of Daughter Plates from Incubation Plate”). Afterwards, the lid is replaced on the
- 10 parent plate, and the plate is placed in a high temperature incubator. [ranging from 31°C to about 65°C depending on the clone.]

One daughter plate is placed in the luminometer and the 1X assay method is used. After the assay, the plate is placed in the ambient incubator, and the second daughter plate is placed in the luminometer. For the second plate, the

- 15 0.02X assay method is used. This plate is discarded, and the first plate is returned from the incubator to the luminometer. The repeat assay method is used (i.e., no reagent is injected). Afterwards, the plate is again returned to the ambient incubator.

The above steps are completed for all plates before proceeding with

- 20 processing.

To begin the second set of measurements, the plate from the high temperature incubator is placed in the shaker to mix.

The plate in the ambient incubator is returned to the luminometer and the repeat assay method is again used. The plate is returned afterwards to the

- 25 ambient incubator.

Two white assay plates again are obtained from the plate feeder and placed in the pipetter. The plate on the shaker is placed in the pipetter, and the lid removed and placed on the nearby locator. Two daughter plates are again made using the pipetting procedure (see “Create Pair of Daughter Plates from

- 30 Incubation Plate”). Afterwards, the lid is replaced on the parent plate, and the plate is returned to the high temperature incubator.

One daughter plate is placed in the luminometer and the 1X assay method is again used. The plate is discarded after the assay. The second daughter plate is

then placed in the luminometer and the 0.06X assay method is used. This plate is also discarded.

The above steps are completed for all plates before proceeding with processing.

5 In the final set of measurements, the plate from the high temperature incubator is again placed in the shaker to mix.

The plate in the ambient incubator is returned to the luminometer and the repeat assay method is again used. The plate is discarded afterwards.

10 One white assay plate is taken from the plate feeder and placed in the pipetter. The plate from the shaker is placed in the pipetter, and the lid removed and placed on the nearby locator. One daughter plate is made using the pipetting procedure (see "Create Single Daughter Plate from Incubation Plate"). The lid is replaced on the parent plate and the plate is discarded.

15 The daughter plate is placed in the luminometer and the 1X assay method is used. The plate is discarded after the assay.

Buffers and Assay Reagents

Buffer A: 325 mM K₂HPO₄; 6.5 mM CDTA; 0.1% Triton X-100

20 Buffer B: 1X CCLR (Promega E153A); 1.25 mg/ml lysozyme; 0.04% gelatin

Buffer C: 10 mM HEPES; 150 mM NaCl; 1 mg/ml BSA; 5% glycerol; 0.1 mM EDTA

25 1X Assay reagent: 5 µM Luciferin; 175 µM ATP; 20 mM Tricine, pH 8.0; 0.1 mM EDTA
0.02X Assay reagent: 1:50 dilution of 1X Assay reagent
0.06X Assay reagent: 1:16.7 dilution of 1X Assay reagent

Pipetting Procedures

A. Pipetting Cells Into the Lysis Plate

30 Non-aseptic procedure using fixed tips

On the pipetter deck:

-place a plate containing approximately 200 µl JM109 cells per well without lid

-Lysate Plate containing 20 μ l of Buffer A

Procedure:

1. Move the tips to the washing station and wash with 1 ml.
2. Move to the cell plate and withdraw 60 μ l.
- 5 3. Move to the Lysate Plate and dispense 45 μ l.
4. Repeat steps 1-3 for all 96 samples.
5. At the conclusion of the procedure, step 1 is repeated to clean the tips.

Post-procedure:

- Place Lysate Plate onto the shaker.
- 10 - Place lid on plate with cells and place on carousel.
- Place Lysate Plate into the CO₂ freezer.

B. Dilution From Lysis Plate to Incubation Plate

On the pipetter deck:

- 15 - Lysate Plate containing 240 μ l of lysate
- Incubation Plate without lid containing 285 μ l of Buffer C

Procedure:

1. Move the tips to the washing station and wash with 0.5 ml.
2. Move to the Lysate Plate and withdraw 30 μ l.
- 20 3. Move to the Incubation Plate and dispense 15 μ l by direct contact with the buffer solution.
4. Repeat steps 1-3 for all 96 samples.
5. At the conclusion of the procedure, step 1 is repeated to clean the tips.

Post-procedure:

- 25 - Place Incubation Plate on shaker.
- Discard Lysate Plate.

C. Create Pair of Daughter Plates From Incubation Plate

This procedure is done twice

On the pipetter deck:

- 30 - Incubation Plate containing 100-300 μ l of solution without lid
- Two empty Assay Plates (white)

Procedure:

1. Move the tips to the washing station and wash with 0.5 ml.
2. Move to the Incubation Plate and withdraw 50 μ l.
3. Move to the first Assay Plate and dispense 20 μ l.
- 5 4. Move to the second Assay Plate and dispense 20 μ l.
5. Repeat steps 1-4 for all 96 samples.
6. At the conclusion of the procedure, step 1 is repeated to clean the tips.

Post-procedure:

- 10 1. Replace lid on Incubation Plate.
2. Place Incubation Plate in incubator.
3. Place first Assay Plate in luminometer.
4. Place second Assay Plate on carousel.

D. Create Single Daughter Plate From Incubation Plate

15 **On the pipetter deck:**

- Place incubation Plate containing 100-300 μ l of solution without lid and
- Empty Assay Plate (white)

Procedure:

1. Move the tips to the washing station and wash with 0.5 ml.
- 20 2. Move to the Incubation Plate and withdraw 40 μ l.
3. Move to the Assay Plate and dispense 20 μ l.
4. Repeat steps 1-3 for all 96 samples.
5. At the conclusion of the procedure, step 1 is repeated to clean the tips.

Post-procedure:

25 **25**

- Discard Incubation Plate and lid on Incubation Plate.
- Place Assay Plate in luminometer.

E. Dilution of Cells Into New Cell Plate

Aseptic procedure using fixed tips

30 **On the pipetter deck:**

- plate containing approximately 200 μ l of cells without lid
- new cell plate containing 180 μ l of Growth Medium without lid

Procedure:

1. Move to the cell plate and withdraw 45 μ l.
2. Move to the Cell Plate and dispense 20 μ l volume by direct liquid-to-liquid transfer.
- 5 3. Move to waste reservoir and expel excess cells.
4. Move to isopropanol wash station aspirate isopropanol to sterilize tips.
5. Move to wash station, expel isopropanol and wash tips.
6. Repeat steps 1-4 for all 96 samples.

Post-procedure:

- 10 - Replace lid on original plate of cells and place onto carousel.
- Replace lid on new cell plate and place into refrigerator.

Notes:

This procedure is used to prepare the cell plates used in the main analysis procedure. 180 μ l of M9 minimal growth medium is added by the reagent dispenser to each of the new cell plates just prior to initiating the pipetting procedure. The dispenser is flushed with 75% isopropanol before priming with medium. The medium also contains selective antibiotics to reduce potential contamination.

20 Luminometer Procedures**A. 1X Assay Method**

1. Place plate into luminometer.
2. Inject 100 μ l of 1X Assay reagent.
3. Measure luminescence for 1 to 3 seconds.
- 25 4. Repeat for next well.
5. Continue until all wells are measured.

B. 0.02X Assay Method

1. Place plate into luminometer.
- 30 2. Inject 100 μ l of 0.02X Assay reagent.
3. Measure luminescence for 1 to 3 seconds.
4. Repeat for next well.
5. Continue until all wells are measured.

C. 0.06X Assay Method

1. Place plate into luminometer.
2. Inject 100 μ l of 0.06X Assay reagent.
3. Measure luminescence for 1 to 3 seconds.
- 5 4. Repeat for next well.
5. Continue until all wells are measured.

D. Repeat Assay

1. Place plate into luminometer.
- 10 2. Measure luminescence for 1 to 3 seconds.
3. Repeat for next well.
4. Continue until all wells are measured.

In Vivo Selection Method

15 Five to seven nitrocellulose disks having 200-500 colonies per disk (1000-3500 colonies total) are screened per 2 microplates (176 clones) (Wood and DeLuca, 1987). The clones are screened at high temperatures using standard screening conditions.

20 Eight positions in each microplate are reserved from a reference clone using the "best" luciferase (the parent clone for random mutagenesis and codon mutagenesis). The positions of the reserved wells is shown as "X" below.

	XooooooooooooX
	oooooooooooooooo
25	oooXooooXoooo
	oooooooooooooooo
	oooooooooooooooo
	oooXooooXoooo
	oooooooooooooooo
30	XooooooooooooX

The reference clones are made by placing colonies from DNA transformed from the parent clone into the reference wells. To identify these

wells prior to inoculation of the microplate, the wells are marked with a black marking pen on the bottom of each well.

Screening Selection Criteria

The following criteria were used for screening purposes. The

5 temperature chosen for the enzyme stability parameter was such that the parent enzyme would decay 100 to 1000 fold over 10 hours (see Table 1). Criteria 1 is achieved manually; data for criteria 2-6 is generated by robotic analysis. For all criteria, the maximum value as described is selected.

10 1. ***In vivo screen.*** The brightest clones are selected at a given temperature.

15 2. **Expression/specific activity.** The value for normalized luminescence is calculated as the ratio of luminescence to optical density. The value is reported as the ratio with the reference value.

20 3. **Enzyme stability.** Measurements of normalized luminescence of the incubated samples (3 taken over about 15 hours) are fitted to $\ln(L) = \ln(L_0) - (t/\tau)$, where L is normalized luminescence and t is time. τ is a measure of the enzyme stability. The value is reported as the ratio with the reference value, and the correlation coefficients are calculated.

25 4. **Substrate binding.** Measurements of normalized luminescence with 1X and 0.02X are taken at the initial reading set, and 1X and 0.06X are taken at the 5 hour set. The ratio of the 0.02X:1X and 0.06X:1X gives the relative luminescence at 0.02X and 0.06X concentrations. These values, along with the relative luminescence at 1X (i.e., 1), are fitted to a Lineweaver-Burk plot to yield the $K_m:app, total$ for the substrates ATP, luciferin, and CoA. The values are reported as the inverse ratio with the reference value, and the correlation coefficients are calculated.

30 5. **Signal stability.** The luminescence of the initial 1X luminescent reactions are re-measured 3 additional times over about 15 hours. These values are fitted to $\ln(L) = \ln(L_0) - (t/\tau)$ and the integral over

t (15 hours) is calculated. Signal stability is then calculated as

$$S = (1 -$$

$\text{int}(L)/L_0 t)^2$. The values are reported as the inverse ratio with the reference value, and the correlation coefficients are calculated.

5 6. **Composite fitness.** The values of criteria 2 through 5 are combined into a single composite value of fitness (or commercial utility). This value is based on a judgment of the relative importance of the other criteria. This judgment is given below:

10	<u>Criteria</u>	<u>Relative Value</u>
	Enzyme Stability	5
	Signal Stability	2
	Substrate Binding	2
	Expression/Activity	1

15

The composite, $C = \text{Sum}(\text{criteria 2-5 weighted by relative value, e.g., more weight is on stability because that was a major goal})$.

EXAMPLE 2

20 **Software**

Organize data into SQL database

Each file created by a luminometer (96 well, Anthos, Austria) represents the data from one microplate. These files are stored in the computer controlling the luminometer, and connected to the database computer by a network link.

25 From each microplate of samples, nine microplates are read by the luminometer (the original microplate for optical density and eight daughter microplates for luminescence).

Ninety files are created in total; each containing data sets for 96 samples. Each data set contains the sample number, time of each measurement relative to the first measurement of the plate, luminometer reading, and background corrected luminometer reading. Other file header information is also given. The time that each microplate is read is also needed for analysis. This can be obtained from the robot log or the file creation time. A naming convention for

the files is used by the robot during file creation that can be recognized by SQL (e.g. YYMMDDPR.DAT where YY is the year, MM is the month, DD is the day, P is the initial plate [0-9], and R is the reading [0-8]).

5 Data Reduction and Organization

Normalize luminescence data: For each measurement of luminescence in the eight daughter plates, the normalized luminescence is calculated by dividing the relative light units by the optical density of the original plate. If any value of normalized luminescence is less than zero, assign the value of 0.1 sL where sL is
10 the standard deviation for measurements of normalized luminescence.

Calculate relative measurement time: For each normalized luminescence measurement, the time of the measurement is calculated relative to the first measurement of the sample. For example, the times of all luminescence measurements of sample B6 in plate 7 (i.e., 7:B06) are calculated relative to the
15 first reading of 7:B06. This time calculation involves both the time when the plate is read and the relative time of when the sample is read in the plate.

Calculate enzyme stability (τ): For each sample, use linear regression to fit $\ln(L_{1x}) = \ln(L_0) - (t/\tau)$ using the three luminescence measurements with 1X substrate concentrations (Plates 1, 5, 8). Also calculate the regression
20 coefficient.

Calculate substrate binding ($K_{m:app,total}$): Using microplates from the first set of readings (Plates 1 and 2), calculate the $L_{0.02x,rel}$ by dividing measurements made with substrate concentrations of 0.02X by those of 1X. Similarly, calculate the $L_{0.06x,rel}$ using microplates of the second set of readings (Plates 5 and
25 6), by dividing measurements made with substrate concentrations of 0.06X by those of 1X.

For each sample, use linear regression to fit $1/L = (K_{m:app,total}/L_{max:app}) (1/[S]) + (1/L_{max:app})$ using

	L	[S]
30	$L_{0.02x,rel}$	0.02
	$L_{0.06x,rel}$	0.06
	1 ($L_{1x,rel}$)	1

$K_{m:app,total}$ is calculated as the slope/intercept. Also calculate the regression coefficient.

Calculate signal stability (S): For each sample, use linear regression to fit $\ln(L) = \ln(L_0) - (t/\tau)$ using the four luminescence measurements of the initial 5 microplate with 1X substrate concentrations (Plates 1, 3, 4, and 7). Also calculate the regression coefficient. From the calculated values of τ and L_0 , calculate the integral of luminescence by $\text{int}(L) = \tau L_0 (1 - \exp(-t_f/\tau))$, where t_f is the average time of the last measurement (e.g., 15 hours). The signal stability is calculated as $S = (1 - \text{int}(L)/L_i t_f)^2$, where L_i is the initial measurement of 10 normalized luminescence with 1x substrate concentration (Plate 1).

[Note: To correct for evaporation, an equation $S = (1 + K - \text{int}(L)/L_i t_f)^2$, may be used where $1/K = 2(\text{relative change of liquid volume at } t_f)$.]

Calculate the reference value surfaces: A three dimensional coordinate system can be defined by using the grid positions of the samples within a 15 microplate as the horizontal coordinates, and the calculated values for the samples (L_i , τ , $K_{m:app,total}$, or S) as the vertical coordinates. This three dimensional system is referred to as a “plate map”. A smooth surface in the plate maps representing a reference level can be determined by least squares fit of the values determined for the 8 reference clones in each microplate. For each 20 of the 10 initial microplates of samples, respective reference surfaces are determined for the criteria parameters L_i , τ , $K_{m:app,total}$, and S (40 surfaces total).

In the least squares fit, the vertical coordinates (i.e., the criteria parameters) are the dependent variables, the horizontal coordinates are the independent variables. A first order surface (i.e., $z = ax + by + c$) is fitted to the 25 values of the reference clones. After the surface is calculated, the residuals to each reference clone are calculated. If any of these residuals is outside of a given cutoff range, the reference surface is recalculated with omission of the aberrant reference clone.

If a first order surface does not sufficiently represent the values of the 30 reference clones, a restricted second order surface is used (i.e., $z = a(x^2 + ky^2) + bx + cy + d$, where k is a constant).

Calculate the reference-normalized values: For the criteria parameter of each sample, a reference-normalized value is determined by calculating the ratio

or inverse ratio with the respective reference value. The reference-normalized values are L_i / L_{ir} , τ / τ_r , $K_{mr} / K_{m:app,total}$, and S_r / S , where reference values are calculated from the equations of the appropriate reference surface.

Calculate the composite scores: For each sample, calculate

5 $C = 5(\tau / \tau_r) + 2(S_r / S) + 2(K_{mr} / K_{m:app,total}) + (L_i / L_{ir})$.

Determine subgroupings: For the criteria parameters L_i , τ , $K_{m:app,total}$, S , and C , delimiting values (i.e., bin sizes) for subgroupings are defined as gL , $g\tau$, gK_m , gS , and gC . Starting with the highest values for L_i , τ , or C , or the lowest values of $K_{m:app,total}$ or S , the samples are assigned to bins for each criteria

10 parameter (the first bin being #1, and so on).

Display sorted table of reference-normalized values: Present a table of data for each sample showing in each row the following data:

- sample identification number (e.g., 7:B06)
- composite score (C)
- 15 - reference-normalized enzyme stability (τ / τ_r)
- correlation coefficient for enzyme stability
- bin number for enzyme stability
- reference-normalized signal stability (S_r / S)
- correlation coefficient for signal stability
- 20 - bin number for signal stability
- reference-normalized substrate binding ($K_{mr} / K_{m:app,total}$)
- correlation coefficient for substrate binding
- bin number for substrate binding
- reference-normalized expression/specific activity (L_i / L_{ir})
- 25 - bin number for expression/specific activity

The table is sorted by the composite score (C).

Present sorted table of criteria parameters.

Present a table of data for each sample showing in each row the following data:

- 30 - sample identification number
- composite score (C)
- enzyme stability (τ)
- correlation coefficient for enzyme stability

- bin number for enzyme stability
- signal stability (S)
- correlation coefficient for signal stability
- bin number for signal stability

5 - substrate binding ($K_{m:app,total}$)

- correlation coefficient for substrate binding
- bin number for substrate binding
- expression/specific activity (L_i)
- bin number for expression/specific activity

10 The table is sorted by the composite score (C); the reference clones are excluded from the table. Same entry coding by standard deviation as described above.

Present sorted table of reference-normalized values

15 This is the same procedure as the final step of the data reduction procedure. The table will show:

- sample identification number
- composite score (C)
- reference-normalized enzyme stability (τ/τ_r)
- correlation coefficient for enzyme stability

20 - bin number for enzyme stability

- reference-normalized signal stability (S_r/S)
- correlation coefficient for signal stability
- bin number for signal stability

25 - reference-normalized substrate binding ($K_{mr}/K_{m:app,total}$)

- correlation coefficient for substrate binding
- bin number for substrate binding
- reference-normalized expression/specific activity (L_i/L_{ir})
- bin number for expression/specific activity

30 The table is sorted by the composite score (C); the reference clones are excluded from the table. Same entry coding by standard deviation as described above.

Present sorted table of criteria parameters for reference clones

This is the same procedure as described above for criteria parameters, except for only the reference clones. The table will show:

- sample identification number
- 5 - composite score (C)
- enzyme stability (τ)
- correlation coefficient for enzyme stability
- bin number for enzyme stability
- signal stability (S)
- 10 - correlation coefficient for signal stability
- bin number for signal stability
- substrate binding ($K_{m:app,total}$)
- correlation coefficient for substrate binding
- bin number for substrate binding
- 15 - expression/specific activity (L_i)
- bin number for expression/specific activity

The table is sorted by the composite score (C). Same entry coding by standard deviation as described above.

Present sorted table of reference-normalized values

20 This is the same procedure as described above for reference-normalized values, except for only the reference clones. The table will show:

- sample identification number
- composite score (C)
- reference-normalized enzyme stability (τ/τ_r)
- 25 - correlation coefficient for enzyme stability
- bin number for enzyme stability
- reference-normalized signal stability (S_r/S)
- correlation coefficient for signal stability
- bin number for signal stability
- 30 - reference-normalized substrate binding ($K_{mr}/K_{m:app,total}$)
- correlation coefficient for substrate binding
- bin number for substrate binding
- reference-normalized expression/specific activity (L_i/L_{ir})

- bin number for expression/specific activity

The table is sorted by the composite score (C). Same entry coding by standard deviation as described above.

Sort table

5 Any table may be sorted by any entries as primary and secondary key.

Display histogram of table

For any table, a histogram of criteria parameter vs. bin number may be displayed for any criteria parameter.

Display plate map

10 For any plate, a plate map may be displayed showing a choice of:

- any luminescence or optical density measurement
- L_i
- L_i reference surface
- L_i / L_{ir}

15 - τ

- τ reference surface
- τ / τ_r
- correlation coefficient of τ

- S

20 - S reference surface

- S_r / S
- correlation coefficient of S

- $K_{m:app,total}$
- K_m reference surface

25 - $K_{mr} / K_{m:app,total}$

- correlation coefficient for $K_{m:app,total}$
- composite score (C)

The plate maps are displayed as a three dimensional bar chart. Preferably, the bars representing the reference clones are indicated by color or some other

30 means.

Display drill-down summary of each entry

For L_i , τ , $K_{m:app,total}$, and S, any entry value in a table may be selected to display the luminescence and optical density reading underlying the value

calculation, and a graphical representation of the curve fit where appropriate. Preferably the equations involved and the final result and correlation coefficient will also be displayed.

5 L_i or L_i/L_r . Display the optical density and luminescence value from the chosen sample in Plate 0 and Plate 1.

τ or τ/τ_r . Display the optical density and luminescence value from the chosen sample in Plate 0, Plate 1, Plate 5, and Plate 8. Display graph of $\ln(L_1X)$ vs. t , showing data points and best line.

10 S or S_r/S . Display the optical density and luminescence value from the chosen sample in Plate 0, Plate 1, Plate 3, Plate 4, and Plate 7. Display graph of $\ln(L)$ vs. t , showing data points and best line.

$K_{m:app,total}$ or $K_{mr}/K_{m:app,total}$. Display the optical density and luminescence value from the chosen sample in Plate 0, Plate 1, Plate 2, Plate 5, and Plate 6.

Display graph of $1/L$ vs. $1/[S]$, showing data points and best line.

15 **EXAMPLE 3**

Preparation Of Novel Luciferases

The gene shown in Figure 45 contains a single base pair mutation which encodes an amino acid substitution at position 249, T to M. This clone has a spectral maximum of 552 nm which is yellow shifted from the sequence of Luc.

20 This mutant was selected as an original template because it produces about 5 times brighter luminosity *in vivo* which allowed for more efficient screening.

C-terminus mutagenesis

To eliminate the peroxisome targeting signal (SKL), the L was mutated to a STOP codon and the 3 codons immediately upstream were randomized according to the oligonucleotide mutagenesis procedure described herein. The mutagenic oligonucleotide designed to accomplish this also introduces a unique SpeI site to allow mutant identification without sequencing. The mutants were screened *in vivo* and 13 colonies picked, 12 of which contained the SpeI site.

N-terminus mutagenesis

30 To test if expression could be improved, the 3 codons immediately downstream from the initiation Met were randomized as described herein. The mutagenic oligo designed to accomplish this also introduces a unique ApaI site

to allow mutant identification without sequencing. Seven clones were selected, and six of the isolated plasmids were confirmed to be mutants.

Shuffling of C- and N-terminus mutants

The C- and N-terminus mutagenesis were performed side-by-side. To 5 combine the N- and C-terminus mutations, selected clones from each mutagenesis experiment were combined with the use of recombination mutagenesis according to the recombination mutagenesis protocol described herein. The shuffled mutants were subcloned into amp^S pRAM backbone and screened in DH5 F'IQ (BRL; Hanahan, 1985). A total of 24 clones were picked, 10 only 4 contained both the N- and C- terminus mutations. These 4 clones were used as templates for randomization of the cysteine positions in the gene.

Mutagenesis to randomize cysteine positions/Random mutagenesis and recombination mutagenesis in the Luc gene

There are 7 cysteine positions in LucPpe2. It is known that these 15 positions are susceptible to oxidation which could cause destabilization of the protein. Seven oligonucleotides were ordered to randomize the cysteine positions.

The oligonucleotides were organized into two groups based upon the conservation of cysteine in other luciferase genes from different families. Group 20 1 randomizes the conserved cysteine positions C-60, C-80, and C-162. Group 2 randomizes cysteines that are not strictly conserved at positions C-38, C-127, C-221, and C-257.

The four selected templates from the N- and C-terminus mutagenesis 25 were sub-cloned into an ampicillin-sensitive backbone and single-stranded DNA was prepared for each of the templates. These templates were combined in equal amounts and oligonucleotide mutagenesis was completed as described herein. It was determined by plating an aliquot of the mutS transformation prior to overnight incubation that each of the 2 groups contained 2 x 10⁴ independent transformants. MutS-DNA was prepared for the 2 groups and was then 30 transformed into JM109 cells for screening. Mutants from group 1 were screened *in vivo* and picks were made for a full robotic run. Five clones were selected that had improved characteristics. Mutants from group 2 were screened *in vivo* and picks were made for a full robotic run. The temperature incubator on

the robot was set at 33°C for this set of experiments. Ten clones were selected that had improved characteristics. The fifteen best picks from both groups of the cysteine mutagenesis experiments were shuffled together as described herein and 18 of the best clones were selected after robotic processing.

5 The “best” clone from the above experiment (Luc31-1G8) was selected as a template for subsequent rounds of mutagenesis. (The high temperature robot incubator temperature was set to 42°C.) Another complete round of mutagenesis was completed.

10 The 18 best clones from the above mutagenesis were picked and clone (Luc39-5B10) was selected as the best clone and was used as a template for another round of mutagenesis. (The high temperature robot incubator temperature was set at 49°C).

15 After this cycle, 6 of the best clones were selected for sequencing (the nucleotide sequence and inferred amino acid sequence of five of the clones is shown in Figures 22-26 and 27-31, respectively). Based upon the sequence data, nine positions were selected for randomization and seven oligos were designed to cover these positions. Based upon data generated from the robot, it was determined that the best clone from the group of six clones that were sequenced was clone (Luc49-7C6, Figures 22 and 27). The luciferase gene from this clone 20 was sub-cloned into an ampicillin-sensitive pRAM backbone and single stranded DNA was prepared. The randomization of the selected positions was completed according to the oligonucleotide mutagenesis procedure listed herein.

25 The randomization oligonucleotides were divided into 4 groups, and transformants from these experiments were picked and two robotic runs were completed. Ten clones were selected from the two experiments. (The high temperature robot incubator temperature on robot was set at 56°C).

The best 10 picks from the above two experiments, and the best 18 picks from the previous population of clones were shuffled together (recombination mutagenesis protocol).

30 The 18 best clones were selected and clone Luc58-0A5 was determined to be the best clone. This clone was then used as a template for another round of mutagenesis. The high temperature robot incubator temperature was set at 58°C.

Clone Luc71-504 was selected as a new lead clone and another round of mutagenesis was completed. Incubator set at 60°C.

The best 18 picks were selected. The nucleotide sequence and inferred amino acid sequence of 4 clones from experiment 78 are shown in Figures 32-35 and 36-39, respectively, and the best clone from this group was determined to be clone Luc78-0B10. The thermostability of clones at various temperatures is presented in the Figures.

EXAMPLE 4

Mutagenesis Strategy from Clone Luc78-0B10 to Luc90-1B5

10 Twenty-three oligonucleotides were prepared to change 28 positions to consensus. All of the oligonucleotides were tested individually using oligonucleotide directed mutagenesis with single stranded DNA from clone *luc78-0B10* as a template to determine which oligonucleotides gave an improvement in thermostability. Table 4 lists the mutagenic oligonucleotides.

15

Table 4

OLIGO SYNTHESIS			
	DESCRIPTION	NUMBER	SEQ ID NO.
5	A17 to T	6215	48
	M25 to L	6216	49
	S36 to P; remove <i>Nsi</i> I site	6217	50
	A101 to V, S105 to N	6218	51
	I125 to V	6219	52
	K139 to Q	6220	53
10	V145 to I	6221	54
	V194 to I	6222	55
	V203 to L, S204 to P	6231	56
	A216 to V	6232	57
	A229 to Q	6233	58
	M249 to T (reversion)	6234*	59
15	T266 to R, K270 to E	6235	60
	E301 to D	6236	61
	N333 to P, F334 to G	6237	62
	R356 to K	6238	63
	I363 to V	6246	64
	A393 to P	6247	65
20	R417 to H	6248	66
	G482 to V	6249	67
	N492 to T	6250	68
	F499 to Y, S501 to A	6251	69
	L517 to V	6252	70
	F537 to L	6253	71

*Note that oligonucleotide #6234 does not change a consensus position. This oligonucleotide causes a reversion of position 249 to the wild-type Ppe-2 codon.

30 Although reversion of this position was shown to increase thermostability at 62°C, reversion of this position decreased light output.

Three oligonucleotide-directed mutagenesis experiments with clone *luc78-0B10* as a template were completed. The oligonucleotides for these 35 experiments were divided in the following manner:

a. 6215, 6234, 6236, 6248 (found to give increased thermostability)

b. 6215, 6217, 6218, 6219, 6220, 6221, 6222, 6231, 6233, 6234, 6236, 6238, 6247, 6248, 6249, 6251, 6253 (found to be neutral or have increased thermostability).

c. All 23 oligonucleotides.

5 Selections from the three experiments listed above were screened with the robotic screening procedure (experiment 84, see Table 1) using *luc78-0B10* as a control. Selections from experiment 84 were recombined using the recombination mutagenesis procedure and then screened with the robotic screening procedure (experiment 85).

10 Single stranded DNA was prepared from three clones, *luc85-3E12*, *luc85-4F12*, *luc85-5A4*. The nucleotide sequence and inferred amino acid sequence of *luc85-4F12* are shown in Figures 40 and 41, respectively. These clones were used as templates for oligonucleotide-directed mutagenesis to improve codon usage. Positions were selected based upon a codon usage table
15 published in *Nucleic Acids Research*, vol. 18 (supplement) 1990, page 2402. The table below lists oligonucleotides that were used to improve codon usage in *E. coli*.

Table 5

	Description	Oligo Synthesis #	SEQ ID NO.
5	L7(tta-ctg), remove <i>Apa</i> I site	6258	72
	L29(tta-ctg)	6259	73
	T42(aca-acc)	6260	74
	L51, L56(tta-ctg), L58(ttg-ctg)	6261	75
	L71(tta-ctg)	6262	76
	L85(ttg-ctg)	6263	77
10	L95(ttg-ctg), L97(ctt-ctg)	6273	78
	L113, L117(tta-ctg)	6274	79
	L151, L153(tta-ctg)	6275	80
	L163(ctc-ctg)	6276	81
	R187(cga-cgt)	6277	82
	L237(tta-ctg)	6279	83
15	R260(cga-cgc)	6280	84
	L285, L290(tta-ctg), L286(ctt-ctg)	6281	85
	L308(tta-ctg)	6282	86
	L318(tta-ctg)	6283	87
	L341(tta-ctg), T342(aca-acc)	6284	88
	L380(ttg-ctg)	6285	89
20	L439(tta-ctg)	6286	90
	L456(ctc-ctg), L457(tta-ctg)	6293	91
	T506(aca-acc), L510(cta-ctg)	6305	92
	R530(aga-cgt)	6306	93

25

In the first experiment, the three templates listed above from experiment 85 were combined and used as a templates for oligonucleotide-directed mutagenesis. All of the oligonucleotides were combined in one experiment and clones resulting from oligonucleotide-directed mutagenesis were screened using 30 the robotic screening procedure as experiment 88. There were a low percentage of luminescent colonies that resulted from this experiment, so another oligonucleotide-directed mutagenesis experiment was completed in which the oligonucleotides were combined in the following groups:

- 35 a. 6258, 6273, 6280, 6286
- b. 6259, 6274, 6281, 6293
- c. 6260, 6275, 6282, 6294

- d. 6261, 6276, 6283, 6305
- e. 6262, 6277, 6284, 9306
- f. 6263, 6279, 6285

5 It was discovered that samples from group b had a low number of luminescent colonies, and it was hypothesized that one of the oligonucleotides in group b was causing problems. Selections were made from all of the experiments with the exception of experiment b. Samples were then run through the robotic screening procedure (experiment 89). Selections from experiments
10 88 and 89 were shuffled together with the recombination mutagenesis protocol and were then screened with the robotic screening procedure (experiment 90).

Materials and Methods

A. Mutagenesis Protocol

15 The mutant luciferases disclosed herein were produced via random mutagenesis with subsequent *in vivo* screening of the mutated genes for a plurality of characteristics including light output and thermostability of the encoded luciferase gene product. The mutagenesis was achieved by generally following a three-step method:

- 20 1. Creating genetic diversity through random mutagenesis. Here, error-prone PCR of a starting sequence was used to create point mutations in the nucleotide sequence. Because error-prone PCR yields almost exclusively single point mutations in a DNA sequence, a theoretical maximum of 7 amino acid changes are possible per nucleotide mutation.
25 In practice, however, approximately 6.1 amino acid changes per nucleotide is achievable. For the 550 amino acids in luciferase, approximately 3300 mutants are possible through point mutagenesis.
- 30 2. Consolidating single point mutations through recombination mutagenesis. The genetic diversity created by the initial mutagenesis is recombined into a smaller number of clones by sPCR. This process not only reduces the number of mutant clones, but because the rate of mutagenesis is high, the probability of linkage to negative mutations is significant. Recombination mutagenesis unlinks positive mutations from

negative mutations. The mutations are "re-linked" into new genes by recombination mutagenesis to yield the new permutations. Then, after re-screening the recombination mutants, the genetic permutations that have the "negative mutations" are eliminated by not being selected.

5 Recombination mutagenesis also serves as a secondary screen of the initial mutants prepared by error-prone PCR.

3. Broadening genetic diversity through random mutagenesis of selected codons. Because random point mutagenesis can only achieve a limited number of amino acid substitutions, complete randomization of selected 10 codons is achieved by oligonucleotides mutagenesis. The codons to be mutated are selected from the results of the preceding mutagenesis processes on the assumption that for any given beneficial substitution, other alternative amino acid substitutions at the same positions may produce even greater benefits. The positions to be mutated are identified 15 by DNA sequencing of selected clones.

B. Initial mutagenesis experiments

Both the N-terminus and the C-terminus of the starting sequence were modified by oligonucleotide-directed mutagenesis to optimize expression and 20 remove the peroxisomal targeting sequence. At the N-terminus, nine bases downstream of the initiation codon were randomized. At the C-terminus, nine bases upstream of the termination codon were randomized. Mutants were analyzed using an *in vivo* screen, resulting in no significant change in expression.

Six clones from this screen were pooled, and used to mutate the codons 25 for seven cysteines. These codons were randomized using oligonucleotide-directed mutagenesis, and the mutants were screened using the robotic screening procedure. From this screen, fifteen clones were selected for directed evolution.

C. Generating and Testing Clones

30 Several very powerful and widely known protocols are used to generate and test the clones of the present invention. Unless noted otherwise, these laboratory procedures are well known to one of skill in the art. Particularly noted as being well known to the skilled practitioner is the polymerase chain

reaction (PCR) devised by Mullis and various modifications to the standard PCR protocol (error-prone PCR, sPCR, and the like), DNA sequencing by any method (Sanger or Maxxam & Gilbert's methodology), amino acid sequencing by any method (e.g., the Edman degradation), and electrophoretic separation of 5 polynucleotides and polypeptides/proteins.

D. Vector Design

A preferred vector (pRAM) (see Figure 20) used for the mutagenesis procedure contains several unique features that allow for the mutagenesis 10 strategy to work efficiently:

The pRAM vector contains a filamentous phage origin, f1, which is necessary for the production of single-stranded DNA.

Two SfiI sites flank the gene. These sites were designed by so that the gene to be subcloned can only be inserted in the proper orientation.

15 The vector contains a *tac* promoter.

Templates to be used for oligonucleotide mutagenesis contain a 4 base-pair deletion in the *bla* gene which makes the vector ampicillin-sensitive. The oligonucleotide mutagenesis procedure uses a mutant oligonucleotide as well as an ampicillin repair oligonucleotide that restores function to the *bla* gene. This 20 allows for the selection of a high percentage of mutants. (If selection is not used, it is difficult to obtain a high percentage of mutants.)

E. Uses of Luciferases

The mutant luciferases of the present invention are suitable for use in any 25 application for which previously known luciferases were used, including the following:

ATP Assays. The greater enzyme stability means that reagents designed for detection of ATP have a greater shelf-life and operational-life at higher temperatures (e.g., room temperature). Therefore, a method of detecting ATP 30 using luciferases with increased thermostability is novel and useful.

Luminescent labels for nucleic acids, proteins, or other molecules.

Analogous to advantages of the luciferases of the present invention for ATP assays, their greater shelf-life and operational-life is a benefit to the reliability

and reproducibility of luminescent labels. This is particularly advantageous for labeling nucleic acids in hybridization procedures where hybridization temperatures can be relatively high (e.g., greater than 40°C). Therefore, a method of labeling nucleic acids, proteins, or other molecules using luciferases 5 of the present invention is novel and useful.

Genetic reporter. In the widespread application of luciferase as a genetic reporter, where detection of the reporter is used to infer the presence of another gene or process of interest, the increased thermostability of the luciferases provides less temperature dependence of its expression in living cells and in cell-10 free translations and transcription/translation systems. Therefore, a method using the luciferases of the present invention as genetic reporters is novel and useful.

Enzyme immobilization. Enzymes in close proximity to physical surfaces can be denatured by their interaction with that surface. The high density 15 immobilization of luciferases onto a surface to provide strong localized luminescence is improved by using thermostable luciferases. Therefore, a method of immobilizing luciferases onto a solid surface using luciferases of the present invention is novel and useful.

Hybrid proteins. Hybrid proteins made by genetic fusion genes encoding 20 luciferases and of other genes, or through a chemical coupling process, benefit by having a greater shelf-life and operational-life. Therefore, a method of producing hybrid proteins through genetic means or chemical coupling using the luciferases of the present invention is novel and useful.

High temperature reactions. The light intensity of a luciferase reaction 25 increases with temperature until the luciferase begins to denature. Because the use of thermostable luciferases allows for use at greater reaction temperatures, the luciferases of the present invention are novel and useful for performing high temperature reactions.

Luminescent solutions. Luminescence has many general uses, including 30 educational, demonstrational, and entertainment purposes. These applications benefit from having enzymes with greater shelf-life and operational-life. Therefore, a method of making luminescent solutions using the luciferases of the present invention is novel and useful.

F. Firefly luciferase

The firefly luciferase gene chosen for directed evolution was *LucPpe2* isolated from *Photuris pennsylvanica*. The luciferase was cloned from fireflies collected in Maryland by Wood et al. and later was independently cloned by Dr. 5 Leach using fireflies collected in Oklahoma (Ye et al., 1997). A mutant of this luciferase (T249M) was made by Wood et al. and used in the present invention because it produced approximately 5-fold more light when expressed in colonies of *E. coli*.

Overview of Evolution Process: Directed evolution was achieved 10 through a recursive process, each step consisting of multiple cycles of 1) creating mutational libraries of firefly luciferase followed by 2) screening the libraries to identify new mutant clones having a plurality of desired enzymological characteristics.

To begin the process, three mutational libraries were created using error-prone PCR (Fromant *et al.*, 1995). Each library was screened first by visual 15 evaluation of luminescence in colonies of *E. coli* (Wood and De Luca, 1987), and then by quantitative measurements of enzymological properties in *E. coli* cell lysates. Approximately 10,000 colonies were examined in the visual screen, from which 704 were selected for quantitative analysis. From each quantitative 20 screen 18 clones were selected. The three sets of 18 clones each were pooled together, and a new mutational library was created using DNA shuffling to generate intragenetic recombinations (sPCR; Stemmer, 1994). The results were screened to yield another set of 18 clones. The entire process was completed by combining this set of 18 clones with 18 clones from the previous 25 round of evolution, creating another mutational library by DNA shuffling, and screening as before.

Screening method: In the qualitative visual screen, colonies were selected only for their ability to sustain relatively bright luminescence. The thermal stability of the luciferase within the colonies of *E. coli* was progressively 30 challenged in successive rounds of evolution by increasing the temperature of the screen. The selected colonies were inoculated into wells of 96-well plates each containing 200 μ l of growth medium.

In the quantitative screens, lysates of the *E. coli* cultures were measured for 1) luminescence activity, 2) enzyme stability, 3) sustained enzymatic turnover, and 4) substrate binding.

“Luminescence activity” was measured as the ratio of luminescence 5 intensity to the optical density of the cell culture.

“Enzyme stability” was determined by the rate of activity loss from cell lysates over 10 hours. In successive rounds of evolution the incubation temperature of the lysates was increased.

“Sustained enzymatic turnover” was determined by the rate of 10 luminescence loss of a signal enzymatic reaction over 10 hours at room temperature.

“Substrate binding” was determined by the relative activity of the lysate when assayed with diluted substrate mixtures. Of these four parameters, the highest priority for selection was placed on thermostability.

15 **Robotic Automation:** Robotic automation was used in the quantitative screens to accurately perform the large number of required quantitative assays on the cultured cells. Overnight cultures were first diluted into fresh medium and grown for 3 hours to produce cultures in mid-log phase growth. The optical densities of each culture was then measured, and aliquots of the cultures were 20 lysed by freeze/thaw and lysozyme. The resulting lysates were further diluted before analysis and incubated at elevated temperatures. Luminescence was measured from aliquots of the diluted lysates, taken at various times, and measured under various conditions as prescribed by the analytical method (see Example 2). Computer analysis of this data yielded the quantitative selection 25 criteria described herein.

Summary of evolutionary progression: After mutagenesis of the N- and C-termini, and randomization of the cysteine codons, a pool of 15 clones was subjected to two rounds of directed evolution as described herein. Five of the 18 clones resulting from this process were sequenced to identify mutations. One of 30 these clones designated, Luc49-7C6, was chosen for more detailed analysis and further mutagenesis. This clone contained 14 new amino acid substitutions compared to the luciferase Luc[T249M].

To assess the potential for other amino acid replacements at the sites of these substitutions, oligonucleotide-directed mutagenesis was used to randomize these codons. The resulting clones were screened as described herein, and 18 selected clones were used to initiate two new rounds of directed evolution. Of 5 the 18 clones resulting from this second set of rounds, the clone designated Luc78-0B10 was chosen for additional study and mutagenesis. This clone encoded a luciferase that contained 23 new amino acid substitutions compared to Luc[T249M].

Using oligonucleotide directed mutagenesis with Luc78-0B10 as the 10 template, codons were selected for substitution to consensus amino acids previously known among beetle luciferases. Selections from this mutagenesis experiment were shuffled together and three clones, determined to be the most stable were then used as templates for oligonucleotide mutagenesis to improve codon usage in *E. coli*. A clone designated Luc90-1B5 selected from this 15 experiment, contained 34 amino acid substitutions relative to Luc[T249M] (see Figures 42 and 43 for the nucleotide sequence and inferred amino acid sequence of *luc90-1B5*, and Figures 44 and 45 for the nucleotide sequence encoding and the inferred amino acid sequence of Luc[T249M]). Out of 25 codons selected for change to consensus amino acids, 11 were replaced in the clone designated 20 Luc90-1B5. Only five out of the 30 positions that were selected for improved codon usage were substituted and had little effect on enzyme expression.

Protein purification: Four mutants that are described herein (Luc[T249M], Luc49-7C6, Luc78-0B10, and Luc90-1B5) were purified using a previously published procedure (Hastings et al., 1996).

25 Enzymological characterization: Purified proteins were diluted in 25 mmol/L HEPES pH 7.8, 150 mmol/L NaCl, 0.1 mmol/L EDTA, 1 mg/ml BSA. Enzyme stability was determined from diluted proteins incubated at different temperatures, and aliquots were removed at different time points. A linear regression of the natural log of the luminescence and time was calculated. 30 Half-life was calculated as the $1n(0.5)/\text{slope}$ of the regression.

G. PCR Mutagenesis Protocol (Random Mutagenesis)PCR mutagenesis reactions

1. Prepare plasmid DNA from a vector containing the gene of interest, estimate DNA concentration from a gel.

5 2. Set up two 50 μ l reactions per group:

There are three groups of mutagenic conditions using different skewed nucleotide concentrations.

The conditions listed herein yield in the range of from 8-10% wild-type Luc colonies after subcloning phenotypic for each generated parent clone. The 10 rate of mutagenesis is estimated by the number of luminescent colonies that are present after mutagenesis. Based upon results of clones mutated in the range of 8-10%, it was determined that this level of mutagenesis produces on average approximately 2-3 amino acid changes per gene. If the mutagenesis rate is selected so that on average there is one amino acid change per gene, then on 15 average 50% of the clones will have no mutations. (Bowie et al., 1990).

For the master mix: add all components (see Table 6) except polymerase, vortex, spin briefly, add polymerase, and mix gently.

Table 6

Component	A to T/T to A	A to C/T to G	G to A/C to T
dATP	0.3 mM	0.1 mM	0.25 mM
dCTP	2.75 mM	4 mM	1 mM
dGTP	0.06 mM	0.02 mM	0.05 mM
dTTP	0.625 mM	0.3 mM	0.6 mM
⁺⁺ pRAMtailUP	0.4 pmol/ μ l	0.4 pmol/ μ l	0.4 pmol/ μ l
⁺⁺ pRAMtailDN	0.4 pmol/ μ l	0.4 pmol/ μ l	0.4 pmol/ μ l
*Taq Polymerase	1 U/ μ l	1 U/ μ l	1 U/ μ l
^o MgCl ₂	6.77 mM	5.12 mM	2.7 mM
^o MnCl ₂	0.5 mM	0.5 mM	0.3 mM
DNA	50 ng total	50 ng total	50 ng total
10X PCR buffer	1X	1X	1X
Autoclaved nanopure water	To 50 μ l	To 50 μ l	To 50 μ l

35 * Taq Polymerase is purchased from Perkin Elmer (N808-0101).

^o MnCl₂ and MgCl₂ are made fresh from 1 M stocks. The stocks are filter sterilized and mixed with sterile water to make the 10 mM and 25 mM stocks which are then stored in Polystyrene Nalgene containers at 4°C.

⁺⁺ pRAMtailUP: 5'-gtactgagacgacgccagcccaagcttaggcctgagt-3' (SEQ ID NO:38); pRAMtailDN: 5'-ggcatgagcgtgaactgactgaactagcggccgccgag-3' (SEQ ID NO:39)

5 10X PCR polymerase buffer:

- 100 mM Tris-HCl pH 8.4 from 1 M stock
- 500 mM Kcl
- Primers are diluted from a 1 nmol/ μ l stock to a 20 pmol/ μ l working stock.

10 Cycle in thermal cycler: 94°C for 1 minute (94°C for 1 minute, 72°C for 10 minutes) 10X

3. Purify reaction products with Wizard PCR purification kit (Promega Corporation, Madison, Wisconsin, part#A718c):

- transfer PCR reaction into a new tube containing Promega 100 μ l Direct Purification buffer (Promega part#A724a)
- add 1 ml of Wizard PCR Purification Resin (Promega part#A718c) Promega and incubate at room temperature for 1 minute
- pull resin though Wizard minicolumn
- wash with 80% ethanol
- spin in microcentrifuge to remove excess ethanol
- elute into 50 μ l sterile nanopure water (allow water to remain on column for at least 1 minute)

Amplification¹ Of Mutagenesis Reaction

1. Set up five 50 μ l reactions (see Table 7) per group.

Table 7

	Components	Concentration	Amount in 50 μ l	Final concentration
5	dATP	10 mM	1 μ l	0.2 mM
	dCTP	10 mM	1 μ l	0.2 mM
	dGTP	10 mM	1 μ l	0.2 mM
	dTTP	10 mM	1 μ l	0.2 mM
	+pRAM18UP	20 pmol/ μ l	1 μ l	0.4 pmol/ μ l
10	+pRAM19DN	20 pmol/ μ l	1 μ l	0.4 pmol/ μ l
	Pfu polymerase	2 U/ μ l	1 μ l	0.04 μ g/ μ l
	°10X buffer	10X	5 μ l	1X
	DNA		10 μ l	
	Water		24.6 μ l	

15

- To master mix: add all components, except polymerase, vortex, spin briefly, add polymerase, mix gently.

° 10X reaction buffer for Native Pfu polymerase contains 20 mM MgCl₂, so no additional MgCl₂ needs to be added

20

+ primers:

pRAM18UP -5'-gtactgagacgacgccag-3' (SEQ ID NO:40)

pRAM19DN -5'-ggcatgagcgtgaactgac-3' (SEQ ID

NO:41)

Cycling conditions: 94°C for 30 seconds (94°C for 20 seconds,

25

65°C for 1 minute, 72°C for 3 minutes) 25X

(Perkin-Elmer Gene Amp® PCR System 2400)

2. Load 1 μ l on a gel to check amplification products

¹ This amplification step with Pfu polymerase was incorporated for 2 reasons: (a) To increase DNA yields for the production of large numbers of transformants. (b) To reduce the amount of template DNA that is carried over from the mutagenic PCR reaction: (Primers for the second amplification reaction are nested within the mutagenic primers. The mutagenic primers were designed with non-specific tails of 11 and 12 bases respectively for the upstream and downstream primers. The nested primers will amplify DNA that was previously amplified with the mutagenic primers, but cannot amplify pRAM template DNA.)

3. Purify amplification reaction products with Wizard PCR purification kit (Promega Corporation, part#A718c):
 - transfer PCR reaction into a new tube containing 100 μ l Direct Purification buffer (Promega, Part#A724a)
 - 5 - add 1 ml of Wizard PCR Purification Resin (Promega Part#A718c) and incubate at room temperature for 1 min
 - pull resin though Wizard minicolumn
 - wash with 80% Ethanol
 - spin in microcentrifuge to remove excess Ethanol
 - 10 - elute with 88 μ l sterile nanopure water (allow water to remain on column for at least 1 min)

Subcloning of amplified PCR mutagenesis products

1. Digest the DNA with *Sfi* I as follows:
 - 2 μ l *Sfi* I (Promega Part #R639a)
 - 15 - 10 μ l 10X buffer B (Promega Part #R002a)
 - 88 μ l of DNA from Wizard PCR prep (see step 3 above)
 - mix components and overlay with 2 drops of mineral oil; incubate at 50°C for 1 hour
2. Remove salts and *Sfi* I ends with Wizard PCR purification as described herein, and elute into 50 μ l sterile nanopure water
- 20 3. Ligation into pRAM (+/r) backbone (set up 4 ligations per group):
 - 0.025 pmol pRAM backbone
 - 0.05 pmol insert (usually in the range of 6 to 12 μ l of insert)
 - 1 μ l of T4 DNA Ligase (Promega part M180a)
 - 25 - 2 μ l of 10X ligase buffer (Promega part C126b, divide into 25 μ l aliquots, do not freeze/thaw more than twice)
 - water to 20 μ l
 - ligate for 2 hours at room temperature
 - heat reactions for 15 minutes at 70°C to inactivate ligase

30 *Transformation and plating*

1. Butanol precipitate samples to remove excess salts (n-Butanol from Sigma, St. Louis, Missouri, part #BT-105):

(if ethanol precipitation is used instead of butanol, a wash with 70% ethanol as needed. Excess salt will cause arcing during the electroporation which causes the reaction to fail.)

5 - add water to 50 μ l
 - add 500 μ l of n-butanol
 - mix until butanol/ligation mix is clear and then spin for 20 min at room temperature
 - drain butanol into waste container in fume hood

10 - resuspend in 12 μ l water, spin 30 sec at full speed

2. Preparation of cell/DNA mix (set up 4 transformations plus one with reference clone DNA):
 - while DNA is precipitating, place electroporation cuvettes on ice
 - fill 15 ml Falcon snap-cap tubes with 3 ml S.O.C. medium and place on ice

15 - thaw JM109 electrocompetent cells on ice (50 μ l per ligation reaction)
 - pipette 10 μ l of the bottom layer from step 1 (or 0.5 μ l ref.clone DNA) into competent cells

20 (small amounts of butanol carry-over do not adversely effect the transformation efficiency)
 - place cell/DNA mix on ice

25 3. Electroporation:
 - carry tubes, cuvettes, and cell/DNA mix on ice to electroporation device
 - pipette cell-DNA mix into a cuvette and zap. Instrument settings:
 Cuvette gap: 0.2 cm
 Voltage: 2.5 kV

30 Capacitance: 25 μ F
 Resistance: 200 Ohms
 Time constant: 4.5 msec

- pipette 1 ml SOC (contains KCl; media prep #KCLM) into cuvette, quickly pour into recovery tube (transformation efficiency is reduced if cells are allowed to sit in cuvette)
- place the recovery tube on ice until all samples are processed
- 5 - allow the cells to recover at 37°C for 30-60 minutes
- plate on LB + amp plates with nitrocellulose filters (# of colonies is about 20% higher if cells recover 60 minutes, possibly due to cell replication.)
- (Best colony density for screening is 500 per plate. For the 10 current batch of cells plate about 500 to 750 µl)

H. Recombination Mutagenesis Protocol or DNA shuffling

DNase I digestion of plasmid DNA

- 1. Prepare 2% low melting point gel
 - 15 - use 0.8 g agarose in 40 ml (NuSieve #50082)
 - use large prep comb
 - make sure it is solidified prior to digesting
- 2. Prepare 4 µg of pooled plasmid DNA for digest
- 3. Prepare 1 U/µl DNase dilution on ice according to the table 20 below:

Table 8

DNase I ⁺	0.74 µl
10X DnaseI buffer	10 µl
1% gelatin*	10 µl
Water to 100 µl	

⁺ DNase I from Sigma (D5791)

* Gelatin was added to keep the DNase I from sticking to the walls of the tubes.

30 This dilution can be kept on ice for at least 30 min without loss in activity.

4. Digest (set up at room temperature):

35 prepare two digests with 1.0 U and 1.5 U DNase I per 100 µl reaction:

- 10 μ l of 10X DNase I buffer (500 mM Tris, 10 mM MgCl₂ pH 7.8)

7.8)

- x μ l DNA (2 μ g of pooled plasmid DNA from step 2)

- 1 or 1.5 μ l of the 1 U/ μ l enzyme dilution

5 - sterile nanopure water to 100 μ l

- incubate at room temperature for 10 minutes

- stop reaction by addition of 1 μ l of 100 mM CDTA

Purification from agarose gel

1. Run DNase digested fragments on gel

10 - add 10 μ l of 10X loading buffer to each DNase I digest

- load all on a 2% Low melting point agarose gel

- run about 30 min at 120-150 V

- load pGEM DNA marker in middle lane

2. Isolate fragments

15 - cut out agarose slice containing fragments in the size range of 600-1000 bp using a razor blade

- cut into pieces that weigh about 0.3 g

- melt the gel slices at 70°C

- add 300 μ l of Phenol (NaCl/Tris equilibrated) to the melted

20 agarose, vortex for about 1 minute at max speed

- spin for 10 min at 4°C

- remove the top layer into a tube containing an equal volume of Phenol/Chloroform/Isoamyl (saturated with 300 mM NaCl /100 mM Tris pH 8.0), vortex and centrifuge for 5 minutes

25 at RT

- remove the top layer into a tube containing chloroform and vortex and centrifuge.

- remove the top layer into a tube with 2 vol. of 95% cold Ethanol; place in -70°C freezer for 10 min (no additional

30 salts are needed because of the High Salt Phenol)

- spin at 4°C for 15 minutes.

- wash with 70% Ethanol, drain and air dry for ~10 min

- resuspend in 25 to 50 μ l of sterile nanopure water

- store at -70°C until ready for use

Assembly reaction

Set up 4 reactions (see Table 9) and pool when completed.

5

Table 9

	Component	Concentration	Amount in μ l	Final concentration
	dATP	10 mM	1	200 μ M
	dCTP	10 mM	1	200 μ M
	dGTP	10 mM	1	200 μ M
10	dTTP	10 mM	1	200 μ M
	DNA*	1-10 ng	5	
	Tli	3 U/ μ l	0.4	0.24 U/ μ l
15	10X Thermo buffer	10X	5	1X
	MgCl ₂	25 mM	4	2 mM
	gelatin	1%	5	0.1%
	water		To 50 μ l	

* Because the DNA used for this reaction has been fragmented, it is difficult to estimate a concentration. The easiest way is to load 5 μ l of the DNaseI digested DNA to an agarose gel and run the gel until the dye enters the wells (1-2 min). Fragments from a typical 2 μ g DNA digest which were resuspended in 100 μ l of water give a DNA concentration of about 1 to 10 ng/ μ l.

Cycling conditions: 94°C for 30 seconds (94°C for 20 seconds, 65°C for 25 minutes, 72°C for 2 minutes) 25X

Amplification of assembly

Usually 5 amplification reactions (see Table 10) will produce enough DNA for a full 8 plate robotic run.

Table 10

Component	Concentration	Amount in μ l	Final concentration
dATP	10 mM	1	200 μ M
dCTP	10 mM	1	200 μ M
dGTP	10 mM	1	200 μ M
dTTP	10 mM	1	200 μ M
pRAMtailUP*	20 pmol/ μ l	2	0.8 pmol/ μ l
pRAMtailDN*	20 pmol/ μ l	2	0.8 pmol/ μ l
Pfu native polymerase ⁺	2 U/ μ l	1	0.04 U/ μ l
10X native Pfu buffer ^o	1X	5	1X
DNA	1-10 ng	5	
water		water to 50 μ l	

* Note that the concentration of primers is twice as high as in a typical amplification reaction.

^o The Pfu 10X buffer contains 20 mM MgCl₂, so it is not necessary to add MgCl₂.

⁺ Pfu polymerase is ordered from Stratagene part #600135. Cycling conditions: 94°C for 30 seconds (94°C for 20 seconds, 65°C for 1 minute, 72°C for 3 minutes) 25x

Subcloning of assembly amplification

Purify amplification products with Wizard PCR purification:

- pool 5 amplification reactions
- transfer into a new tube that contains 100 μ l of Direct Purification buffer
- add 1 ml of Wizard PCR Purification Resin, incubate at RT for 1 minute
- pull Resin though Wizard minicolumn
- wash with 80% ethanol and spin in microcentrifuge to remove excess ethanol
- elute with 88 μ l of sterile nanopure water (allow water to remain on column for at least 1 minute)

2. Digest with *Sfi* I:

- 2 μ l *Sfi* I
- 10 μ l 10X buffer B
- 88 μ l of DNA from Wizard PCR prep

- mix components and overlay with 2 drops of mineral oil;
incubate at 50°C for 1 hour

3. Band isolation:
Sometimes after amplification of the assembly reaction a
5 band that is smaller than the gene-sized fragment is produced.
This small fragment has been shown to subclone about 10-fold
more frequently than the gene sized fragment if the sample is not
band isolated. When this contaminating band is present, it is
necessary to band isolate after *Sfi* I digestion.

10 - load the DNA to a 0.7% agarose gel
- band isolate and purify with the Gene Clean kit from Bio 101
- elute DNA with 50 µl sterile nanopure water, check
concentration on gel (This type of purification with
standard agarose produced the highest number of
15 transformants after subcloning. Other methods tried: Low
melt with Phenol chloroform, Gene clean with low melt,
Wizard PCR resin with standard agarose, Pierce Xtreme
spin column with Low melt (did not work with standard
agarose)).

20 4. Ligate into pRAM [+/r] backbone: (See ligation and
transformation protocol above)

Large scale preparation of pRAM backbone

1. Streak an LB amp plate with pRAMMCS [+/r] (This vector
25 contains a synthetic insert with a *Sac* II site in place of a gene.
This vector contains the new ribosome binding site, but it will be
cut out when the vector is digested with *Sfi* I.

2. Prepare a 10 ml overnight culture in LB supplemented with amp.
3. The next day inoculate 1 L of LB supplemented with amp and
30 grow for 16-20 hours.

4. Purify the DNA with the Wizard Maxi Prep kit. (Promega
#A7270) (use 4 preps for 1 L of cells)

5. Digest the Plasmid with *Sfi* I. (Use 5 U per microgram) Overlay with mineral oil and digest for at least two hours.
6. Ethanol precipitate to remove salts. Resuspend in water.
7. Digest with *Sac* II for 2 hours. (Keep digest volume to 2 ml or less). It is possible that part of the plasmid could be partially digested. If the vector is cut with an enzyme that is internal to the two *Sfi* I sites, it will keep the partially digested fragments from joining in a ligation reaction.
8. Load entire digest onto a column (see 9). The volume of the sample load should not be more than 2 ml. If it is it will be necessary to ethanol precipitate.
9. The column contains Sephadryl S-1000 and is stored with 20% ethanol to prevent bacterial contamination. Prior to loading the sample the column must be equilibrated with cold running buffer for at least 24 hours. If the column has been sitting more than a couple of months it may be necessary to empty the column, equilibrate the resin 3-4 washes in cold running buffer, and then re-pour the column. After the column is poured it should be equilibrated overnight so that the resin is completely packed.
10. Collect fractions of about 0.5 ml. Typically the DNA comes off between fractions 25 and 50. Load a 5 μ l aliquot from a range of fractions to determine which fractions contain the backbone fragment. The small insert fragment will start to come off the column before all of the backbone is eluted, so it will be necessary to be conservative when fractions are pooled. For this reason typically 40-60% of the DNA is lost at this step.
11. Pool the fractions that contain the backbone.
12. Ethanol precipitate the samples. Resuspend in a volume that produces about 10-50 ng/ μ l.

13. Store at -70°C.

Column running buffer: (store at 4°C)

5 mM EDTA

5 100 mM NaCl

50 mM Tris-HCL pH 8.0

10 $10 \mu\text{g/ml}$ tRNA (R-8759, Sigma)

I. Oligonucleotide Mutagenesis

10 Prepare Ampicillin-sensitive single stranded DNA of the template to be mutated. Design a mutagenic primer that will randomly generate all possible amino acid codons.

Mutagenesis reaction:

15

Table 11

Component	Final concentration
Single Stranded Template	0.05 pmol
Mutagenic Oligonucleotide	1.25 pmol
Ampicillin Repair Oligo (Promega q631a)	0.25 pmol
20 10X annealing buffer*	1X
Water to 20 μl	

*10X Annealing buffer:

-200 mM Tris-HCl, pH 7.5

25 -100 mM MgCl₂

-500 mM NaCl

Heat reaction at 60°C for 15 minutes and then immediately place on ice.

Synthesis reaction:

Table 12

Component	Amount
Water	5 µl
10X synthesis buffer*	3 µl
T4 DNA Polymerase (Promega m421a)	1 µl (10 Units)
T4 DNA Ligase (Promega 180a)	1 µl (3 Units)

10 *10X Synthesis buffer

100 mM Tris-HCl, pH 7.5

5 mM dNTPs

10 mM ATP

20 mM DTT

15 Incubate at 37°C for 90 minutes.

Transform into Mut-S strain BMH 71-18 (Promega strain Q6321)

-Place Synthesis reaction in a 17X100 mm tube.

-Add BMH 71-18 competent cells that have been thawed on ice to synthesis reaction.

20 -Incubate on ice for 30 min

-Heat Shock cells at 42°C for 90 seconds.

-Add 4 ml of LB medium and grow cells at 37°C for 1 hour. Add Ampicillin to a final concentration of 1.25 ug/ml and then grow overnight at 37°C.

25 Isolate DNA with Wizard Plus Purification system (Promega a7100)

Transform isolated DNA into JM109 electrocompetent cells and transform onto LB Ampicillin plates.

J. Screening procedure

30 JM109 clones (from a transformation reaction) are plated onto nitrocellulose filters placed on LB amp plates at a screening density of about 500 colonies per plate.

As listed in the Random Mutagenesis procedure, approximately 10% of the clones to be selected will have to be as stable as the same sequenced or better than source. Or stated another way, about 50 colonies per plate will be suitable for selection. There are 704 wells available for a full eight plate robotic run, so 5 at least 15 LB amp plates will be needed for a full robotic run.

After overnight growth at 37°C the plates containing the transformants are removed from the incubator and placed at room temperature.

The nitrocellulose filter is lifted on one side and 500 μ l of 10 mM IPTG is added to each of the plates. The filter is then placed back onto the plate to 10 allow diffusion of the IPTG into the colonies containing the different mutant luciferase genes. The plates are then incubated for about 4 hours at room temperature.

One (1) ml of a solution contains 1 mM luciferin and 100 mM sodium citrate is pipetted onto a slide warmer that is set at 50°C. A nitrocellulose filter 15 that contains mutant luciferase colonies and has been treated with IPTG is then placed on top of the luciferin solution. After several minutes, the brightest colonies are picked with tooth picks which are used to inoculate wells in a microtiter plate that contain M9- minimal media with 1% gelatin.

After enough colonies are picked to 8 microtiter plates, the plates are 20 placed in an incubator at 350 rpm at 30°C incubation and are grown overnight.

In the morning the overnight plates are loaded onto the robot and the cell dilution procedure is run. (This procedure dilutes the cultures 1:10 into induction medium). The new plates are grown for 3 hours at 350 rpm at 30°C.

After growth, the plates are loaded to the robot for the main assay 25 procedure.

Minimal Media:

6 g/Liter Na_2HPO_4
3 g/Liter KH_2PO_4
0.5 g/Liter NaCl
30 1 g/Liter NH_4Cl
2 mM MgSO_4
0.1 mM
1 mM Thiamine-HCl

0.2% glucose
12 µg/ml tetracycline
100 µg/ml ampicillin

5 *Overnight media contains 1% gelatin

*Induction media contains 1 mM IPTG and no gelatin.

S.O.C. Media:

10 -10 mM NaCl
-2.5 mM KC1
-20 mM MgCl₂
-20 mM glucose
-2% bactotryptone
-0.5% yeast extract

15 Summary of Exemplary Evolutionary Progression

1. Start with *LucPpe2*[T249M]
2. Mutate 3 amino acids at N- and C-termini
3. Mutate 7 cysteines
4. Perform two iterations of evolution → *Luc49-7C6*
- 20 5. Mutagenesis of altered codons (9)
6. Two iterations of evolution → *Luc78-0B10*
7. Mutagenesis of consensus codons (28)
8. Mutagenesis of codon usage (24) → *Luc90-1B5*

25 One Iteration of Recursive Process

1. 1 clone → 3 libraries using error-prone PCR
 - 3 x Visual screen (about 10,000 clones each)
 - 3 x Quantitative screen (704) clones each)
2. 3 x 18 clones → library using sPCR
 - Visual screen (about 10,000 clones)
 - Quantitative screen (704 clones)
- 30 3. 18 + 18 → library using sPCR
 - Visual screen (about 10,000 clones)

- Quantitative screen (704 clones)

4. Output: 18 clones

5

EXAMPLE 5

Mutagenesis Strategy from Clone Luc90-1B5 to Luc133-1B2 and Luc146-1H2

Upon storage, luciferin degrades and the degradation products inhibit luciferase. The production of inhibitors causes an apparent instability in the 10 reagent containing both luciferase and luciferin. There are two ways to reduce this problem: 1) Store the luciferin and luciferase at pH 5.5-6.0 to reduce the rate of luciferin degradation, and/or 2) Evolve an enzyme that is resistant to the luciferin degradation products.

LucPpe2 mutants that were evolved after clone Luc90-1B5 were evolved 15 to be more stable at low pH and have resistance to luciferin degradation products. These mutant enzymes are useful, for example, in an ATP detection kit. One embodiment of such a kit comprises a mixture of luciferin and luciferase. A luminescent reaction occurs when a sample comprising ATP is added to the mixture.

20 Three populations of random mutants were produced using clone Luc90-1B5 as a template. These three populations were screened on the robot as experiments 114, 115, and 117. Robotic screens for experiments 114, 115, 116, 117, 118, 119, and 122 were completed as described previously except that buffer C was prepared with citrate buffer pH 4.5 instead of HEPES buffer pH 25 7.8, and the assay reagent was prepared with HEPES pH 7.1 with 10 μ M ATP instead of Tricine pH 8.0 and 175 μ M ATP. These screening conditions were biased to select clones that have increased retention of luminescence activity over time at pH 4.5 at 48°C and increased luminescence activity when assayed at pH 7.1 with 10 μ M ATP. Seventeen clones from experiment 114, seven clones 30 from experiment 115, and ten clones from experiment 116 were shuffled together using sPCR and selected mutants from this screen were run on the robot as experiment 117. Eighteen clones were selected from experiment 117.

The clone that was determined to have the most improved characteristics (increased retention of luminescence activity over time at pH 4.5 and 48°C and increased luminescence activity when assayed at pH 7.1 with 10 μ M ATP) was clone Luc117-3C1 and it was selected as a template for random mutagenesis.

5 Two populations of random mutants were screened and then run on the robot as experiments 118 and 119. Seven clones from experiment 118 and five clones from experiment 119 were saved.

Clones from experiments 114, 115, 116, 117, 118, and 119 were selected based upon the following characteristics: brighter luminescence than Luc90-1B5, 10 and increased retention of luminescence activity over time at pH 4.5. These select clones were shuffled together and were run on the robot as experiment 122. Eleven clones from this experiment were saved.

15 Three populations of random mutants were prepared from clone Luc122-4D5 and run on the robot as experiments 125, 126, and 127. Thirteen clones from experiment 125, four clones from experiment 126, and three clones from experiment 127 were shuffled together and run on the robot as experiment 128. For experiments 125, 126, 127 and 128 the screen for K_m was altered to select for clones that are more resistant to luciferin degradation products. The clones were also screened for retention of luminescence over time at pH 4.5.

20 Instead of screening for substrate utilization, a screen for resistance to inhibitor was conducted. In place of the 0.06X dilution of substrates, a 75:25 mix of D to L luciferin in 1X assay buffer was used and designated as "0.75X". In place of the 0.02X dilution of substrates, a 50:50 mix of D to L luciferin in 1X assay buffer and was designated as "0.5X". The 1X assay buffer in these 25 experiments contained the following: 10 μ M ATP, 50 mM HEPES pH 7.8, 8 mM MgSO₄, and 0.1 mM EDTA. The 0.75X sample contained 75 μ M D-luciferin and 25 μ M L-luciferin. The 0.5X sample contained 50 μ M D-luciferin and 50 μ M L-luciferin. The 1X sample contained 250 μ M D-luciferin. A K_m regression was used as before and a K_m value was calculated. Normalized values 30 of greater than 1 indicate more resistance to inhibitor. Clones from these experiments that were shown to have greater resistance to L-luciferin were also more resistant to luciferin degradation products.

To more easily measure resistance to inhibitor on the robotic system, a new variable "Q" was designated. The "Q" variable replaces the K_m variable used previously. The luminescence ratio is calculated the same as in the K_m measurement, then the natural log (ln) of each luminescence ratio is calculated 5 (Y-axis). The X-axis is an arbitrary time that is entered by the user. The first time point is zero and the samples are measured with 1X assay buffer that contains

250 μ M D-luciferin. The next two time points have the same time value (i.e., 4 hours to simulate incubation of luciferin) and samples are measured with 1X 10 assay buffer that contains a 50:50 mixture (as described above) of D-luciferin to L-luciferin. A linear regression correlating ln(lum ratio) to time is calculated. Q is calculated as the ln(0.5)/slope. Normalized values of "Q" greater than 1 indicate more resistance to inhibitor. Experiments 133 and higher were run using this program.

15 Sixteen clones from experiment 128 were shuffled with clones from experiment 122 and run on the robot as experiment 133. Two samples, Luc133-1B2 and Luc133-0D11, were selected as templates for random mutagenesis and run on the robot as experiments 145 and 146, respectively. The clone that showed an increased retention of luminescence over time at pH 4.5 and the most 20 resistance to inhibitor was clone Luc146-1H2. Moreover, at pH 4.5 and 48°C, Luc133-1B2 and Luc146-1H2 had increased thermostability relative to Luc90-1B5, and increased resistance to inhibitor (Figures 54-61). A comparison of the luminescence signal for Luc49-7C6, Luc78-0B10, Luc90-1B5, Luc133-1B2, and Luc146-1H2 is shown in Figure 59. A comparison of the thermostability at 25 50°C for clones for Luc49-7C6, Luc78-0B10, Luc90-1B5, Luc133-1B2, and Luc146-1H2 is shown in Figure 60. Figures 55-58 show the nucleotide sequence encoding and the inferred amino acid sequence of Luc133-1B2 and Luc146-1H2.

Materials and Methods

Assay to detect resistance to luciferase inhibitor

30 A 10 mM stock solution of luciferin is incubated at 50°C in 50 mM HEPES, pH 7.8, to accelerate the production of luciferin breakdown products. At different time points an aliquot is removed and then placed at -20°C. After incubation is complete, assay reagent (100 μ M Luciferin, 1 μ M ATP, 50 mM

HEPES, pH 7.8 and 8 mM MgSO₄) is prepared with luciferin from each of the different time points and a diluted lysate is then assayed with each assay reagent.

The lysate is prepared as follows. Overnight cultures of clones to be tested are prepared in LB supplemented with 100 µg/ml AMP. The cultures are 5 diluted 1:10 in M-9 minimal media supplemented with 1 mM IPTG, 100 µg/ml AMP and grown for 3 hours at 30°C. Forty-five µl of cells is mixed with 20 µl of Buffer A and frozen. The mixture is thawed, 175 µl of Buffer B added, and the resulting mixture diluted 1:10 in Buffer C. A regression of luminescence versus time of luciferin incubation is then calculated, and from this graph half-life is extrapolated. A longer half-life means that the mutant being tested is more 10 resistant to luciferin breakdown products.

EXAMPLE 6

Mutagenesis Strategy from LucPplYG to Clone Luc81-6G01

15 The luciferases from the luminous beetle, *Pyrophorus plagiophthalmus*, had been shown previously to generate different colors of luminescence (LucPpl). Analysis of these luciferases revealed that the different colors were caused by discreet amino acid substitutions to their protein sequences. This allowed the possibility to make a pair of genetic reporters capable of emitting a 20 multiplexed luminescent signal, thus enabling quantitation of two biomolecular events simultaneously from within the same living system.

Amino acid substituted LucPpl were prepared which have the following properties:

Physical stability of the luciferases

25 Although the luminescence activity of LucPpl within colonies of *E. coli* appeared to be thermostable to above 60°C, in lysates these luciferases had relatively low stability. They were particularly unstable in the presence of Triton X-100 detergent. When lysates are prepared containing the commonly used firefly luciferase, the enzyme retains greater than 90% activity over 5 hours at 30 room temperature. In contrast, the activity of the LucPpl luciferases would decrease several fold over the same period.

The thermostabilities of the LucPpl luciferases are also near the physiological temperature of mammalian cells. The green-emitting luciferase

(*LucPp1/GR*) and red-emitting luciferase (*LucPp1/RD*) have different thermostabilities which may cause differences in the behaviors as genetic reporters within cells. The influence of temperature should be greatest near the point of denaturation for the enzymes, where small changes in temperature will

5 have the greatest effect on protein structure. In contrast, temperature will have much less affect on protein structure when it is much below the denaturation point. Thus, the differential effect on two enzymes having slightly different denaturation temperatures will be less at relatively lower temperatures. It might therefore be preferable to have the denaturation temperature of the reporter

10 enzyme significantly above the growth temperature of mammalian cells.

Spectral overlap between the luciferases

Although a method was developed to quantify each luciferase in a mixture by using colored filters, the ability to discriminate between the luciferases is limited by their spectral overlap. This overlap reduces the ability

15 to accurately measure both luciferases if their luminescence intensities differ by more than 10 fold. If the intensities differ by more than 50 fold, the luminescence signal of the dimmer luciferase is obscured by the other. Thus, it would be preferable to further separate the luminescence spectra of the two luciferases.

20 Many different mutations were identified which shifted the luminescence spectrum towards the red. But the limit for red luminescence appears to be about 620 nm. Further effort at shifting the spectrum of the red-emitting luciferase into still longer wavelengths might have some benefit. It was found that green-shifting mutations were rare, however, an extensive analysis was not conducted.

25 Measurements from native luciferases show some examples of luminescence below 530 nm, about 15 nm less than the green-emitting prototype enzyme.

Differential physical and enzymological characteristics

Ideally the two luciferase reporters would be identical in all characteristics except for the color of luminescence. However, as noted above,

30 the physical stability of the luciferases was not identical. It was also found that mutations resulting in red-shifted luminescence also caused an increase in the K_M for luciferin. Although some of these differences may be unavoidable, it is not clear whether the properties are fundamentally associated. For instance,

luciferases from different beetle species sometimes have significantly differing K_m even though their luminescence spectra are similar. It may be that much of the differences associated with development of the red-emitting luciferase are due to concomitant perturbations to the integrity of the enzyme structure, as the 5 thermostability of a prototype of the red-emitting luciferase was increased without significantly altering the luminescence spectrum.

Stable luminescence signals

When firefly luciferase was first described as a genetic reporter, the luminescent signal was a relatively brief flash initiated upon injection of the 10 reaction substrates. Subsequent development of the luminescent chemistry made the assay more convenient by enabling a stable signal for several minutes. Presently, such stabilized assays are standard for general laboratory applications. However, to allow high throughput screening in pharmaceutical research, the luminescence signal was further stabilized to extend for over an hour. This was 15 necessary to allow sufficient time to assay several thousand samples in a batch. Although the luminescent signal of the new multiplexed luciferases was stable for minutes, they did not provide the extended signal stability needed for high throughput screening. It would be preferable if the signal stability could be further increased while optimizing other properties.

20 Methods to Optimize Luciferase Performance

To prepare luciferases having certain performance, a method for *in vitro* evolution of enzyme function, as described above, was employed. Briefly described, the method is a recursive process of generating random mutations and screening for desirable properties. It was originally developed primarily to 25 increase the thermostability of luciferases, although other enzymological characteristics are also subject to optimization by the screening criteria. A slightly different strategy was used to achieve the properties described above since two related luciferases needed to be optimized concomitantly.

Initially, a single prototype enzyme is subjected to *in vitro* evolution to 30 optimize physical stability and the luminescence signal. In the process, the mutant libraries are also screened for any new mutations causing changes in color. Particular emphasis is placed on isolating green shifted mutants. After initial optimization of a common prototype, a green- and red-emitting form of

the enzymes is created, and these are further optimized separately to harmonize their physical and chemical properties. Particular attention is given to matching their physical stabilities and their substrate binding constants, especially for luciferin.

5 The choice for the initial prototype for optimization was the wild-type yellow-green-emitting luciferase isolated from the luminous beetles (*LucPp/YG*). Of the luciferases originally cloned from *P. plagiophthalmus*, this one produced the brightest luminescence when expressed in *E. coli*. Furthermore, there was concern that the lack of green-shifting mutation resulted
10 because the prior mutagenesis studies were done using a luciferase that already had the greenest luminescence. It was possible that if additional green-shifting mutations existed, they might be more evident when screened in a red-shifted background. The mutagenesis was performed as follows:

Remove peroxisomal targeting sequence

15 The translocation signal at the C-terminus of the luciferases was removed. This was done using oligonucleotide-directed mutagenesis to convert the normal -KSKL to -XXX* (where X represents any amino acid, and * represents a termination codon). Several colonies yielding bright luminescence were selected and used as templates for the next stage of mutagenesis.

20 Removal of sensitive cysteines

The luciferases from *P. plagiophthalmus* have 13 cysteines, which are potentially sensitive to oxidation. This is in contrast to the commonly used firefly luciferase, which has only 4 cysteines. To remove any cysteines that may limit enzyme stability, oligonucleotide-directed mutagenesis was used to
25 randomize the cysteine codons. Three sets of oligonucleotides were used: non-conserved cysteines in regions of low sequence homology (positions 69, 114, 160, 194, 335, and 460), non-conserved cysteines in regions of higher sequence homology (positions 127, 213, 310, and 311), and highly conserved cysteines (positions 60, 80, and 388). The best clones from each of these screens were
30 isolated, and a new mutant library made by sPCR and screened again. At the screening temperature of 29°C, the activity of the wild-type yellow-green-emitting luciferase decreased about 500-fold over 10 hours. The activity of the most stable mutant (Luc20-4C10) was more stable, decreasing only about 2 fold.

First cycle of random mutagenesis

Using the procedure developed previously, three mutant libraries were generated using error-prone PCR and screened. The best mutants from these were recombined into a new library by sPCR and screened again. Finally, the 5 best clones of this screen were recombined with the best clones from the previous oligonucleotide-directed mutagenesis by sPCR, and screened again. At 41°C, the activity of the best mutant from this process (Luc30-4B02) decreased 63 fold over 10 hours, whereas the activity of the parent mutant (Luc20-4C10) decreased greater than 100,000 fold.

10 Sequence analysis

Six of the best mutants from the last screen were isolated and sequenced. This revealed that the amino acids at 16 positions had been changed among the six clones. Thirteen positions had been changed in the preferred mutant, Luc30-4B02. Four of the changes were at the C-terminus in all the isolated mutants, 15 where oligonucleotide mutagenesis had changed the wild-type sequence of -KSKL to -AGG*. Only two of the cysteines had been changed by the previous oligonucleotide mutagenesis; one highly conserved cysteine at position 60 was changed to valine and one moderately conserved cysteine at position 127 was changed to threonine. The remaining amino acid changes were all due to point 20 mutations in the DNA, consistent with error-prone PCR. Interestingly, three of these changed the amino acid into that found in the wild-type green-emitting luciferase (two in mutant Luc30-4B02). Four of the remaining changes brought the mutant sequences closer to the consensus amino acid among other cloned beetle luciferases (two in mutant Luc30-4B02) (Figure 19B). An additional 4 25 codons were changed without affecting the amino acid sequence.

Site-directed mutagenesis

To further explore the potential of the mutations identified in the sequenced mutants, additional mutagenesis experiments were performed using oligonucleotides. Eight of the codons mutated by the error-prone PCR were 30 randomized or partially randomized using oligonucleotide-directed mutagenesis. Four of the remaining cysteine codons were randomized; two highly conserved cysteines (positions 80 and 388) and two cysteines in a region of sequence homology (positions 310 and 311). One leucine was mutated to a

leucine/proline; proline is the consensus amino acid among other beetle luciferases.

The mutagenesis was performed with four sets of oligonucleotides (Table 13), and the best clones from each set were selected. These were 5 recombined by sPCR together with the selected clones from the previous random mutagenesis and screened again. The activity of the best clone from this process (Luc47-7A11) decreased 2.3 fold at 42°C; the activity of the parent clone (Luc30-4B02) decreased greater than 2000 fold.

10

Table 13

Experiment	Mutations
Set A	$C_{80} \rightarrow X + K_{84} \rightarrow X + I_{91} \rightarrow (F, L, I, M, V, S, P, T, A)$
Set B	$I_{288} \rightarrow (F, L, I, M, V, S, P, T, A)$ $C_{310} \rightarrow X + C_{311} \rightarrow X$
Set C	$G_{351} \rightarrow (I, M, V, T, A, N, K, D, E, S, R, G) + L_{350} \rightarrow (L, P) + S_{356} \rightarrow P + L_{359} \rightarrow (F, L, I, M, V, S, P, T, A)$
Set D	$C_{388} \rightarrow X + V_{389} \rightarrow (N, Y, N)$ $K_{457} \rightarrow X$

Second cycle of random mutagenesis

The random mutagenesis process using error-prone PCR was applied 20 again to the best clone from the oligonucleotide-directed mutagenesis (Luc47-7A11). Three libraries were again created and screened, and selected mutants were recombined by sPCR and screened again. Following recombination, the activity of the best mutant (Luc53-0G01) decreased 1.2 fold at 43°C. The parent clone (Luc47-7A11) decreased 150 fold. After recombining the best of these 25 new mutants with the best mutants from the previous oligonucleotide-directed mutagenesis, the activity of the new best mutant (Luc55-2E09) decreased 31 fold at 47°C, compared to 80 fold for the parent (Luc53-0G01).

Third cycle of random mutagenesis

The random mutagenesis process was repeated using the best clone from 30 the previous cycle of mutagenesis (Luc53-0G01). After recombining the selected mutants with the mutants from the second cycle of mutagenesis, the activity of the best clone (Luc81-6G01) decreased 100-fold at 47°C, compared with 750 fold for the parent (Luc53-0G01). The discrepancy in measured activity of Luc53-0G01 in this cycle of mutagenesis compared to the previous

cycle may be due to changes in the assay procedure and recalibration of the incubator temperature. It should be noted that the recorded thermostabilities from each stage of mutagenesis are calculated from robotic data using abbreviated assay procedures. The data are intended to indicate the relative 5 stabilities of enzyme mutants when assayed in the same screen, rather than providing an accurate quantitation of thermostability.

Luminescence

Before making a final selection of the best clone from which to create the green- and red-emitting luciferases, further analysis was done on the best clones 10 from the final screen. Three clones in particular were strong candidates as the final choice: Luc81-0B11, Luc81-5F01, and Luc81-6G01. The luminescence properties of these three mutant enzymes were compared among one another. They were also compared to the wild-type yellow-green-emitting luciferase to gauge the effect of the *in vitro* evolution process.

15 From colonies of *E. coli* expressing the luciferases, Luc81-5F01 and Luc81-6G01 produced luminescence most rapidly at room temperature upon addition of luciferin. The luminescence was more rapid and brighter than colonies expressing the wild-type green- and yellow-green-emitting luciferases. The luminescence from all the selected colonies appeared green-shifted 20 compared to the yellow-green parent clone. When the colonies are heated to 65°C, the yellow-green clone loses most luminescence and the green clone becomes dimmer. Some of the mutant clones lose their luminescence at 65°C, but the three preferred clones remain bright above 70°C. No spectral changes upon heating the colonies were evident until above 70°C, where those clones still 25 retaining activity began to red-shift slightly (sometime, the initial phases of enzyme denaturation are accompanied by a red shift in the luminescence). The luminescence characteristics of the three preferred mutants are quite similar.

The thermostability of the mutant luciferases in cell lysates was compared at room temperature (Figure 63). Dilute lysates were buffered at pH 30 7.5 and contained 1% Triton X-100; typical conditions for lysates of mammalian cells. The luminescence activity of all three mutant enzymes showed no decrease

over 20 hours, whereas the activity of the wild-type yellow-green-emitting luciferase decreased substantially.

For luminescence assays requiring only a few second, the wild-type yellow-green-emitting luciferase produces a very stable signal (the initial rise in 5 the signal evident in the first 2 seconds is due to the response time of the luminometer, not the kinetics of the luminescence reaction) (Figure 64A). However, the signal intensity was reduced about 30% by the presence of 1% Triton X-100 in the lysate (diluted 1:5 with the addition of assay reagent). In contrast, the luminescence intensity of the mutant luciferases was unaffected by 10 the presence of Triton X-100. Under these conditions, the most stable signal was produced by Luc81-6G01, although the signal intensity was somewhat brighter for Luc81-5F01. However, the data are not corrected for the efficiency of enzyme expression in *E. coli*. Thus, differences in luminescence intensity may not correlate to changes in enzyme specific activity, nor is the expression 15 efficiency in *E. coli* necessarily relevant to expression in mammalian cells.

For batches of assays requiring more than an hour to process, the signal stability of the yellow-green-emitting luciferase is inadequate under the conditions tested. The luminescence intensity decreases several fold per hour (Figure 64B). Attempts to correct this by the *in vitro* evolution yielded mixed 20 results. The signal stability of all three mutant enzymes was generally much improved over the parent yellow-green enzyme for three hours after substrate addition. However, this was accompanied by a greater initial decrease in luminescence during the first half-hour. This initial decrease would be more acceptable if it had occurred more rapidly, so that batch processing of samples 25 would not be delayed by 30 minutes in waiting for the signal to stabilize. It may be possible to improve this kinetic behavior by adjusting the assay conditions.

From these results, the mutant Luc81-6G01 was chosen as the best clone from which to subsequently create the green- and red-emitting luciferases. The sequence of Luc81-6G01 (Figures 46-47) and Luc81-0B11 was determined and 30 compared with the sequences of Luc30-4B02 from earlier in the *in vitro* evolution process, and the wild-type yellow-green-emitting luciferase used as the initial parent clone (Figure 19B). Relative to Luc30-4B02, the Luc81-6G01 mutant acquired new mutations in 9 codons, of which 8 caused changes in the

amino acid sequence. Four of these 8 amino acid changes were probably acquired through recombination with clones generated prior to isolation of Luc30-4B02. Two are identical to mutations found in the other clones sequenced along with Luc30-4B02, and two are reversions to the wild-type 5 parent sequence. The remaining four are novel in the sequence of Luc81-6G01. Two of the novel mutations change the amino acid to the consensus amino acid among other cloned luciferases.

Interestingly, in either the sequences of Luc81-6G01 or Luc81-0B11, there is no evidence that the prior oligonucleotide-directed mutagenesis had any 10 beneficial effect. No novel nucleotide sequences appear at any of the targeted codons. The improved enzyme performance following the oligonucleotide-directed mutagenesis apparently was due to recombination of previously acquired mutations. All of the novel amino acid changes in Luc81-6G01 and Luc81-0B11 are at sites not targeted by the oligonucleotides and are due to 15 single-base modifications of the codons, consistent with error-prone PCR. Even though the novel mutations in Luc81-6G01 were not found in the earlier sequence data, it is not certain when they were generated in the process. Most likely they were produced in the second and third cycles of random mutagenesis; however, they may have been present among other selected mutants prior to 20 Luc30-4B02. Relative to the initial yellow-green-emitting luciferase, the Luc81-6G01 mutant has acquired 17 amino acid changes and 3 codon mutations not affecting the amino acid sequence.

The observation that the onset of luminescence within colonies of *E. coli* is faster for the new mutants, and that the luminescence is brighter at higher 25 temperatures, is probably not due to differences in protein expression. Immunoblot analysis of cell expressing the different luciferases showed no significant differences in the amount of polypeptide present. As noted above, the greater light intensity at higher temperatures is due to the increased thermostability of the mutant luciferases. The apparent K_M 's for ATP and 30 luciferin have also changed during the course of the *in vitro* evolution (Table 14). To estimate the K_M values, the mutant luciferases were partially purified from lysates of *E. coli* by differential precipitation using ammonium sulfate (40-

65% saturation fraction). The results show that the K_M 's for both ATP and luciferase are more than 10-fold lower.

When luciferin is added to an *E. coli* colony expressing luciferase, the intracellular concentration of luciferin slowly increases as it diffuses across the cell membrane. Thus, the intracellular concentration of luciferin reaches saturation sooner for those luciferases having the lowest K_M 's. Hence, the mutant luciferases appear brighter sooner than the wild-type parent clone. This also explains why the luminescence of the red-emitting prototype clone appears in *E. coli* colonies much more slowly than the green-emitting luciferase.

10 Analysis of K_M shows that the mutations causing the red luminescence also substantially increase the K_M for luciferin.

Table 14

Luciferase	K_M for ATP (μM)	K_M for luciferin (μM)
YG w.t.	140	21
Luc30-4B02	12	7.8
Luc81-6G01	8.0	1.9

15 From the analysis of luminescence signal *in vitro*, the luminescence from the mutant luciferases might be expected to fade more quickly than the wild-type luciferase during the first 30 minutes. Following this, the luminescence should be most stable in the mutants. However, this has not been noticed in the colonies of *E. coli*, and it may be that the kinetics of luminescence are different within cells compared to diluted enzyme in buffer.

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All publications, patents and patent applications are incorporated herein by reference. While in the foregoing specification, this invention has been described in relation to certain preferred embodiments thereof, and many details 20 have been set forth for purposes of illustration, it will be apparent to those skilled in the art that the invention is susceptible to additional embodiments and that certain of the details herein may be varied considerably without departing from the basic principles of the invention.

WHAT IS CLAIMED IS:

1. A second luciferase that has enhanced resistance to an inhibitor of the luciferase relative to a first reference beetle luciferase.
2. The second luciferase of claim 1 which retains at least 50% more activity in the presence of the inhibitor relative to the reference beetle luciferase.
3. The second luciferase of claim 1 which retains at least 100% more activity in the presence of the inhibitor relative to the reference beetle luciferase.
4. The second luciferase of claim 1 which comprises a plurality of amino acid substitutions relative to the reference beetle luciferase.
5. The second luciferase of claim 4 wherein the reference luciferase is native beetle luciferase.
6. The second luciferase of claim 5 wherein the reference beetle luciferase is *LucPpl*.
7. The second luciferase of claim 5 wherein the reference beetle luciferase is *LucPpe2*.
8. The second luciferase of claim 4 wherein the substitutions are to a consensus amino acid.
9. A luciferase which comprises SEQ ID NO:14, SEQ ID NO:19, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:47, or an enzymologically active portion thereof.
10. An isolated and purified nucleic acid molecule comprising a nucleic acid segment encoding the luciferase of claim 9 or the complement thereof.

11. The isolated and purified nucleic acid molecule of claim 10 comprising a nucleic acid segment comprising SEQ ID NO:1, SEQ ID NO:6, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:42, SEQ ID NO:43 or SEQ ID NO:46.
12. An isolated and purified nucleic acid molecule comprising a nucleic acid segment encoding the second luciferase of claim 1 or the complement thereof.
13. A vector containing the nucleic acid molecule of claim 10.
14. A vector containing the nucleic acid molecule of claim 12.
15. A host cell, the genome of which is augmented with the nucleic acid molecule of claim 10.
16. A host cell, the genome of which is augmented with the nucleic acid molecule of claim 12.
17. A solid substrate comprising the second luciferase of claim 1.
18. A solid substrate comprising the luciferase of claim 9.
19. A fusion protein comprising the second luciferase of claim 1.
20. A fusion protein comprising the luciferase of claim 9.
21. An isolated and purified nucleic acid molecule comprising a nucleic acid segment encoding the fusion protein of claim 19.
22. An isolated and purified nucleic acid molecule comprising a nucleic acid segment encoding the fusion protein of claim 20.

23. A method of using a luciferase, comprising: linking an agent with the second luciferase of claim 1 so as to yield a labeled agent.
24. The use of the second luciferase of claim 1 for detecting ATP, for labeling a molecule, as a genetic reporter, for immobilization onto a solid surface, to produce hybrid proteins, for high temperature reactions, or for creating luminescent solutions.
25. A method of using a luciferase, comprising: linking an agent with the luciferase of claim 9 so as to yield a labeled agent.
26. The use of the luciferase of claim 9 for detecting ATP, for labeling a molecule, as a genetic reporter, for immobilization onto a solid surface, to produce hybrid proteins, for high temperature reactions, or for creating luminescent solutions.
27. A method of using a vector encoding a luciferase, comprising:
 - a) introducing the vector of claim 13 into a host cell; and
 - b) detecting or determining the presence of luciferase in the host cell.
28. A method of using a vector encoding a luciferase, comprising:
 - a) introducing the vector of claim 14 into a host cell; and
 - b) detecting or determining the presence of luciferase in the host cell.
29. A kit comprising: a container comprising the second luciferase of claim 1.
30. The kit of claim 29 wherein the container comprises an aqueous mixture comprising the luciferase.
31. The kit of claim 29 wherein the container comprises lyophilized luciferase.

32. The kit of claim 29 further comprising a container comprising luciferin.
33. The kit of claim 29 wherein the container further comprises luciferin.
34. The kit of claim 31 wherein the container further comprises lyophilized luciferin.
35. The kit of claim 34 wherein the container which comprises luciferin comprises lyophilized luciferin.
36. The kit of claim 29 further comprising packaging material enclosing, separately packaged, the container and instruction means directing the user to correlate the luciferase activity in a sample suspected of having ATP with the level of ATP in the sample.
37. The kit of claim 29 further comprising packaging material enclosing, separately packaged, the container and instruction means directing the user to correlate the luciferase activity in a sample suspected of having an infectious agent that produces ATP with the level or presence of the agent in the sample.
38. A kit comprising: a container comprising the luciferase of claim 9.
39. The kit of claim 38 wherein the container comprises an aqueous mixture comprising the luciferase.
40. The kit of claim 38 wherein the container comprises lyophilized luciferase.
41. The kit of claim 38 further comprising a container comprising luciferin.
42. The kit of claim 38 wherein the container further comprises luciferin.

43. The kit of claim 40 wherein the container further comprises lyophilized luciferin.
44. The kit of claim 41 wherein the container which comprises luciferin comprises lyophilized luciferin.
45. The kit of claim 38 further comprising packaging material enclosing, separately packaged, the container and instruction means directing the user to correlate the luciferase activity in a sample suspected of having ATP with the level of ATP in the sample.
46. The kit of claim 38 further comprising packaging material enclosing, separately packaged, the container and instruction means directing the user to correlate the luciferase activity in a sample suspected of having an infectious agent that produces ATP with the level or presence of the agent in the sample.
47. A method to prepare an enzyme which is not a beetle luciferase and which has enhanced enzymological properties, comprising:
 - a) selecting one or more isolated polynucleotide sequences encoding an enzyme which is not a luciferase and which has at least one enhanced enzymological property from a first population of polynucleotide sequences obtained from a first isolated polynucleotide sequence encoding an enzyme which is not a luciferase, wherein the first isolated polynucleotide sequence is subjected to conditions that yield nucleotide mutations, wherein the enzyme encoded by the one or more selected isolated polynucleotide sequences has at least one enhanced enzymological property relative to the enzyme encoded by the first isolated polynucleotide sequence;
 - b) mutating the selected isolated polynucleotide sequence to yield a second population of polynucleotide sequences, wherein the selected isolated polynucleotide sequence is subjected to oligonucleotide mediated mutagenesis with a plurality of oligonucleotides each

comprising at least one codon that encodes a consensus amino acid which is not present in the first polynucleotide sequence; and
c) repeating step a) and step b) so as to yield a further polynucleotide sequence encoding an enzyme which is not a luciferase having at least one enhanced enzymological property and comprising a plurality of amino acid substitutions relative to the enzyme encoded by the first polynucleotide sequence.

48. The method of claim 47 further comprising isolating the further polynucleotide sequence.
49. The method of claim 47 wherein in step b) a mixture of the selected isolated polynucleotide sequences of step a) are mutated.
50. The method of claim 47 wherein the property is specific activity, enzyme activity, catalytic turnover, Km or substrate utilization.
51. The method of claim 47 wherein the enzyme is DNA polymerase or RNA polymerase.
52. The method of claim 47 where the enzyme is detectable in crude cellular lysates or cells.
53. The method of claim 52 wherein the enzyme is chloramphenicol acetyltransferase, beta-glucuronidase or beta-galactosidase.
54. The method of claim 47 wherein the first polynucleotide sequence is subjected to recombination mutagenesis.
55. The method of claim 47 wherein the first polynucleotide sequence is subjected to point mutagenesis.

56. The method of claim 47 wherein the selection is an automated multi-parameter process.
57. A polynucleotide sequence which is obtained by the method of claim 47.
58. A method to prepare an enzyme that is resistant to an inhibitor, comprising:
 - a) selecting one or more isolated polynucleotide sequences encoding an enzyme which is resistant to an inhibitor from a first population of polynucleotide sequences obtained from a first isolated polynucleotide sequence encoding an enzyme subjected to conditions that yield nucleotide mutations, wherein the enzyme encoded by the one or more selected isolated polynucleotide sequences has increased resistance to an inhibitor relative to the enzyme encoded by the first isolated polynucleotide sequence;
 - b) mutating the selected isolated polynucleotide sequence to yield a second population of polynucleotide sequences; and
 - c) repeating step a) and step b) so as to yield a further polynucleotide sequence encoding an enzyme that is resistant to an inhibitor and comprises a plurality of amino acid substitutions relative to the enzyme encoded by the first polynucleotide sequence.
59. The method of claim 58 further comprising isolating the further polynucleotide sequence.
60. The method of claim 58 wherein in step b) a mixture of the selected isolated polynucleotide sequences of step a) are mutated.
61. The method of claim 58 wherein the further polynucleotide sequence encodes an enzyme that has increased thermostability relative to the first polynucleotide sequence.

62. The method of claim 58 wherein the enzyme is DNA polymerase or RNA polymerase.
63. The method of claim 58 wherein the enzyme is resistant to inhibition by a substrate analog of the enzyme.
64. The method of claim 58 wherein the mutating employs oligonucleotides having at least one codon encoding a consensus amino acid.
65. The method of claim 58 wherein the enzyme is a luciferase.
66. The method of claim 65 wherein the luciferase is a beetle luciferase.
67. The method of claim 66 wherein the first polynucleotide sequence encodes *LucPpe2*.
68. The method of claim 66 wherein the first polynucleotide sequence encodes *LucPpl*.
69. The method of claim 58 wherein a plurality of amino acid substitutions are to a consensus amino acid.
70. The method of claim 58 wherein the first polynucleotide sequence is subjected to recombination mutagenesis.
71. The method of claim 58 wherein the first polynucleotide sequence is subjected to point mutagenesis.
72. The method of claim 58 wherein the selection is an automated multi-parameter process.
73. The method of claim 65 wherein the luciferase has increased luminescence intensity, increased signal stability or decreased Km.

100

74. A polynucleotide sequence which is obtained by the method of claim 58.
75. An enzyme which is encoded by the polynucleotide sequence of claim 57.
76. An enzyme which is encoded by the polynucleotide sequence of claim 76.

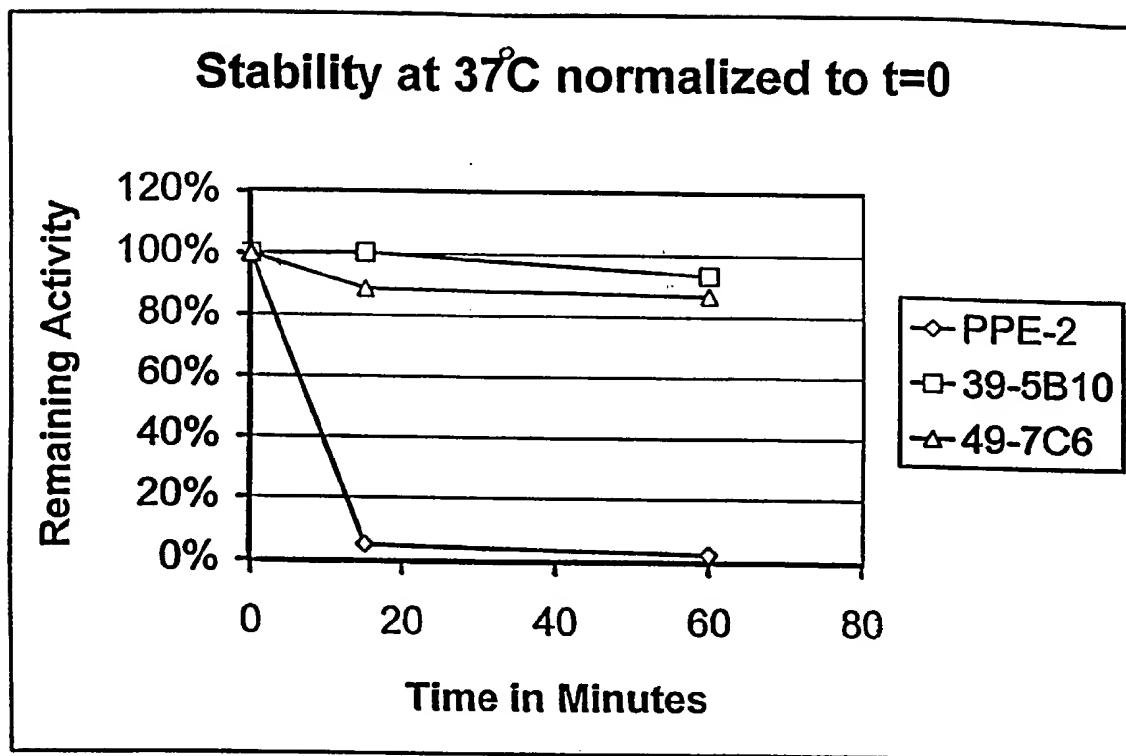


FIG. 1

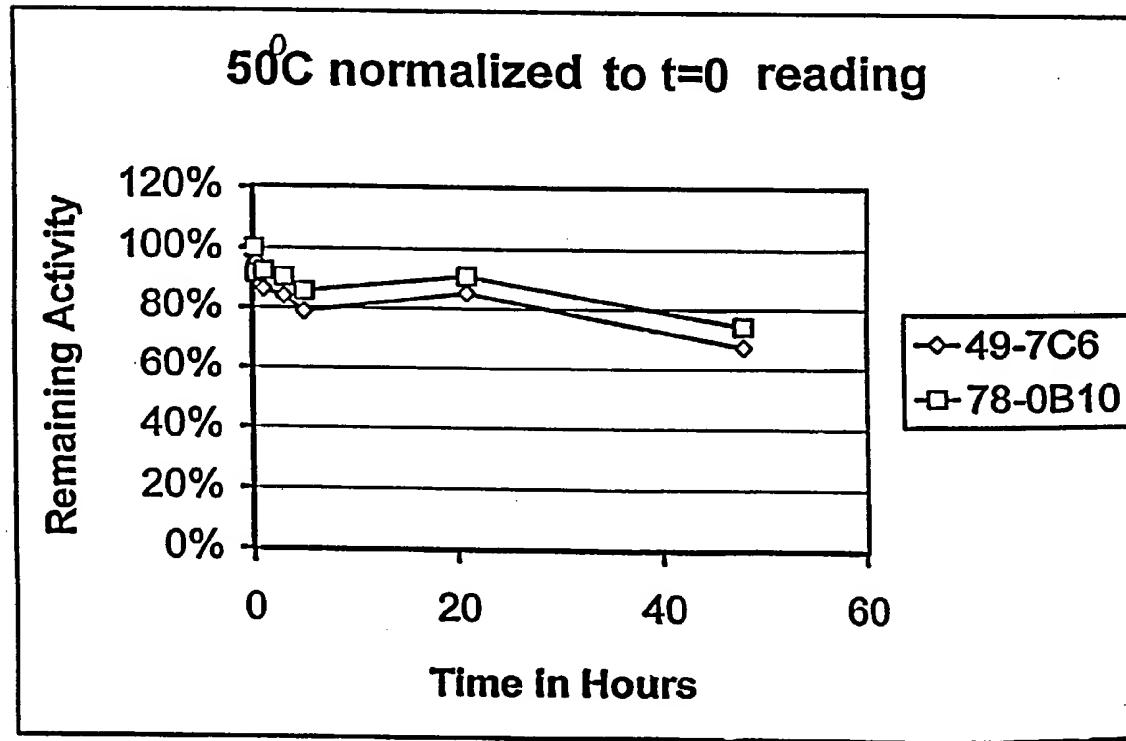


FIG. 2

SUBSTITUTE SHEET (RULE 28)

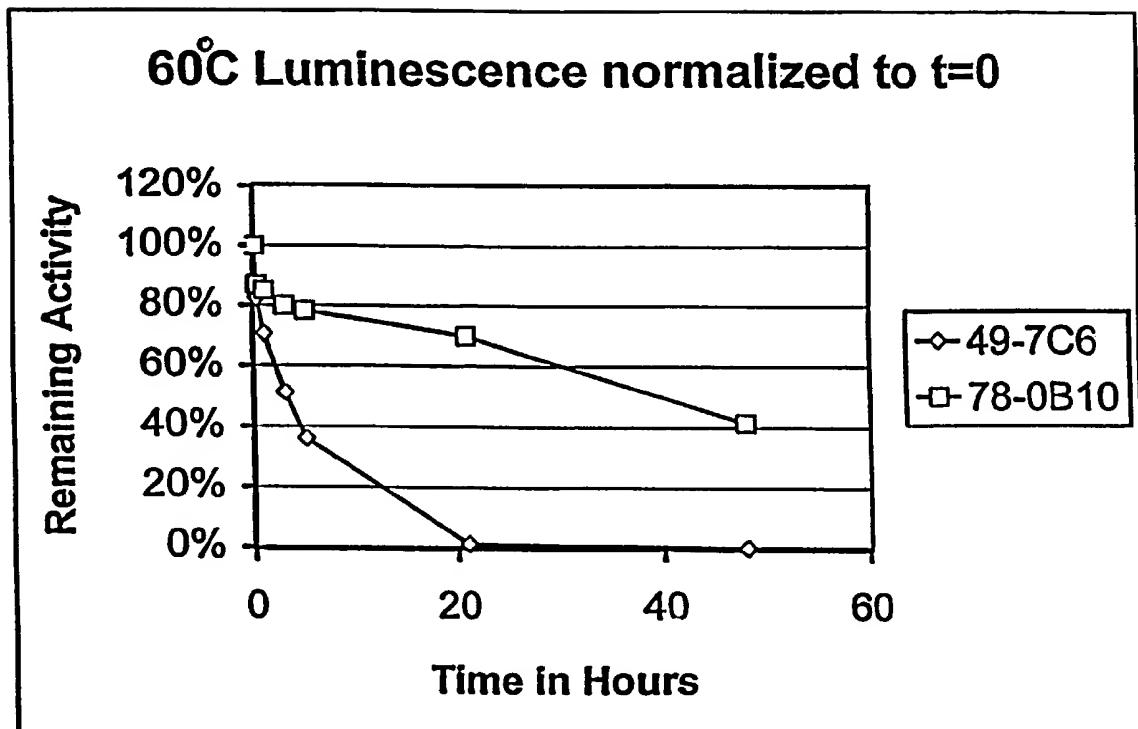


FIG. 3

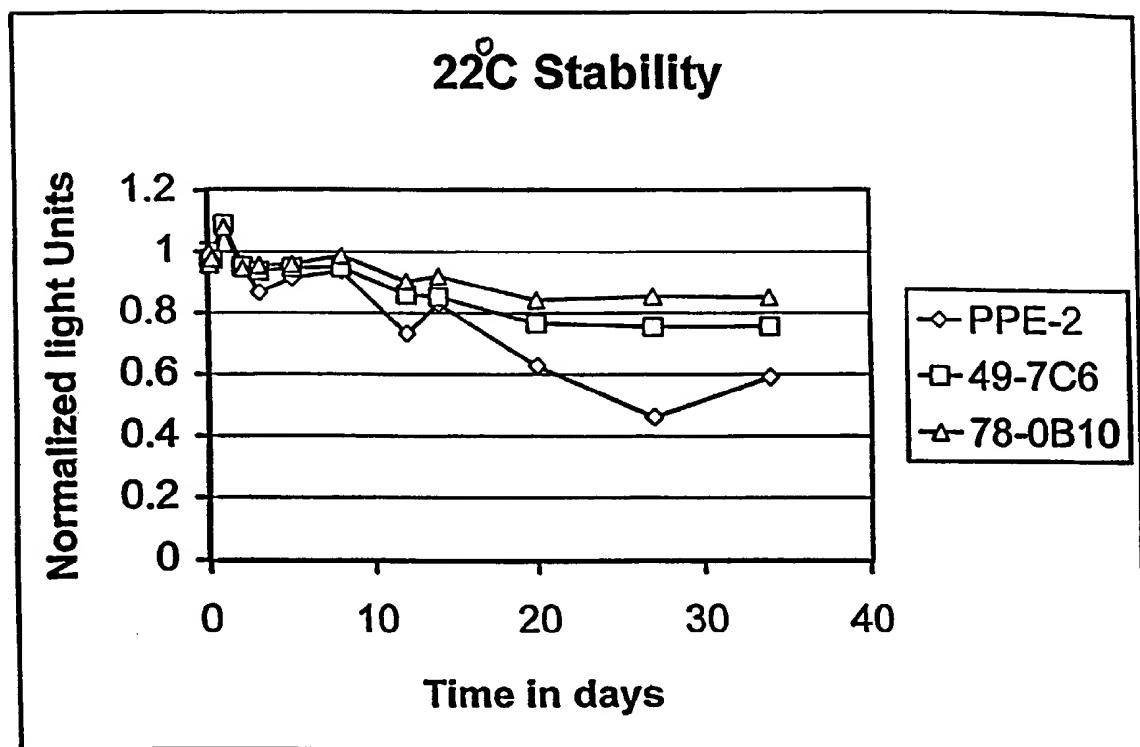


FIG. 4

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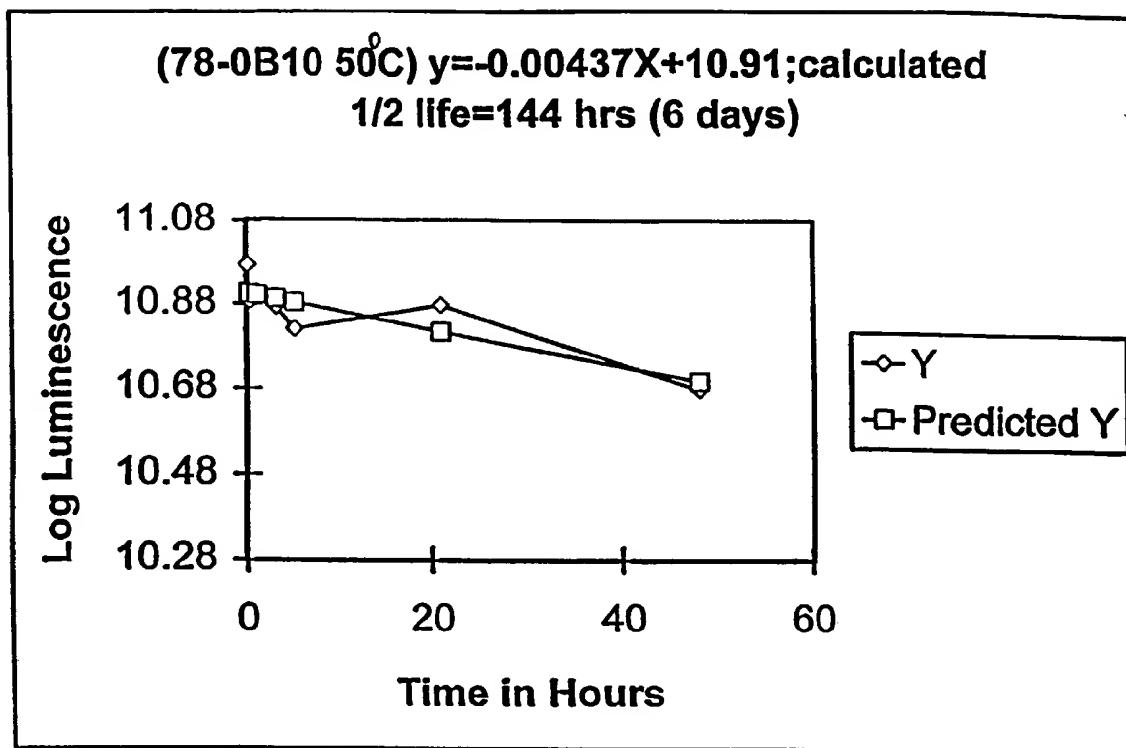


FIG. 5

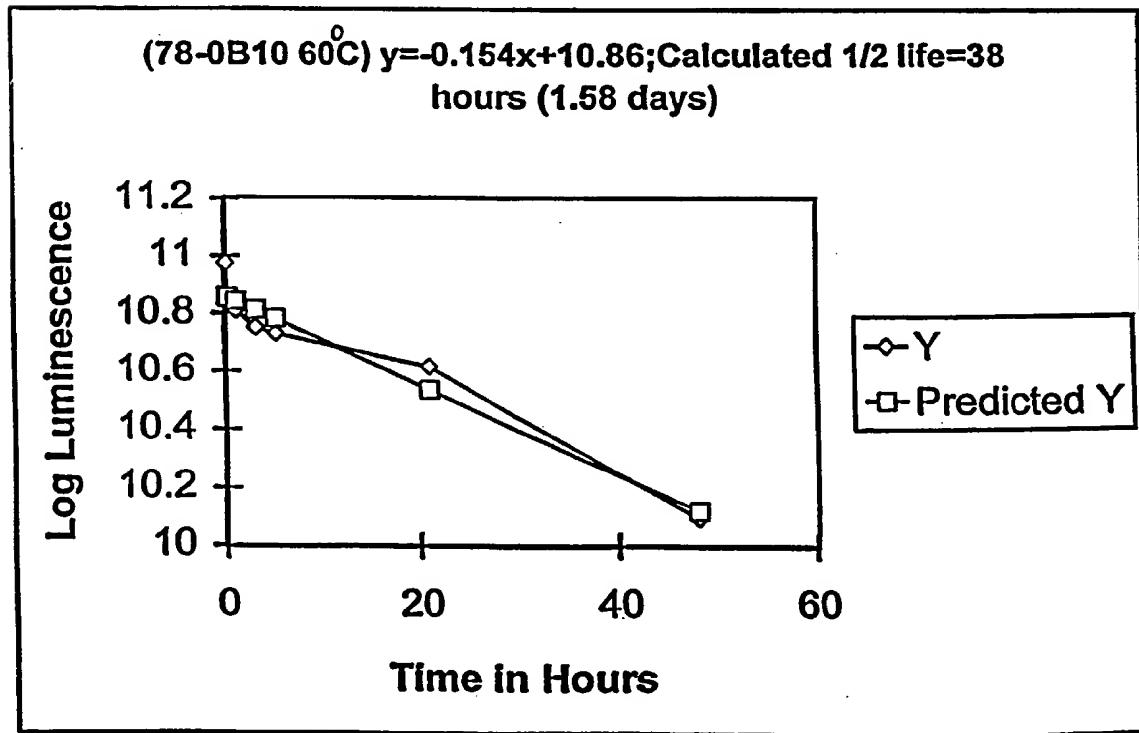


FIG. 6

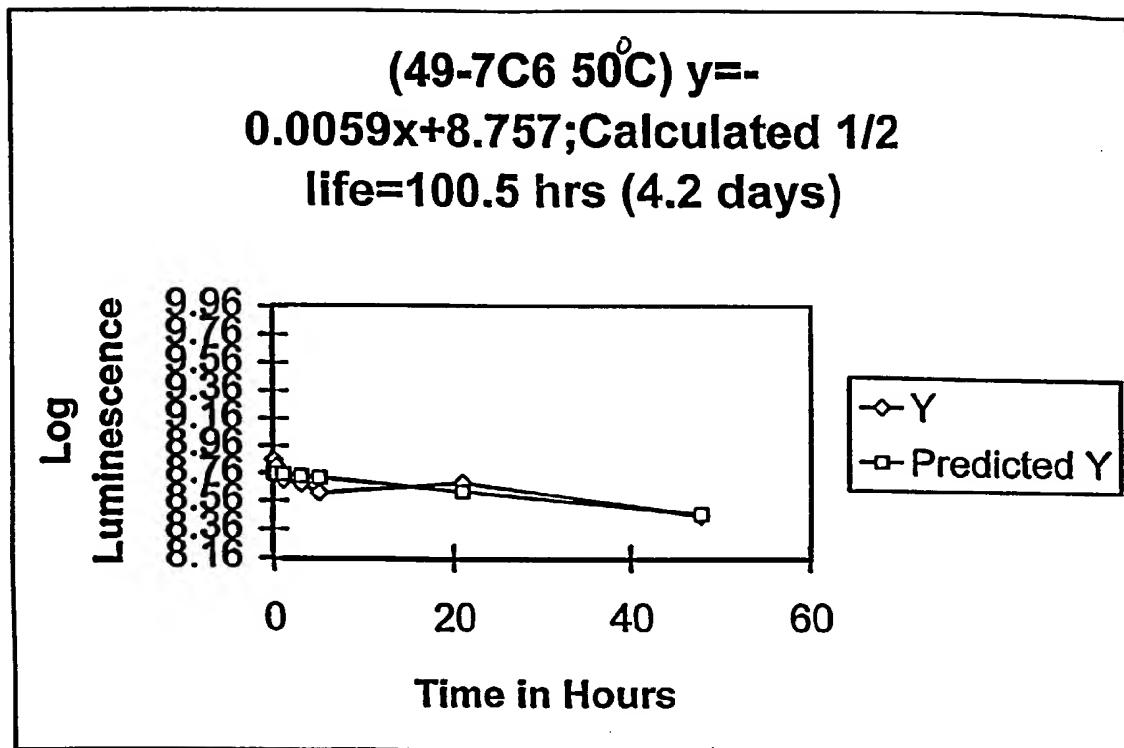


FIG. 7

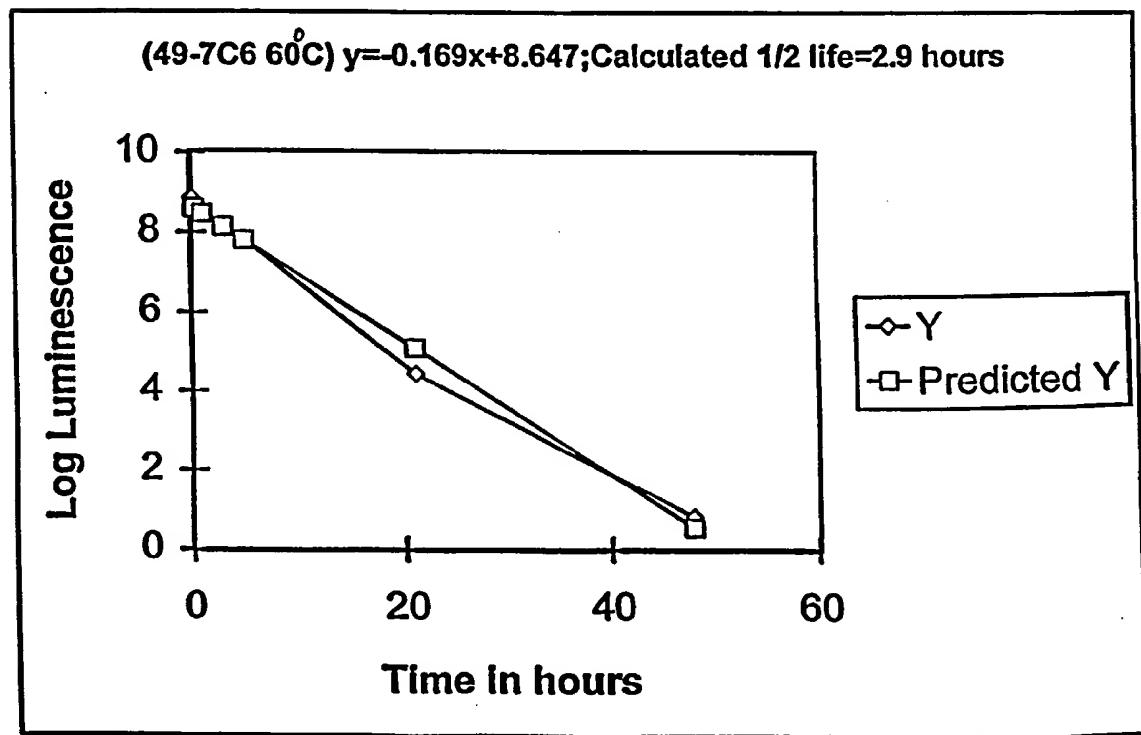


FIG. 8

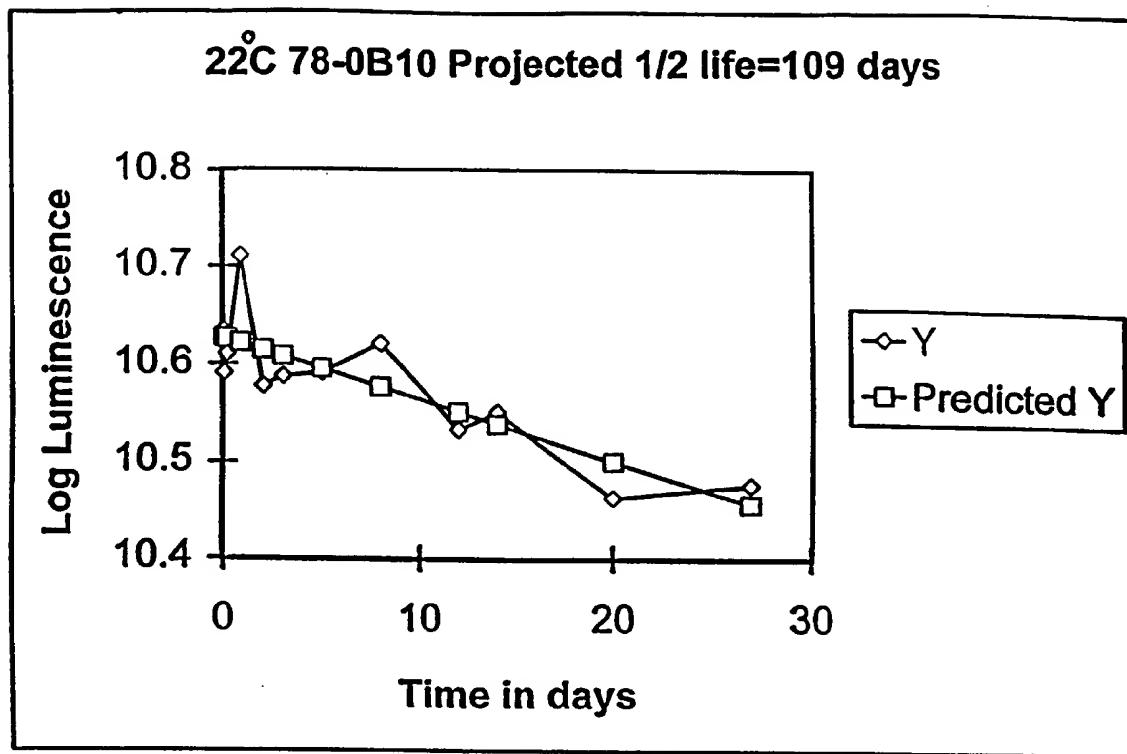


FIG. 9

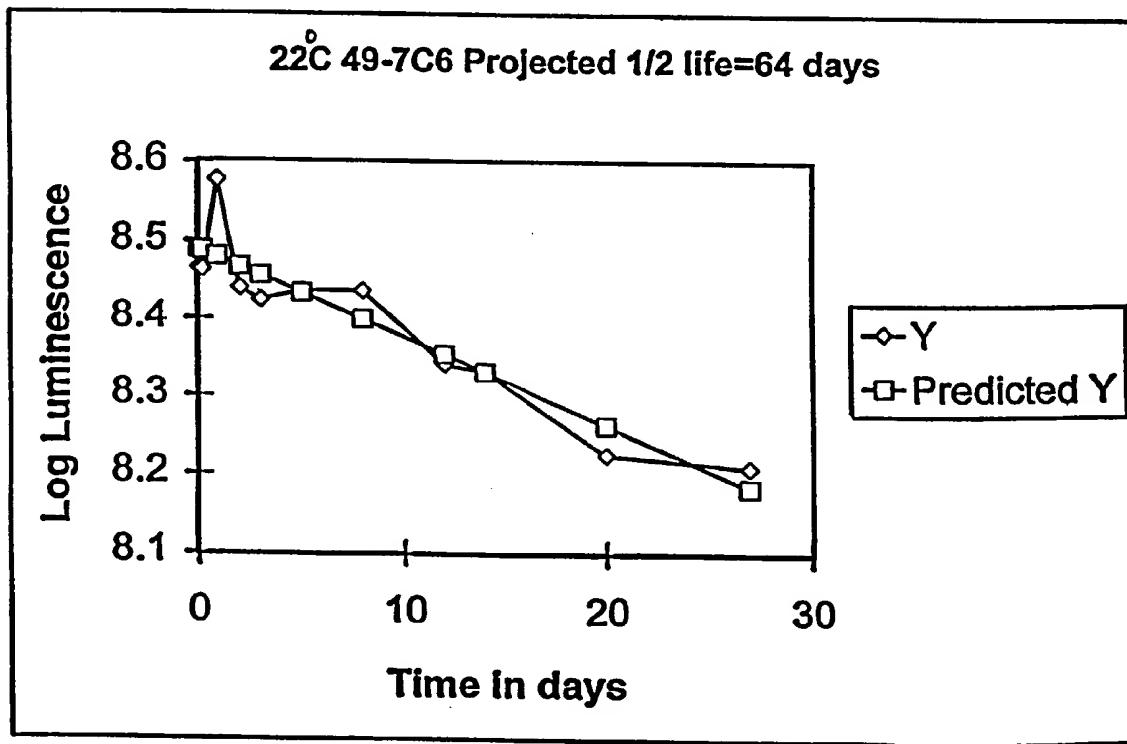


FIG. 10

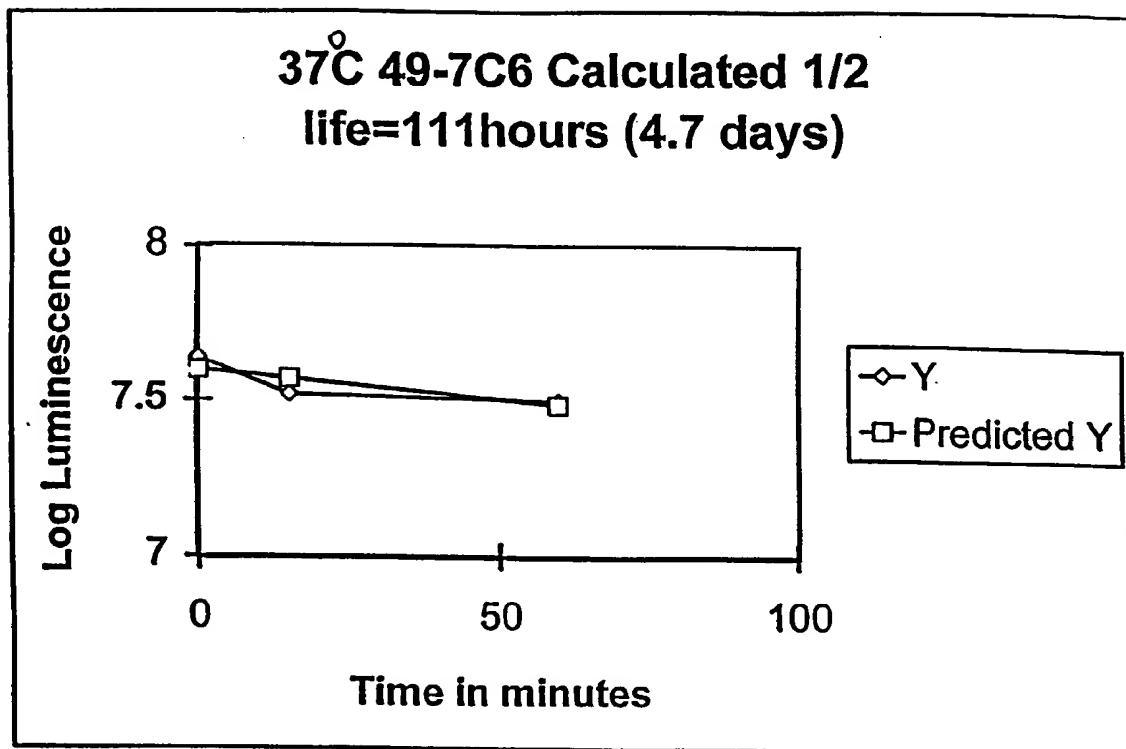


FIG. 11

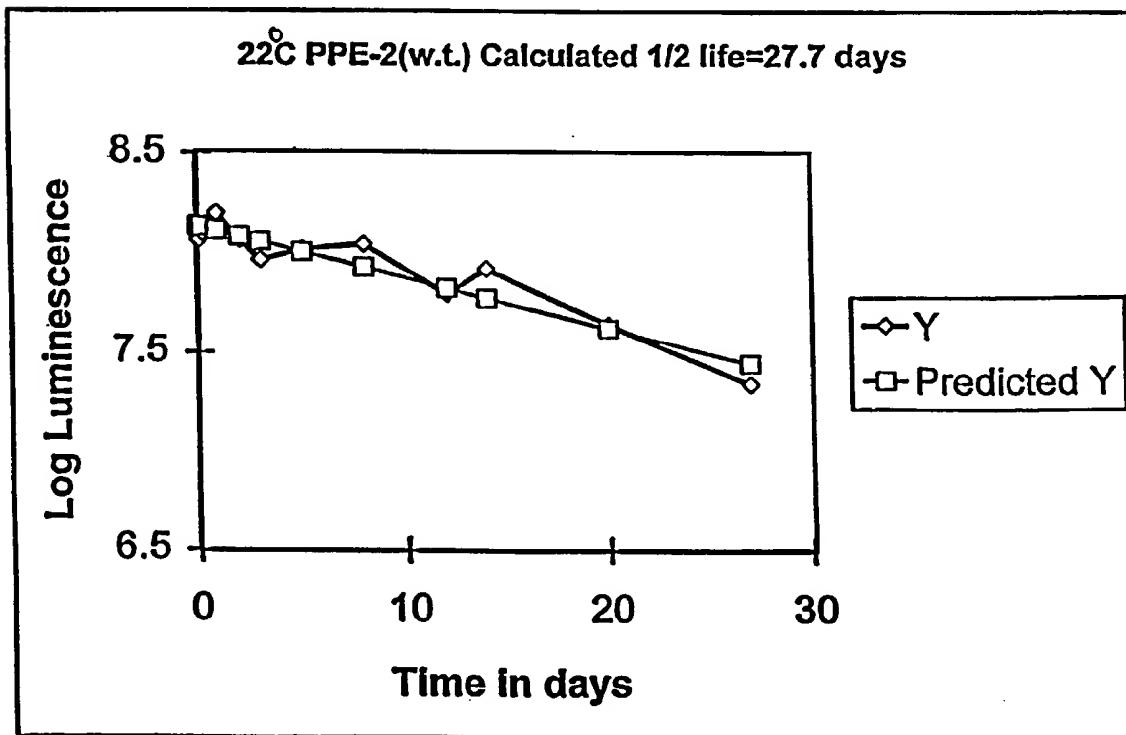


FIG. 12

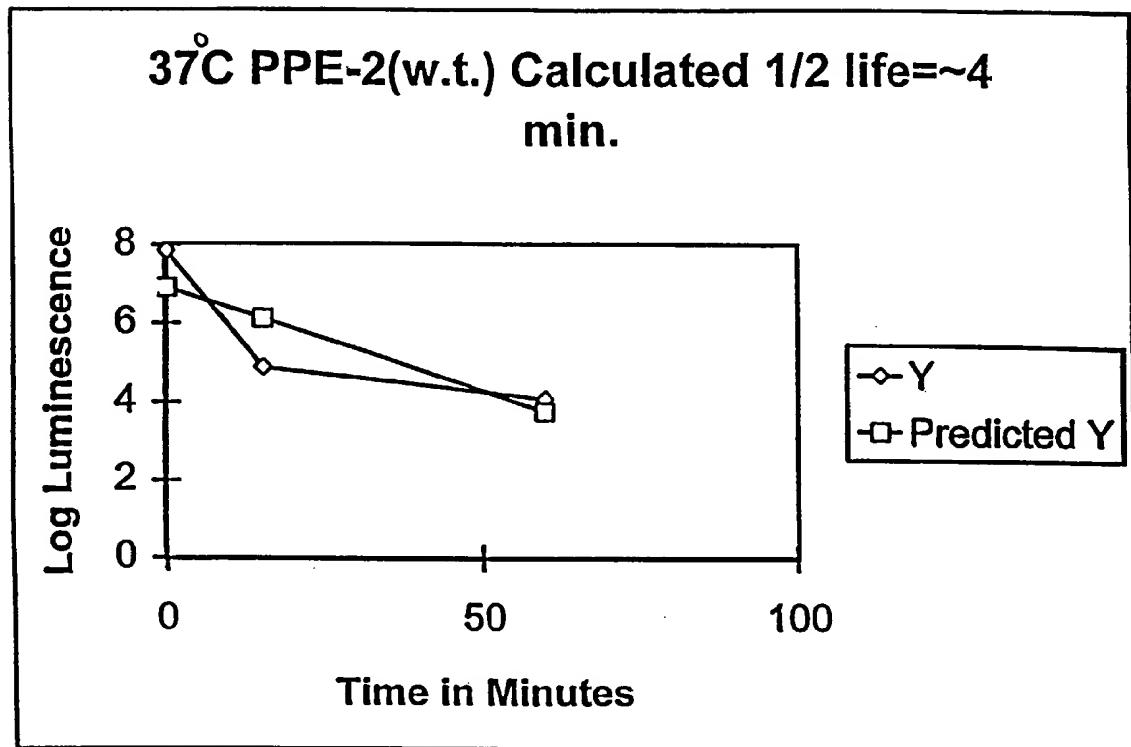


FIG. 13

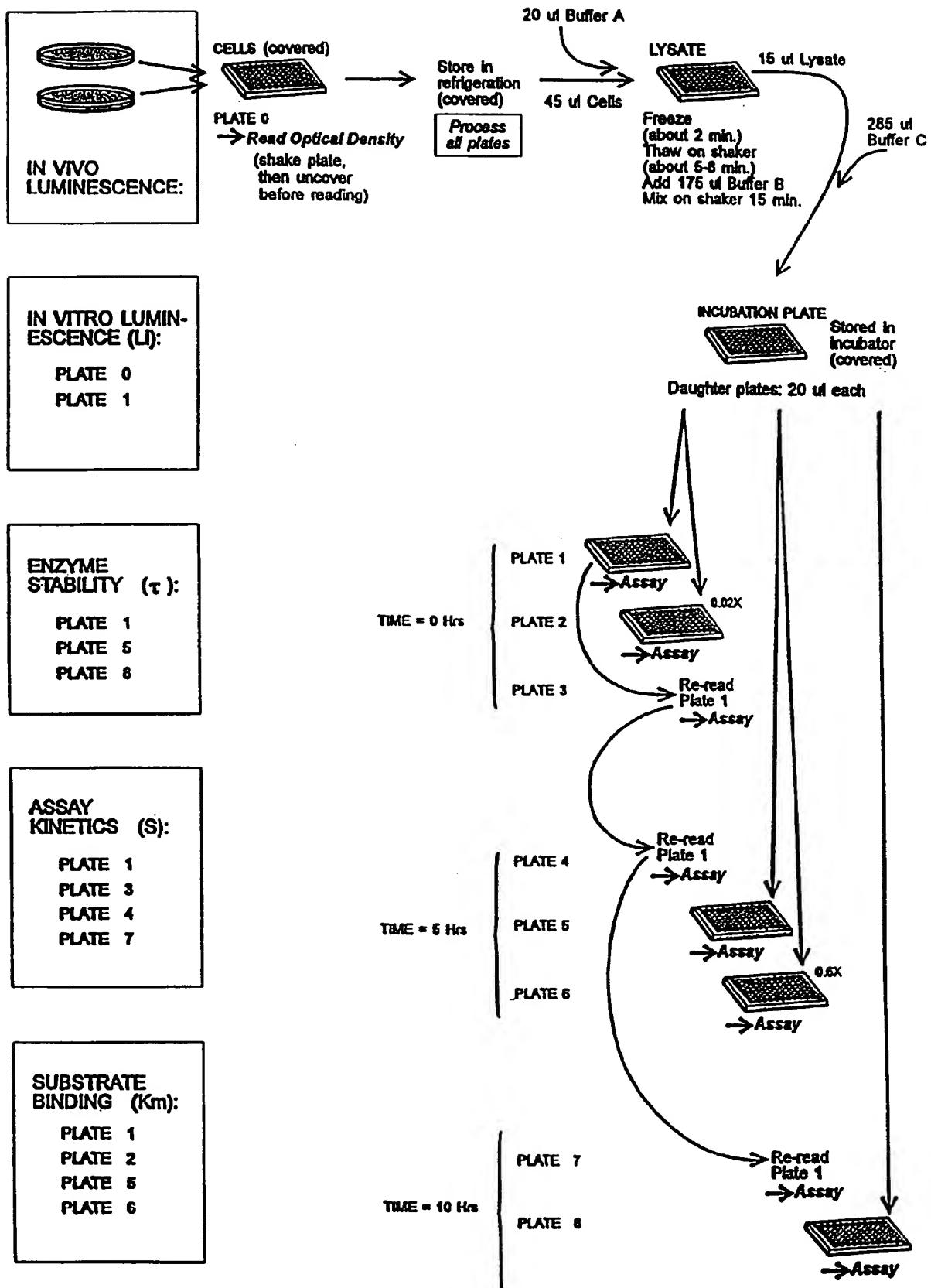
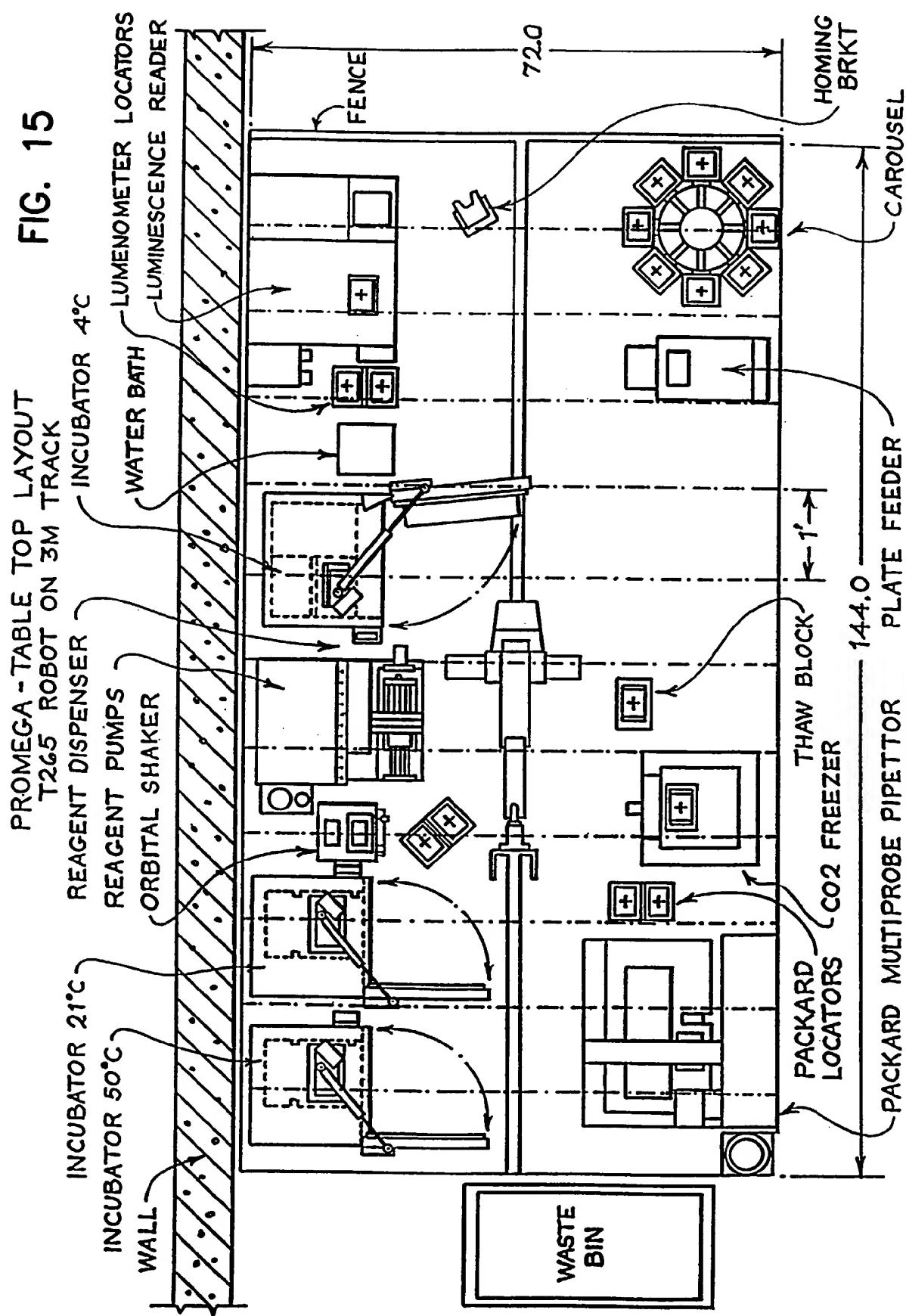


FIG. 14

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FIG. 15



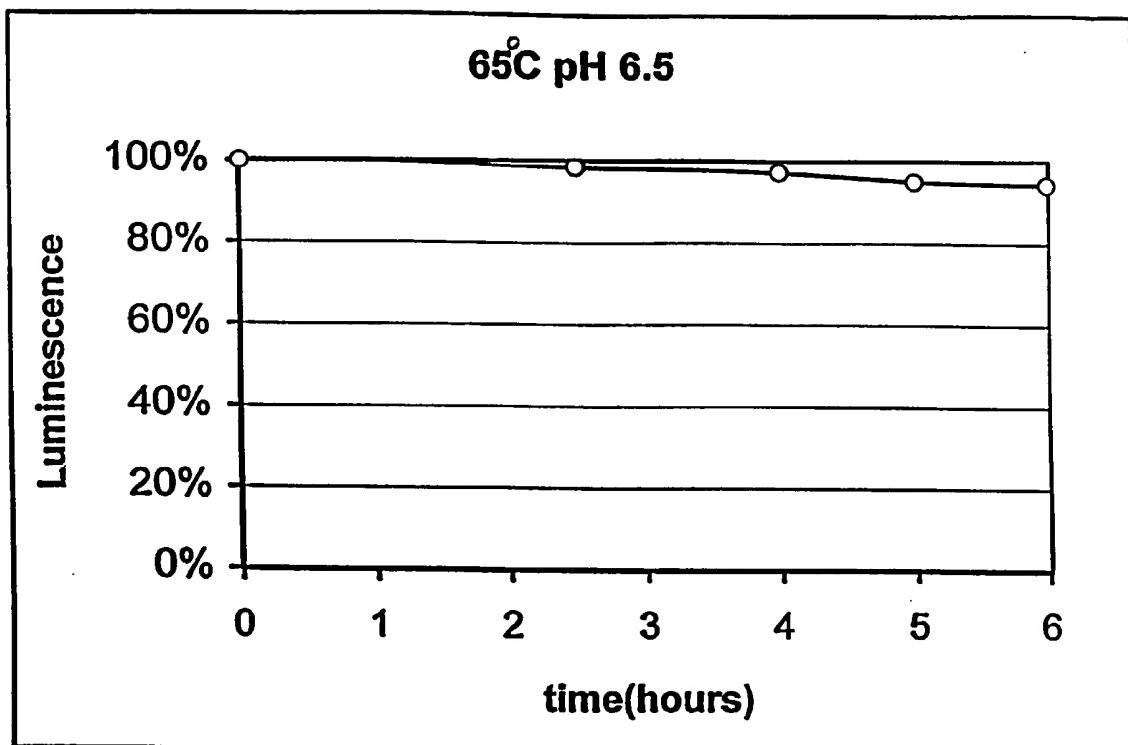


FIG. 16A

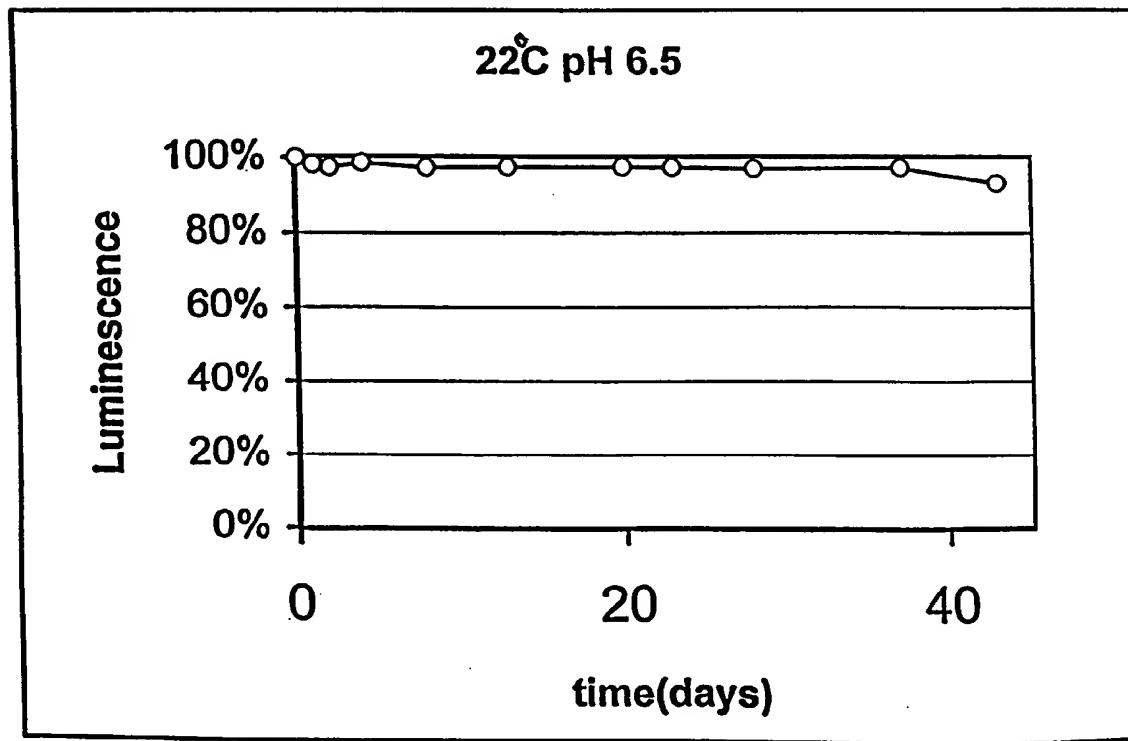


FIG. 16B

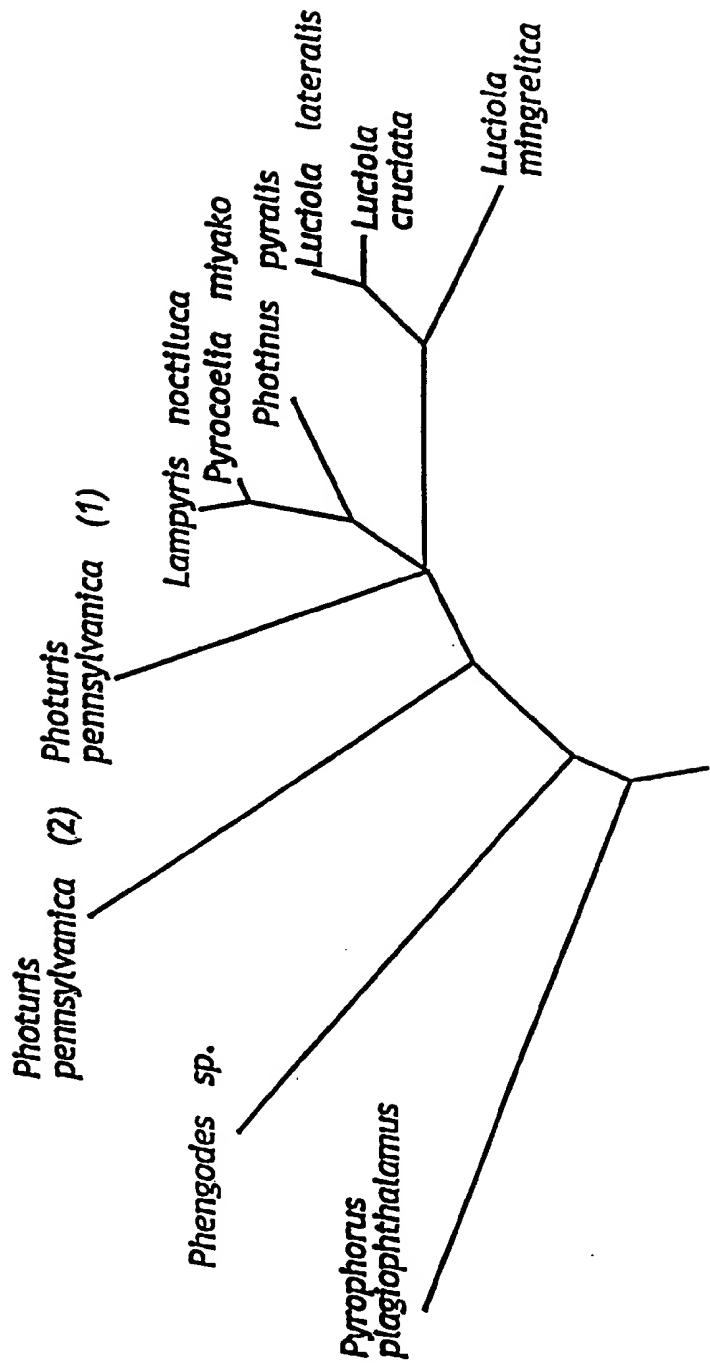


FIG. 17



FIG. 18A

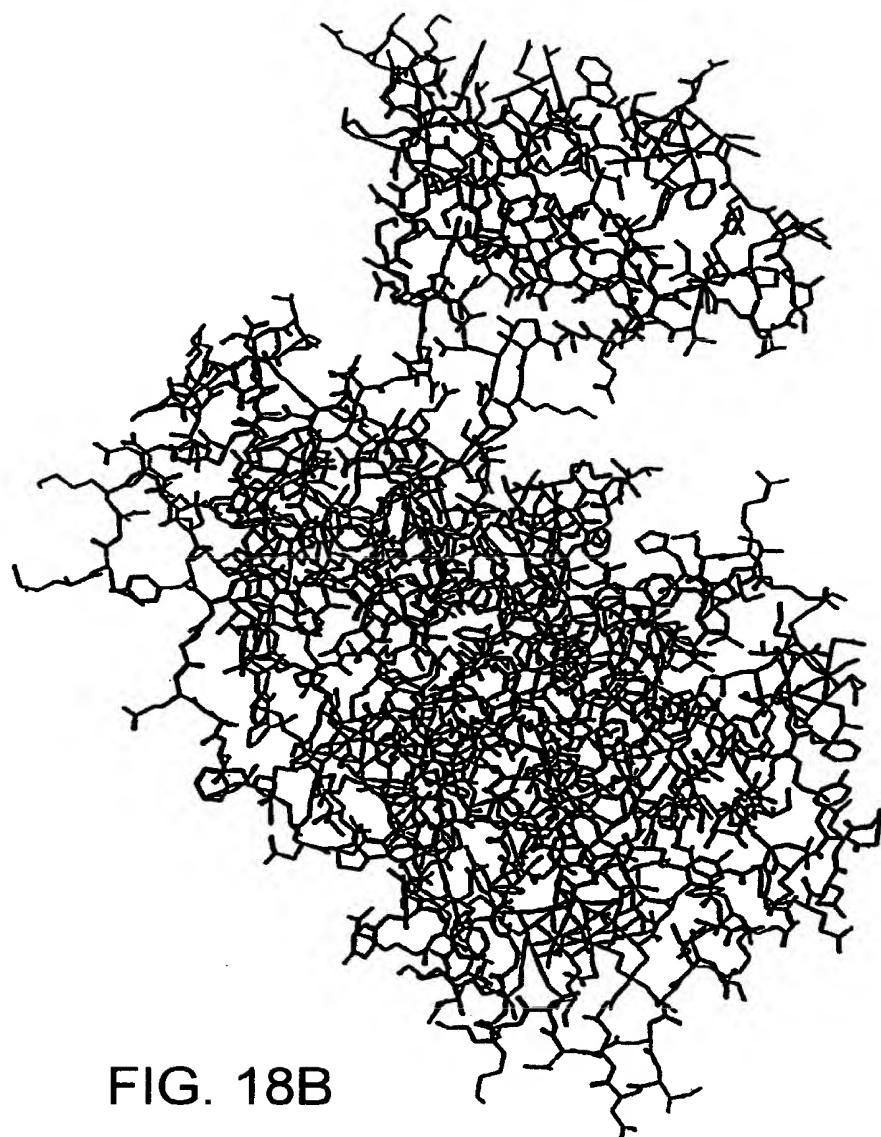


FIG. 18B

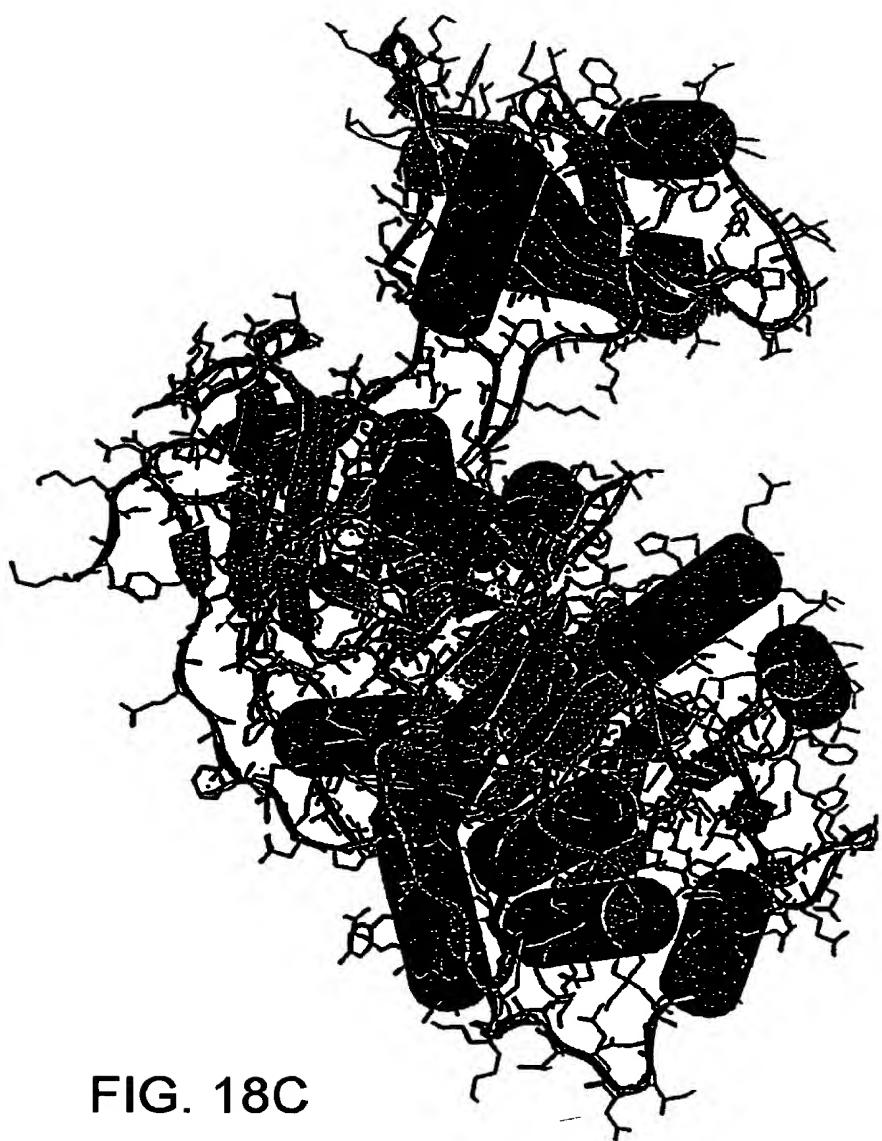


FIG. 18C

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1

50

Lcr MENMENDE.N IVVGPKPFPYP IEEGSAGTQL RKYMERAYKL .GALAFTN
 Lla MENMENDE.N IVYGPEPFYP IEEGSAGAQL RKYMDRYAKL .GALAFTN
 Lmi ME.MEKES.N VVYGPLPFYP IEEGSAGIQL HKYMHQYAKL .GAIAPSN
 Pmi . . . MEDDSKH IMHGRHSIL WEDGTAGEQL HKAMKRYAQV PGTIAFTDAH
 Ppy . . . MED.AKN IKKGPAFPYP LEDGTAGEQL HKAMKRYALV PGTIAFTDAH
 Lno . . . MED.AKN IMHGPAPFYP LEDGTAGEQL HKAMKRYAQV PGTIAFTDAH
 Ppe1 . . . MSI.ENN ILIGPPPYP LERGTAGEQL HRAISRYAAV PGTLAYTDVH
 Phg MIKME..EEH VMPGAMPRDL LFEGTAGCQL HRALVKHSHF PE..AIVD
 GR . . . MMKREKN VVYGPPEPLHP LEDLTAGEML FRALRKHSHL PQ..ALVDV
 YG . . . MMKREKN VIYGPEPLHP LEDLTAGEML FRALRKHSHL PQ..ALVDV
 Ppe2 . . . MED..KN ILYGPEPFYP LADGTAGEQM FYALSRYADI SGCLALTN
 49-7C6 A
 78-0B10 A D
 90-1B5 A E D P
 133-1B2 A E D P
 146-1H2 A E D A P
 Cons ---M--- ---G--- ---AG--- ---A---

51

100

Lcr TGVDYSYAEY LEKSCCLGKA LQNYGLVVDG RIALCSENCE EFFIPVIAGL
 Lla TGVDYTYAEY LEKSCCLGEA LKNYGLVVDG RIALCSENCE EFFIPVLAGL
 Lmi TGVDISYQEY FDITCRLAEE MKNPGMKPEE HIALCSENCE EFFIPVLAGL
 Pmi AEVNITYSEY FEMSCRЛАET MKRYGLGLQH HIAVCSETSL QFFMPVC
 Ppy IEVNITYAEY FEMSVRLAEE MKRYGLANTNH RIVVCS
 Lno AEVNITYSEY FEMACRLAET MKRYGLGLQH HIAVCSEN
 Ppe1 TELEVITYKEF LDVTCRLAEE MKNYGLGLQH TISVCE
 Phg THEIISYAKI LDMSCRЛАAVS FQKYGLTQNN IIGICSENNL NFFNPVIAAF
 GR GEEWISYKEF FETTCLLAQS LHNCGYKMSD VVSICAENN
 YG GDESLSYKEF FEATCLLAQS LHNCGYK
 Ppe2 TKEVLYEEF LKLSCRЛАES FKKYGLQND TIAVCE
 49-7C6 I
 78-0B10 V
 90-1B5 V
 133-1B2 S V
 146-1H2 S V
 Cons ---Y--- ---L--- ---G--- ---C-E--- -FF-P---

101

150

Lcr FIGVGVAPTN EIYTLRELVH SLGISKPTIV FSSKKGLDKV ITVQKTVTT
 Lla FIGVGVAPTN EIYTLRELVH SLGISKPTIV FSSKKGLDKV ITVQKTVAT
 Lmi YIGVAVAPTN EIYTLRELNH SLGIAQPTIV FSSRKGLPKV LEVQKTVTC
 Pmi FIGVGVAPTN DIYNERELYN SLFISQPTIV FCSKRALQKI LGVQK
 Ppy FIGVGAVAPN DIYNERELLN SMNISQPTVV FVSKGLQKI LNVQK
 Lno FIGVGVA
 Ppe1 YVGVATAPTN DIYNERELYN SLSISQPTIV SCSKRALQKI LGVQ
 Phg YLGITVATVN DTYTDRELSE TLNITKPKML FCSKQSLPIV MKTMK
 GR YIGMIVAPVN EGYIPDELCK VMGISRPQLV FCTKNILNKV LEVQSRTDF
 YG YIGMIVAPVN ESYIPDELCK VMGISKPQIV FCTKNILNKV LEVQSRTNF
 Ppe2 YLGIIAAPVS DKYIERELIH SLGIVKPRII FCSKNTFQKV LNVKS
 49-7C6
 78-0B10 S
 90-1B5 V N V SI
 133-1B2 V N V SI
 146-1H2 V N V SI
 Cons ---G---A--- ---Y---EL--- ---I---P---

FIG. 19A

	151		200		
Lcr	KTIVILD SKV	DYRGYQCLDT	FIKRNTPPGF	QASSFKTVEV	.DRKEQVALI
Lla	KTIVILD SKV	DYRGYQSMDN	FIKKNTPQGF	KGSFKTVEV	.NRKEQVALI
Lmi	KKIVILD SKV	NPGQHDCMET	FIKKHVELGP	QPSSFVPIDV	KNRKQHVALL
Pmi	QKIVILD SRE	DYMGKQSMYS	FIESHLPA GF	NEYDYIPDSF	.DRETATALI
Ppy	QKIIIMDSKT	DYQGFQSMYT	FVTSHLPPGF	NEYDFVPESF	.DRDKTIALI
Lno	QKIVILD SRE	DYMGKQSMYS	FIESHLPA GF	NEYDYIPDSF	.DRETATALI
Ppe1	KKIIILDGKK	DYLGYQSMQS	FMKEHVPANF	NVSAFKPLSF	.DLDR.VACI
Phg	QKLLIIDS MQ	DIGGIEBCVHS	FVSRYTDEHF	DPLKFVPLDF	.DPREQVALI
GR	KRIIILD AVE	NIHGCESLPN	FISRYSDGNI	A..NFKPLHY	.DPVEQVAAI
YG	KRIIILD TVE	NIHGCESLPN	FISRYSDGNI	A..NFKPLHY	.DPVEQVAAI
Ppe2	ETIIILD LNE	DLGGYQCLNN	FISQNSDINL	DVKFKPNSF	.NRDDQVALV
49-7C6				Y	
78-0B10			S	Y	
90-1B5			S	Y	I
133-1B2	D		S	Y	I
146-1H2			S	Y	SI
Cons	----I-D---	----G-----	F-----	-----	A--
	201		250		
Lcr	MNSSGSTGLP	KGVQLTHENT	VTRFSHARDP	IYGNQVSPGT	AVLTVVVFHH
Lla	MNSSGSTGLP	KGVQLTHENA	VTRFSHARDP	IYGNQVSPGT	AILTVVVFHH
Lmi	MNSSGSTGLP	KGVRITHEGA	VTRFSHARDP	IYGNQVSPGT	AILTVVVFHH
Pmi	MNSSGSTGLP	KGVDLTHMNV	CVRFSHCRDP	VFGNQIIPDT	AILTVIPFHH
Ppy	MNSSGSTGLP	KGVALPHRTA	CVRFSHARDP	IFGNQIIPDT	AILSVVVFHH
Lno	MNSSGSTGLP	KGVELTHQNV	CVRFSHCRDP	VFGNQIIPDT	AILTVIPFHH
Ppe1	MNSSGSTGLP	KGVPISHRNT	IYRFSHCRDP	VFGNQIIPDT	TILCAVPFHH
Phg	MTSSGTTGLP	KGVMLTHRNI	CVRFVHSRDP	LFGTRFIPET	SILSLVPFHH
GR	LCSSGTTGLP	KGVMQTHRNV	CVRLIHALDP	RVGTQLIPGV	TVLVYLPFFH
YG	LCSSGTTGLP	KGVMQTHQNI	CVRLIHALDP	RAGTQLIPGV	TVLVYLPFFH
Ppe2	MFSSGTTGVS	KGVMLTHKNI	VARFSHCKDP	TFGNAINPTT	AILTVIPFHH
49-7C6			LA		
78-0B10	P		LA		
90-1B5	LP		LA		
133-1B2	LP		IA	S	
146-1H2	LP		IA	S	
Cons	--SSG-TG--	KGV---H---	--R--H--DP	--G----P--	--L---PF-H
	251		300		
Lcr	GFGMFTTILGY	LICGFRVVML	TKFDEETFLK	TLQDYKCTSV	ILVPTLFAIL
Lla	GFGMFTTILGY	LTCGFRIVML	TKFDEETFLK	TLQDYKCSSV	ILVPTLFAIL
Lmi	GFGMFTTILGY	FACGYRVVML	TKFDEELFLR	TLQDYKCTSV	ILVPTLFAIL
Pmi	VFQMFITLGY	LTCGFRIVML	YRFEEELFLR	SLQDYKIQSA	LLVPTLFSFF
Ppy	GFGMFTTILGY	LICGFRVVML	YRFEEELFLR	SLQDYKIQSA	LLVPTLFSFF
Lno	GFGMFTTILGY	LTCGFRIVML	YRFEEELFLR	SLQDYKIQSA	LLVPTLFSFF
Ppe1	AFGTFTNLGY	LICGFHVVL	YRFNEHFLQ	TLQDYKCQSA	LLVPTVLAFL
Phg	AFGMFTTILSY	FIVGLXIVMM	KRFDGELFLK	TIQNYKIPTI	VIAPPVMVFL
GR	AFGFSINLGY	FMVGLRVIML	RRFDQEAFLK	AIQDYEVRSV	INVPAIILFL
YG	AFGFSINLGY	FMVGLRVIML	RRFDQEAFLK	AIQDYEVRSV	INVPAIILFL
Ppe2	GFGMTTILGY	FTCGFRVALM	HTFEEKLFLQ	SLQDYKVEST	LLVPTLMAFF
49-7C6	M	V			L
78-0B10	M	V			L
90-1B5	M	V			L
133-1B2	M	V			L
146-1H2	M	V			L
Cons	-F-----L-Y	--G-----	--F----FL-	--Q-Y-----	--P-----

FIG. 19A (Continued)

301

350

Lcr NKSELLNKYD LSNLVEIASG GAPLSKEVGE AVARRFNLPG VRQGYGLTET
 Lla NRSELLDKYD LSNLVEIASG GAPLSKEIGE AVARRFNLPG VRQGYGLTET
 Lmi NKSELLIDKFD LSNLVEIASG GAPLAKEVGE AVARRFNLPG VRQGYGLTET
 Pmi AKSTLVVDKYD LSNLHEIASG GAPLAKEVGE AVAKRFKLPG IRQGYGLTET
 Ppy AKSTLIDKYD LSNLHEIASG GAPLSKEVGE AVAKRFKLPG IRQGYGLTET
 Lno AKSTLVVDKYD LSNLHEIASG GAPLAKEVGE AVAKRFKLPG IRQGYGLTET
 Ppe1 AKNPLVVDKYD LSNLHEIASG GAPLSKEISE IAAKRFKLPG IRQGYGLTET
 Phg AKSHLVVDKYD LSSIKEIATG GAPLGPALAN AVAKRLKLGG IIQGYGLTET
 GR SKSPLVVDKYD LSSLRELCCG AAPLAKEVAE IAVKRLNLPG IRCGFGLTES
 YG SKSPLVVDKYD LSSLRELCCG AAPLAKEVAE VAVKRLNLPG IRCGFGLTES
 Ppe2 AKSALVEKYD LSHLKEIASG GAPLSKEIGE MVKKRFKLNP VRQGYGLTET

49-7C6

78-0B10

90-1B5

133-1B2

146-1H2

Cons ----L--K-D LS---E---G -APL----- -----R--L-- ---G-GLTE-

351

400

Lcr TSALIIITPEG DDKPGASGKV VPLFKAKVID LDTKKSLGP N RRGEVCVKGP
 Lla TSALIIITPEG DDKPGASGKV VPLFKAKVID LDTKKTLGP N RRGEVCVKGP
 Lmi TSALIIITPEG DDKPGASGKV VPLFKVKVID LDTKKTLGV N RRGEICVKGP
 Pmi TSALIIITPEG DDKPGACGKV VPFFTAKIVD LDTGKTLGV N QRGEELCVKGP
 Ppy TSAILITPEG DDKPGAVGKV VPFFFEAKVVD LDTGKTLGV N QRGEELCVRGP
 Lno TSALIIITPEG DDKPGACGKV VPFFSAKIVD LDTGKTLGV N QRGEELCVKGP
 Ppe1 TCAIVITAEG EFKLGAVGKV VPFYSLKVLD LNTGKKLGPN ERGEICFKGP
 Phg CCAVLITPHN KIKTGSTGQV LPYVTAKIVD TKTGKNLGP N QTGEELCPKSD
 GR TSANIHSLRD EFKSGSLGRV TPLMAAKIAD RETGKALGP N QVGEELCIKGP
 YG TSANIHSLSGD EFKSGSLGRV TPLMAAKIAD RETGKALGP N QVGEELCVKGP
 Ppe2 TSAVLITPDT DVPKGSTGKI VPFHAVKVVD PTTGKILGP N ETGELYFKGD

49-7C6

NN

P

78-0B10

KG

A

P

A

90-1B5

KG

AK

P

P

133-1B2

KG

AK

P

P

146-1H2

KG

AK

L

P

P

Cons --A----- -----G--G-- -P----K--D --T-K-LG-N --GE-----

401

450

Lcr MLMKGYVNNP EATKELIDEE GWLHTGDIGY YDEEKHFFIV DRLKSLIKYK
 Lla MLMKGYVDNP EATREIIDEE GWLHTGDIGY YDEEKHFFIV DRLKSLIKYK
 Lmi SMLMGYSNNP EATRETIIDEE GWLHTGDIGY YDEDEEHFFIV DRLKSLIKYK
 Pmi MIMKGYVNNP EATNALIDKD GWLHSGDIAY YDKDGHFFIV DRLKSLIKYK
 Ppy MIMSGYVNNP EATNALIDKD GWLHSGDIAY WDEDEEHFFIV DRLKSLIKYK
 Lno MIMKGYVNNP EATSALIDKD GWLHSGDIAY YDKDGHFFIV DRLKSLIKYK
 Ppe1 MIMKGYIINP EATRELIIDEE GWHSGDIY FDEDEGHFYIV DRLKSLIKYK
 Phg IIMKGYYQNE EETRLVIDKD GWLHSGDIY YDTDGNFHIV DRLKELIKYK
 GR MVSKGYVNNV EATKEAIDDD GWLHSGDFGY YDEDEEHFYVV DRYKELIKYK
 YG MVSKGYVNNV EATKEAIDDD GWLHSGDFGY YDEDEEHFYVV DRYKELIKYK
 Ppe2 MIMKSYYNNE EATKAIINKD GWLRSGDIAY YDNDGHFYIV DRLKSLIKYK

49-7C6

G

DN

78-0B10

G

DN

90-1B5

G

DN

133-1B2

G

DN

146-1H2

G

DN

Cons -----Y--N- E-T---I--- GW---GD--Y -D-----V DR-K-LIKYK

FIG. 19A (Continued)

451	500
Lcr	GYQVPPAEL E SVLLQHPSIF DAGVAGVPDP VAGELPGAVV VLESGNMTE
Lla	GYQVPPAEL E SVLLQHPNIF DAGVAGVPDP IAGELPGAVV VLEKGKSMTE
Lmi	GYQVPPAEL E SVLLQHPNIF DAGVAGVPDP DAGELPGAVV VMKGKTMTE
Pmi	GYQVPPAEL E SILLQHPFIF DAGVAGIPDP DAGELPAAVV VLEEGKMMTE
Ppy	GYQVAPAELE SILLQHPNIF DAGVAGLPDD DAGELPAAVV VLEHGKTMTE
Lno	GYQVPPAEL E SILLQHPFIF DAGVAGIPDP DAGELPAAVV VLEEGKTMTE
Ppe1	GYQVPPAEL E ALLLQHPIE DAGVAGVPDE VAGDLPGAVV VLKEGKSITE
Phg	AYQVAPAELE ALLLQHPIA DAGVTGIPDE EAGELPAACV VLEPGKTMTE
GR	GSQVAPAELE EILLKNPCIR DVAVVGIPDL EAGELPSAFV VIQPGKEITA
YG	GSQVAPAELE EILLKNPCIR DVAVVGIPDL EAGELPSAFV VKQPGKEITA
Ppe2	GYQVAPAEIE GILLQHPIIV DAGVTGIPDE AAGELPAAGV VVQTGKYLNE

49-7C6

78-0B10

90-1B5

133-1B2

146-1H2

Cons --QV-PAE-E --LL--P-I- D--V-G-PD- -AG-LP-A-V V---GK----

501	550
Lcr	KEVMDYVASQ VSNAKRLRGG VRFVDEVPKG LTGKIDGRA. IREILKKPV.
Lla	KEVMDYVASQ VSNAKRLRGG VRFVDEVPKG LTGKIDGKA. IREILKKPV.
Lmi	KIEIVDYVNSQ VVNHKRLRGG VRFVDEVPKG LTGKIDAKV. IREILKKPQ.
Pmi	QEIVMDYVAGQ VTASKRLRGG VKFVDEVPKG LTGKIDSRK. IREILTMGQK
Ppy	KEIVDYVASQ VTTAKKLRRGG VVFVDEVPKG LTGKLDARK. IREILIKAKK
Lno	QEIVMDYVAGQ VTASKRLRGG VKFVDEVPKG LTGKIDGRK. IREILMMGKK
Ppe1	KEIQDYVAGQ VTSSKRLRGG VEFVKEVPKG FTGKIDTRK. IKEILIKAQK
Phg	KEVMDYLAER VTPTKRLRGG VLFVNNIPKG ATGKLVRTE. LRRLLLTQRA.
GR	KEVYDYLAER VSHTKYLRRGG VRFVDSIPRN VTGKITRKE L LKQOLLEKS..
YG	KEVYDYLAER VSHTKYLRRGG VRFVDSIPRN VTGKITRKE L LKQOLLEKS..
Ppe2	QIVQNFVSSQ VSTAKWLRGG VKFLDEIPKG STGKIDRKV. LRQMFEKH..

49-7C6

78-0B10 D

90-1B5 DY A

133-1B2 DY A

146-1H2 DY A

Cons ----- V---K-LRGG V-F----P-- -TGK-----

551	
LcrAKM
LlaAKM
LmiAKM
PmiSKL
Ppy	G...GKS K KL
LnoSKL
Ppe1	GKS K SKAKL
PhgAKL
GRSKL
YGSKL
Ppe2KS K KL

49-7C6	TNG*
78-0B10	TNG*
90-1B5	TNG*
133-1B2	TNG*
146-1H2	TNG*

FIG. 19A (Continued)

Lcr:	<i>Luciola cruciata</i>	Phg:	<i>Phengodes</i> sp.
Lla:	<i>Luciola lateralis</i>	Lmi:	<i>Luciola mingrelica</i>
Gr:	<i>Pyrophorus plagiophthalmus</i>	(green)	
YG:	<i>Pyrophorus plagiophthalmus</i>	(yellow green)	
Pmi:	<i>Pyrocoelia miyako</i>	Ppy:	<i>Photinus pyralis</i>
Lno:	<i>Lampyris noctiluca</i>		
Ppe-2:	<i>Photuris pennsylvanica</i> (2)		
Ppe-1:	<i>Photuris pennsylvanica</i> (1)		
Cons:	Sites which are strictly conserved		

FIG. 19A (Continued)

	1	50
Lcr	MENMENDE.N	IVVGPKPFYP
Lla	MENMENDE.N	IVYGPEPFYP
Lmi	ME.MEKES.N	VVYGPLPFYP
Pmi	...MEDDSKH	IMHGRHSIL
Ppy	...MED.AKN	IKKGPAFPYP
Lno	...MED.AKN	IMHGPAPFYP
Ppe1	...MSI.ENN	ILIGPPPYP
Ppe2	...MED..KN	ILYGPEPFYP
Phg	MIKME..EEH	VMPGAMPRDL
GR	...MMKREKN	VVYGPPEPLHP
YG	...MMKREKN	VIYGPEPLHP
30-4B02	...MMKREKN	VIYGPEPLHP
81-6G01	...MMKREKN	VIYGPEPLHP
Cons	---	M-----G-----AG-----A-----

	51	100
Lcr	TGVDYSYAEY	LEKSCCLGKA
Lla	TGVDYTYAEY	LEKSCCLGEA
Lmi	TGVDISYQEY	FDITCRLAEA
Pmi	AEVNITYSEY	FEMCRLAET
Ppy	IEVNITYAEY	FEMSVRLAEA
Lno	AEVNITYSEY	FEMACRLAET
Ppe1	TELEVTYKEF	LDVTCRLAEA
Ppe2	TKENVLYKEF	LKLSCRLAES
Phg	THEIISYAKI	LDMSCRLAVS
GR	GEEWISYKEF	FETCILLAQS
YG	GDESLSYKEF	FEATCILLAQS
30-4B02	GDESLSYKEF	FEATVILLAQS
81-6G01	GDESLSYKEF	FEATVILLAQS
Cons	-----Y-----	L-----G-----C-E---FF-P-----

	101	150
Lcr	FIGVGVAPTN	EIYTLRELVH
Lla	FIGVGVAPTN	EIYTLRELVH
Lmi	YIGVAVAPTN	EIYTLRELNH
Pmi	FIGVGVAPTN	DIYNERELYN
Ppy	FIGVAVAPAN	DIYNERELLN
Lno	FIGVGVASTN	DIYNERELYN
Ppe1	YGVVATAPTN	DIYNERELYN
Ppe2	YLGIIAAPVS	DKYIERELIH
Phg	YLGITVATVN	DTYTDRSELSE
GR	YIGMIVAPVN	EGYIPDELCK
YG	YIGMIVAPVN	ESYIPDELCK
30-4B02	YIGMIVAPVN	ESYIPDELCK
81-6G01	YIGMIVAPVN	ESYIPDELCK
Cons	--G---A---	--Y---EL---I---P---

21/70

151

200

Lcr	KTIVILD SKV	DYRGYQCLDT	FIKRNTPPGF	QASSFKTVEV	.DRKEQVALI
Lla	KTIVILD SKV	DYRGYQSMDN	FIKKQNTPGF	KGSSFKTVEV	.NRKEQVALI
Lmi	KKIVILD SKV	NFGGHDCMET	FIKKHVELGF	QPSSFVPIDV	KNRKQHALL
Pmi	QKIVILD SRE	DYMGKQSMYS	FIESHLPAGF	NEYDYIPDSF	.DRETATALI
Ppy	QKIIIMDSKT	DYQGFQSMYT	FVTSHLPPGF	NEYDFVPESF	.DRDKTIALI
Lno	QKIVILD SRE	DYMGKQSMYS	FIESHLPAGF	NEYDYIPDSF	.DRETATALI
Ppe1	KKIIILDGKK	DYLGYQSMQS	FMKEHVPANF	NVS AFKPLSF	.DLDR.VACI
Ppe2	ETIIILDNE	DLGGYQCLNN	FISQNSDINL	DVKFKPNSF	.NRDDQVALV
Phg	QKLLIIDSMQ	DIGGIECVHS	FVSRYTDEHF	DPLKFVPLDF	.DPREQVALI
GR	KRIIILDAVE	NIHGCESLPN	FISRYSDGNI A.	.NFKPLHY	.DPVEQVAII
YG	KRIIILDTVE	NIHGCESLPN	FISRYSDGNI A.	.NFKPLHY	.DPVEQVAII
30-4B02	KRIIILDTVE	NIHGCESLPN	FISRYSDGNI A.	.NFKPLHY	.DPVEQVAII
81-6G01	KRIIILDTVE	NIHGCESLPN	FISRYSDGNI A.	.NFKPLHF	.DPVEQVAII
Cons	-----I-D-----	-----G-----	F-----	-----A-----	

201

250

Lcr	MNSSGSTGLP	KGVQLTHENT	VTRFSHARDP	IYGNQVSPGT	AVLTVVPFHH
Lla	MNSSGSTGLP	KGVQLTHENA	VTRFSHARDP	IYGNQVSPGT	AILTVVPFHH
Lmi	MNSSGSTGLP	KGVRITHEGA	VTRFSHAKDP	IYGNQVSPGT	AILTVVPFHH
Pmi	MNSSGSTGLP	KGVDLTHMNV	CVRFSHCRDP	VFGNQIIPDT	AILTVIPFH
Ppy	MNSSGSTGLP	KGVALPHRTA	CVRFSHARDP	IFGNQIIPDT	AILSVVPPFH
Lno	MNSSGSTGLP	KGVELTHQNV	CVRFSHCRDP	VFGNQIIPDT	AILTVIPFH
Ppe1	MNSSGSTGLP	KGVPISHRNT	IYRFSHCRDP	VFGNQIIPDT	TILCAVPPFH
Ppe2	MFSSGTTGVS	KGVMLTHKNI	VARFSHCKDP	TFGNAINPTT	AILTVIPFH
Phg	MTSSGTTGLP	KGVMLTHRNI	CVRFVHSRDP	LFGTRFIPET	SILSLVPPFH
GR	LCSSGTTGLP	KGVMQTHRNV	CVRLIHALDP	RVGTQLIPGV	TVLVYLPFFH
YG	LCSSGTTGLP	KGVMQTHQNI	CVRLIHALDP	RAGTQLIPGV	TVLVYLPFFH
30-4B02	LCSSGTTGLP	KGVMQTHQNI	CVRLIHALDP	RAGTQLIPGV	TVLVYLPFFH
81-6G01	LCSSGTTGLP	KGVMQTHQNI	CVRLIHALDP	RAGTQLIPGV	TVLVYLPFFH
Cons	--SSG-TG--	KGV---H---	--R--H--DP	--G----P--	--L---PF-H

251

300

Lcr	GFGMPTTLGY	LICGFRVVML	TKFDEETFLK	TLQDYKCTSV	ILVPTLFAIL
Lla	GFGMPTTLGY	LTCGFRIVML	TKFDEETFLK	TLQDYKCSV	ILVPTLFAIL
Lmi	GFGMPTTLGY	FACGYRVVML	TKFDEELFLR	TLQDYKCTSV	ILVPTLFAIL
Pmi	VFQMFPTTLGY	LTCGFRIVLM	YRFEEELFLR	SLQDYKIQSA	LLVPTLFSFF
Ppy	GFGMPTTLGY	LICGFRVVML	YRFEEELFLR	SLQDYKIQSA	LLVPTLFSFF
Lno	GFGMPTTLGY	LTCGFRIVLM	YRFEBELFLR	SLQDYKIQSA	LLVPTLFSFF
Ppe1	AFGTFTNLGY	LICGFHVVL	YRFNEHLFLQ	TLQDYKCQSA	LLVPTVLAFL
Ppe2	GFGMFTTLGY	FTCGFRVALM	HTFEEKLFLQ	SLQDYKVEST	LLVPTLMAFF
Phg	AFGMFTTLSY	FIVGLKIVMM	KRFDGELFLK	TIQNYKIPTI	VIAPPVMVFL
GR	AFGFSINLGY	FMVGLRVIML	RRFDQEAFLK	AIQDYEVRSV	INVPAIILFL
YG	AFGFSINLGY	FMVGLRVIML	RRFDQEAFLK	AIQDYEVRSV	INVPAIILFL
30-4B02	AFGFSINLGY	FMVGLRVIML	RRFDQEAFLK	AIQDYEVRSV	INVPAIILFL
81-6G01	AFGFSITLGY	FMVGLRVIMF	RRFDQEAFLK	AIQDYEVRSV	INVPSVILFL
Cons	-F-----L-Y	--G-----	--F-----FL-	--Q-Y-----	--P-----

FIG. 19B (Continued)

301

350

Lcr	NKSELLNKYD	LSNLVEIASG	GAPLSKEVGE	AVARRFNLPG	VRQGYGLTET
Lla	NRSELLDKYD	LSNLVEIASG	GAPLSKEIGE	AVARRFNLPG	VRQGYGLTET
Lmi	NKSELIDKFD	LSNLTEIASG	GAPLAKEVGE	AVARRFNLPG	VRQGYGLTET
Pmi	AKSTLVDKYD	LSNLHEIASG	GAPLAKEVGE	AVAKRFLKPG	IRQGYGLTET
Ppy	AKSTLIDKYD	LSNLHEIASG	GAPLSKEVGE	AVAKRFLPG	IRQGYGLTET
Lno	AKSTLVDKYD	LSNLHEIASG	GAPLAKEVGE	AVAKRFLKPG	IRQGYGLTET
Ppe1	AKNPLVDKYD	LSNLHEIASG	GAPLSKEISE	IAAKRFLKPG	IRQGYGLTET
Ppe2	AKSALVEKYD	LSHLKEIASG	GAPLSKEIGE	MVKRFLKLF	VRQGYGLTET
Phg	AKSHLVDKYD	LSSIKEIATG	GAPLGPALAN	AVAKRLKLGG	IIQGYGLTET
GR	SKSPLVDKYD	LSSLRELCCG	AAPLAKEVAE	IAVKRLNLPG	IRCGFGLTES
YG	SKSPLVDKYD	LSSLRELCCG	AAPLAKEVAE	VAVKRLNLPG	IRCGFGLTES
30-4B02	SKSPLVDKYD	LSSLRELCCG	AAPLAKEVAE	VAAKRLNLPG	IRCGFGLTES
81-6G01	SKSPLVDKYD	LSSLRELCCG	AAPLAKEVAE	VAAKRLNLPG	IRCGFGLTES
Cons	-----L--K-D	LS---E---G	-APL-----	-----R--L--	---G-GLTE-

351

400

Lcr	TSAlIITPEG	DDKPGASGKV	VPLFKAKVID	LDTKKSLGP	RRGEVCVKGP
Lla	TSAlIITPEG	DDKPGASGKV	VPLFKAKVID	LDTKKTLGP	RRGEVCVKGP
Lmi	TSAlIITPEG	DDKPGASGKV	VPLFKVKVID	LDTKKTLGV	RRGEICVKGP
Pmi	TSAlIITPEG	DDKPGACGKV	VPFFTAKVID	LDTGKTLGV	QRGELCVKGP
Ppy	TSAlIITPEG	DDKPGAVGKV	VPFFFEAKVVD	LDTGKTLGV	QRGELCVRGP
Lno	TSAlIITPEG	DDKPGACGKV	VPFFSAKVID	LDTGKTLGV	QRGELCVKGP
Ppe1	TCAIVITAEQ	EFKLGAVGKV	VPFYSLKVL	LNTGKKLGP	ERGEICFKGP
Ppe2	TS AVLITPDT	DVRPGSTGKI	VPFHAVKVD	PTTGKILGP	ETGELYFKGD
Phg	CAAVLITPHN	KIKTGSTGQV	LPYVTAKVID	TKTGKNLGP	QTGELCFKSD
GR	TSANIHSLRD	EFKSGSLGRV	TPLMAAKIAD	RETGKALGP	QVGELCIKGP
YG	TSANIHSLRD	EFKSGSLGRV	TPLMAAKIAD	RETGKALGP	QVGELCVKGP
30-4B02	TSANIHSLRD	EFKSGSIGRV	TPLMAAKIAD	RETGKALGP	QVGELCIKGP
81-6G01	TSANIHSLRD	EFKSGSLGRV	TPLMAAKIAD	RETGKALGP	QVGELCIKGP
Cons	--A-----	-----G--G--	-P----K--D	--T-K-LG-N	--GE-----

401

450

Lcr	MLMKGYVNPN	EATKELIDEE	GWLHTGDIY	YDEEKHFFIV	DRLKSLIKYK
Lla	MLMKGYVDNP	EATREIIDE	GWLHTGDIY	YDEEKHFFIV	DRLKSLIKYK
Lmi	SLMLGYSNNP	EATRETIIDEE	GWLHTGDIY	YDEEKHFFIV	DRLKSLIKYK
Pmi	MIMKGYVNPN	EATNALIDKD	GWLHSGDIAY	YDKDGHFFIV	DRLKSLIKYK
Ppy	MIMSGYVNPN	EATNALIDKD	GWLHSGDIAY	WDEDEHFFIV	DRLKSLIKYK
Lno	MIMKGYVNPN	EATSALIDKD	GWLHSGDIAY	YDKDGHFFIV	DRLKSLIKYK
Ppe1	MIMKGYINNP	EATRELIDEE	GWIHSGDIY	FDEDGHFYIV	DRLKSLIKYK
Ppe2	MIMKSYYNNE	EATKAIINKD	GWLHSGDIAY	YDNDGHFYIV	DRLKSLIKYK
Phg	IIMKGYYQNE	EETRLVIDKD	GWLHSGDIY	YDTDGNFHIV	DRLKELIKYK
GR	MVSKGYVN	EATKEAIDDD	GWLHSGDFGY	YDEDEHFYVV	DRYKELIKYK
YG	MVSKGYVN	EATKEAIDDD	GWLHSGDFGY	YDEDEHFYVV	DRYKELIKYK
30-4B02	MVSKGYVN	EATKEAIDDD	GWLHSGDFGY	YDEDEHFYVV	DRYKELIKYK
81-6G01	MVSKGYVN	EATKEAIDDD	GWLHSGDFGY	YDEDEHFYVV	DRYKELIKYK
Cons	-----Y--N-	E-T---I---	GW---GD--Y	-D-----V	DR-K-LIKYK

FIG. 19B (Continued)

23/70

	451	500
Lcr	GYQVPPAEL SVLLQHPSIF DAGVAGVPDP VAGELPGAVV VLESGKNMTE	
Lla	GYQVPPAEL SVLLQHPNIF DAGVAGVPDP IAGELPGAVV VLEKGKSMTE	
Lmi	GYQVPPAEL SVLLQHPNIF DAGVAGVPDP DAGELPGAVV VMEKGKTMTE	
Pmi	GYQVPPAEL SILLQHPPIF DAGVAGIPDP DAGELPAAVV VLEEGKMMTE	
Ppy	GYQVAPAELE SILLQHPNIF DAGVAGLPDD DAGELPAAVV VLEHGKTMTE	
Lno	GYQVPPAEL SILLQHPPIF DAGVAGIPDP DAGELPAAVV VLEEGKTMTE	
Ppe1	GYQVPPAEL ALLLQHPPFIE DAGVAGVPDE VAGDLPAGAVV VLKEGKSITE	
Ppe2	GYQVAPAEIE GILLQHPPYIV DAGVTGIPDE AAGELPAAGV VVQTGKYLNE	
Phg	AYQVAPAELE ALLLQHPPYIA DAGVTGIPDE EAGELPAACV VLEPGKTMTE	
GR	GSQVAPAELE EILLKNPCIR DVAVVGIPDL EAGELPSAFV VIQPGKEITA	
YG	GSQVAPAELE EILLKNPCIR DVAVVGIPDL EAGELPSAFV VKQPGKEITA	
30-4B02	GSQVAPAELE EILLTNPCIR DVAVVGIPDL EAGELPSAFV VKQPGKEITA	
81-6G01	GSQVAPAELE EILLKNPCIR DVAVVGIPDL EAGELPSAFV VKQPGKEITA	
Cons	--QV-PAE-E --LL--P-I- D--V-G-PD- -AG-LP-A-V V---GK---	

	501	550
Lcr	KEVMDYVASQ VSNAKRLRGG VRFVDEVPKG LTGKIDGRA. IREILKRPV.	
Lla	KEVMDYVASQ VSNAKRLRGG VRFVDEVPKG LTGKIDGKA. IREILKRPV.	
Lmi	KEIVDYVNSQ VVNHKRLRGG VRFVDEVPKG LTGKIDAKV. IREILKRPQ.	
Pmi	QEVMGYVAGQ VTASKRLRGG VKFVDEVPKG LTGKIDSRK. IREILTMGQK	
Ppy	KEIVDYVASQ VTAKKLRGG VVFVDEVPKG LTGKLDARK. IREILIKAKK	
Lno	QEVMGYVAGQ VTASKRLRGG VKFVDEVPKG LTGKIDGRK. IREILMMGKK	
Ppe1	KEIQDYVAGQ VTSSKKLRGG VEFVKEVPKG FTGKIDTRK. IKEILIKAQK	
Ppe2	QIVQNFVSSQ VSTAKWLRGG VKFLDEIPKG STGKIDRKV. LRQMPEKH..	
Phg	KEVMDYIAER VTPTKRLRGG VLFVNNIPKG ATGKLRVTE. LRRLLTQRA.	
GR	KEVYDYLAER VSHTKYLRRGG VRFVDSIPRN VTGKITRKE LKQLLEKS..	
YG	KEVYDYLAER VSHTKYLRRGG VRFVDSIPRN VTGKITRKE LKQLLEKS..	
30-4B02	KEVYDYLAER VSHTKYLRRGG VRFVDSIPRN VTGKITRKE LKQLLEK...	
81-6G01	KEVYDYLAER VSHTKYLRRGG VRFVDSIPRN VTGKITRKE LKQLLEK...	
Cons	----- V---K-LRGG V-F---P-- -TGK-----	

	551
LcrAKM
LlaAKM
LmiAKM
PmiSKL
Ppy	G...GKSCL
LnoSKL
Ppe1	GKS SKS KAKL
Ppe2KSCL
PhgAKL
GRSKL
YGSKL
30-4B02AGG*
81-6G01AGG*
Cons	-----K-

FIG. 19B (Continued)

24/70

1

50

Lcr	MENMENDE.N	IVVGPKPFYP	IIEGSAGTQL	RKYMERYAKL	.GAIATNAV
Lla	MENMENDE.N	IVYGPEPFYP	IIEGSAGAQL	RKYMDRYAKL	.GAIATNAL
Lmi	ME.MEKEE.N	VVYGPLPFYP	IIEGSAGIQL	RKYMHQYAKL	.GAIAFSNAL
Pmi	...MEDDSKH	IMHGHRHSIL	LEDGTAGEQL	HKAMKRYAQV	PGTIAFTDAH
Ppy	...MED.AKN	IKKGPAFPYP	LEDGTAGEQL	HKAMKRYALV	PGTIAFTDAH
Lno	...MED.AKN	IMHGPAFPYP	LEDGTAGEQL	HKAMKRYAQV	PGTIAFTDAH
Ppe1	...MSI.ENN	ILIGPPPYP	LEEGTAGEQL	HRAISRYAAV	PGTLAYTDVH
Ppe2	...MED..KN	ILYGPEPFYP	LADGTAGEQM	FYALSRYADI	SGCIALTNAH
Phg	MIKME..EEH	VMPGAMPRDL	LFEGTAGQQL	HRALYKHSYF	PE..AIVDSH
YG	...MMKREKN	VIYGPEPLHP	LEDLTAGEML	FRALRKHSHL	PQ..ALVDVF
	---	XXOOOOOO	XOO-OOOXO	OXXO--OXO	XOOXOXXXOO
	---	OXOO-XX			

51

100

Lcr	TGVDYSYAEY	LEKSCLLGKA	LQNYGLVVVG	RIALCSENCE	EFFIPVIAGL
Lla	TGVDYTYAEY	LEKSCLGEA	LKNYGLVVVG	RIALCSENCE	EFFIPVLAGL
Lmi	TGVDISYQEY	FDITCRLAEEA	MKNFGMKPEE	HIALCSENCE	EFFIPVLAGL
Pmi	AEVNITYSEY	FEMSCRRAET	MKRYGLGLQH	HIAVCSETSL	QFFMPVCGAL
Ppy	IEVNITYAEY	FEMSVRLAEEA	MKRYGLNTNH	RIVVCSSENSL	QFFMPVLGAL
Lno	AEVNITYSEY	FEMACRLAET	MKRYGLGLQH	HIAVCSENSL	QFFMPVCGAL
Ppe1	TELEVTYKEF	LDVTCRRLAEEA	MKNYGLGLQH	TISVCSENCV	QFFMPICAAL
Ppe2	TKENVLYEEF	LKLSCRLLAES	FKKYGLKQND	TIAVCSENGL	QFFLPLIASL
Phg	THEIISYAKI	LDMSCRLLAVS	FQKYGLTQNN	IIGICSENNL	NFFNPVIAAF
YG	GDESLSYKEF	FEATCLLAQS	LHNCGYKMND	VVSICAENN	FFFPIIAAW
	---	XOXOXX--X	XOOXOXOOX-	XXOX-XOOOO	OXO-X-OOX X--O-XOOX

101

150

Lcr	FIGVGVAPTN	EIYTLRELVH	SLGISKPTIV	FSSKKGLDKV	ITVQKTVTTI
Lla	FIGVGVAPTN	EIYTLRELVH	SLGISKPTIV	FSSKKGLDKV	ITVQKTVATI
Lmi	YIGVAVAPTN	EIYTLRELNH	SLGIAQPTIV	FSSRKGLPKV	LEVQKTVTCI
Pmi	FIGVGVAPTN	DIYNERELYN	SLFISQPTIV	FCSKRALQKI	LGVQKKLPVI
Ppy	FIGVAVAPAN	DIYNERELLN	SMNISQPTVV	FVSKKGLQKI	LNVQKKLPPI
Lno	FIGVGVASTN	DIYNERELYN	SLSISQPTIV	SCSKRALQKI	LGVQKKLPPI
Ppe1	YVGVATAPTN	DIYNERELYN	SLSISQPTVV	FTSRNSLQKI	LGVQSRLPPI
Ppe2	YLGIIIAAPVS	DKYTERELIH	SLGIVKPRII	FCSKNTFQKV	LNVKSKLKYV
Phg	YLGITVATVN	DTYTDRELSE	TLNITKPQML	FCSKQSLPIV	MKTMKIMPYV
YG	YIGMIVAPVN	ESYIPDELCK	VMGISKPQIV	FCTKNILNKV	LEVQSRTNFI
	---	XO-XOO-OXO	XX-OXX--OO	XXO-OX-XOO	OOXOOOOXOO

FIG. 19C

151

200

Lcr	KTIVILD8KV DYRGYQCLDT FIKRNTPPGF QASSFKTVEV	.DRKEQVALI
Lla	KTIVILD8KV DYRGYQSMDN FIKKNTPQGF KGSSFKTVEV	.NRKEQVALI
Lmi	KKIVILD8KV NFGGHDCMET FIKKHVELGF QPSSFVPIDV KNRKQHVALL	
Pmi	QKIVILD8RE DYMKGQSMYS FIESHLPAAGF NEYDYIPDSF	.DRETATALI
Ppy	QKIIIMD8KT DYQGFQSMYT FVTSHLPPGF NEYDFVPESF	.DRDKTIALI
Lno	QKIVILD8RE DYMKGQSMYS FIESHLPAAGF NEYDYIPDSF	.DRETATALI
Ppe1	KKIIILDGKK DYLGYQSMQS FMKEHVPANF NVSAFKPLSF	.DLDR. VACI
Ppe2	ETIIILDLNE DLGGYQCLNN FISQNSDINL DVKFKPNSF	.NRDDQVALV
Phg	QKLLIID8MQ DIGGIECVHS FVSRYTDEHF DPLKFVPLDF	.DPREQVALI
YG	KR IIIILD TVE NIHGCESLPN FISRYSDGNI A..NFKPLHY	.DPVEQVAI

OXOX-O-XXO XXO-OXOXOO -OOOXOXOXX O--OOOOOXX -OXOOOO-XO

201

250

Lcr	MNSSG5TGLP KGVLQTHENT VTRFSHARDP IYGNQVSPGT	AVLTVVPFH
Lla	MNSSG5TGLP KGVLQTHENA VTRFSHARDP IYGNQVSPGT	AILTVVPFH
Lmi	MNSSG5TGLP KGVRITHEGA VTRFSHARDP IYGNQVSPGT	AILTVVPFH
Pmi	MNSSG5TGLP KGVDLTHHMNV CVRFSHCRDP VFGNQIIPDT	AILTVIPFH
Ppy	MNSSG5TGLP KGVALPHRTA CVRFSHARDP IFGNQIIPDT	AILSVVPFH
Lno	MNSSG5TGLP KGVELTHQNV CVRFSHCRDP VFGNQIIPDT	AILTVIPFH
Ppe1	MNSSG5TGLP KGVPISHRNT IYRFSHCRDP VFGNQIIPDT	TILCAVPFH
Ppe2	MFSSGTTGVS KGVMLTHKNI VARFSHCRDP TFGNAINPTT	AILTVIPFH
Phg	MFSSGTTGGLP KGVMLTHRNI CVRFVHSRDP LGTRFIPET	SILSLVPFH
YG	LCSSGTTGLP KGVMQTHQNI CVRLIHALDP RAGTQLIPGV	TVLVYLPFFF

XX---X--OO ---OXO-000 OO-XX-OX-- OX-XOXO-OX XX-XXX--X-

251

300

Lcr	GFGMFTTLGY LICGFRVVML TKFDEETFLK TLQDYKCTSV	ILVPTLFAIL
Lla	GFGMFTTLGY LTCGFRIVML TKFDEETFLK TLQDYKCSSV	ILVPTLFAIL
Lmi	GFGMFTTLGY FACGYRVVML TKFDEELFLR TLQDYKCTSV	ILVPTLFAIL
Pmi	VFQMFTTLGY LTCGFRIVLM YRFEEELFLR SLQDYKIQSA	LLVPTLFSFF
Ppy	GFGMFTTLGY LICGFRVVLM YRFEEELFLR SLQDYKIQSA	LLVPTLFSFF
Lno	GFGMFTTLGY LTCGFRIVLM YRFEEELFLR SLQDYKIQSA	LLVPTLFSFF
Ppe1	AFGFTFTNLGY LICGFRVVLM YRFNEHFLQ TLQDYKCQSA	LLVPTVLAFL
Ppe2	GFGMTTTLGY FTCGFRVALM HTFEELFLQ SLQDYKVEST	LLVPTLMAFF
Phg	AFGMFTTSLY FIVGLKIVMM KRFDGELFLK TIQNYKIPTI	VIAPPVMVFL
YG	AFGFSINLGY FMVGLRVIML RRFQEAFLK AIQDYEVRSV	INVPAILFL

X-OXXX-O- XOX-XOXXX OO-OXOX--O XX-O-X0000 XXO-XXXOO

FIG. 19C (Continued)

301

350

Lcr	NKSELLENKYD	LSNLVLEIASG	GAPLSKEVGE	AVARRFNLPG	VRQGYGLTET
Lla	NRSELLDKYD	LSNLVLEIASG	GAPLSKEIGE	AVARRFNLPG	VRQGYGLTET
Lmi	NKSELIDKFD	LSNLTEIASG	GAPLAKEVGE	AVARRFNLPG	VRQGYGLTET
Pmi	AKSTLVDKYD	LSNLHEIASG	GAPLAKEVGE	AVAKRFLPG	IRQGYGLTET
Ppy	AKSTLIDKYD	LSNLHEIASG	GAPLSKEVGE	AVAKRFHLPG	IRQGYGLTET
Lno	AKSTLVDKYD	LSNLHEIASG	GAPLAKEVGE	AVAKRFLPG	IRQGYGLTET
Ppe1	AKNPLVDKYD	LSNLHEIASG	GAPLSKEISE	IAAKRFLPG	IRQGYGLTET
Ppe2	AKSALVEKYD	LSHLKEIASG	GAPLSKEIGE	MVKRFLKLF	VRQGYGLTET
Phg	AKSHLVDKYD	LSSIKEIATG	GAPLGPALAN	AVAKRLKLG	IIQGYGLTET
YG	SKSPLVDKYD	LSSLRELCCG	AAPLAKEVAE	VAVKRLNLPG	IRCGFGLTES

X000-00-0- --X00-XXX- X---X000XO XXX0-XX-00 OOX-X---X

351

400

Lcr	TSAILITPEG	DDKPGASGKV	VPLFKAKVID	LDTKKSLGP	RRGEVCVKGP
Lla	TSAILITPEG	DDKPGASGKV	VPLFKAKVID	LDTKKTLGP	RRGEVCVKGP
Lmi	TSAILITPEG	DDKPGASGKV	VPLFKVKVID	LDTKKTLGV	RRGEICVKGP
Pmi	TSAILITPEG	DDKPGACGKV	VPFFTAKIVD	LDTGKTLGV	QRGELCVKGP
Ppy	TSAILITPEG	DDKPGAVGKV	VPFFFEAKVVD	LDTGKTLGV	QRGELCVRGP
Lno	TSAILITPEG	DDKPGACGKV	VPFFSAKIVD	LDTGKTLGV	QRGELCVKGP
Ppe1	TCAIVITAE	EFKIGAVGKV	VPFFYSLKVID	LNTGKKLGP	ERGEICFKGP
Ppe2	TSAVLITPDT	DVRPGSTGKI	VPFHAVKVV	PTTGKILGP	ETGELYPKGD
Phg	CAVLITPHN	KIKTGSTGQV	LPYVTAKIVD	TKTGKNLGP	QTGELCFKSD
YG	TSANIHSLGD	EFKSGSLGRV	TPLMAAKIAD	RETGKALGP	QVGEICVKGP

OO-X0XXXX XXOX-XO-XO X-XXOO-XX- XX-O-X--O- OX--000000

401

450

Lcr	MLMKGYVN	EATKELIDE	GWLHTGDIY	YDEEKHFFIV	DRLKSLIKYK
Lla	MLMKGYVD	EATREIIDE	GWLHTGDIY	YDEEKHFFIV	DRLKSLIKYK
Lmi	SLMLGYSN	EATRETIIDE	GWLHTGDIY	YDEDEHFFIV	DRLKSLIKYK
Pmi	MIMKGYVN	EATNALID	GWLHSGDIAY	YDKDGHFFIV	DRLKSLIKYK
Ppy	MIMSGYVN	EATNALID	GWLHSGDIAY	WDEDEHFFIV	DRLKSLIKYK
Lno	MIMKGYVN	EATSALID	GWLHSGDIAY	YDKDGHFFIV	DRLKSLIKYK
Ppe1	MIMKGYIN	EATRELIDE	GWIHSGDIY	FDEDGHVYIV	DRLKSLIKYK
Ppe2	MIMKSYYN	EATKAIINK	GWLRSGDIAY	YDNDGHFYIV	DRLKSLIKYK
Phg	IIMKGYYQ	EETRLVID	GWLHSGDIY	YDTDGNFHIV	DRLKELIKYK
YG	MVKSGYVN	EATKEAID	GWLHSGDFGY	YDEDEHFFV	DRYKELIKYK

OXOO-00-X -0-0OX-0XO --000--XO- O-0OXOOX- --X-X----

FIG. 19C (Continued)

27/70

451

500

Lcr	GYQVPPAEL E SVLLQHPSIF DAGVAGVPDP VAGELPGAVV VLESGKNMTE
Lla	GYQVPPAEL E SVLLQHPNIF DAGVAGVPDP IAGELPGAVV VLEKGKSMTE
Lmi	GYQVPPAEL E SVLLQHPNIF DAGVAGVPDP DAGELPGAVV VMEKGKTMT
Pmi	GYQVPPAEL E SILLQHPPIF DAGVAGIPDP DAGELPAAVV VLEEGKMMTE
Ppy	GYQVAPAELE SILLQHPPIF DAGVAGLPDD DAGELPAAVV VLEHGKTMT
Lno	GYQVPPAEL E SILLQHPPIF DAGVAGIPDP DAGELPAAVV VLEEGKTMT
Ppe1	GYQVPPAEL E ALLLQHPPFIE DAGVAGVPDE VAGDLPGAVV VLKEGKSITE
Ppe2	GYQVAPAEIE GILLQHPPYIV DAGVTGIPDE AAGELPAAGV VVQTGKYLNE
Phg	AYQVAPAELE ALLLQHPPYIA DAGVTGIPDE EAGELPAACV VLEPGKTMT
YG	GSQVAPAELE EILLKNPCIR DVAVVGIPDL EAGELPSAFV VKOPGKEITA

OX--X---O- XO--XX-O-X -XX-X-O--X O--O--X-X- -XXO--OXOX

501

550

Lcr	KEVMDYVASQ VSNAKRLRGG VRFVDEVPKG LTGKIDGRA. IREILKPKV.
Lla	KEVMDYVASQ VSNAKRLRGG VRFVDEVPKG LTGKIDGKA. IREILKPKV.
Lmi	KEIVDYVNSQ VVNHKRLRGG VRFVDEVPKG LTGKIDAKV. IREILKPKQ.
Pmi	QEVMQDYVAGQ VTASKRLRGG VKFVDEVPKG LTGKIDSRK. IREILTMGQK
Ppy	KEIVDYVASQ VTTAKKLRGG VVFVDEVPKG LTGKLDARK. IREILIKAKK
Lno	QEVMQDYVAGQ VTASKRLRGG VKFVDEVPKG LTGKIDGRK. IREILMMGKK
Ppe1	KEIQDYVAGQ VTSSEKKLRGG VEFVKEVPKG FTGKIDTRK. IKEILIIAQK
Ppe2	QIVQNFVSSQ VSTAKWLRGG VKFLDEIPKG STGKIDRKV. LRQMFKEH..
Phg	KEVMDYIAER VTPTRKLRRGG VLFVNNIPKG ATGKLVRTE. LRRLLTQRA.
YG	KEVYDYLAER VSHTKYLRRGG VRFVDSIPRN VTGKITRKE LKQOLLEKS..

OOOXOOXOX -OOO-X---- -O-OOX-XX X---OXOXOO XXXXOOOOOO

551

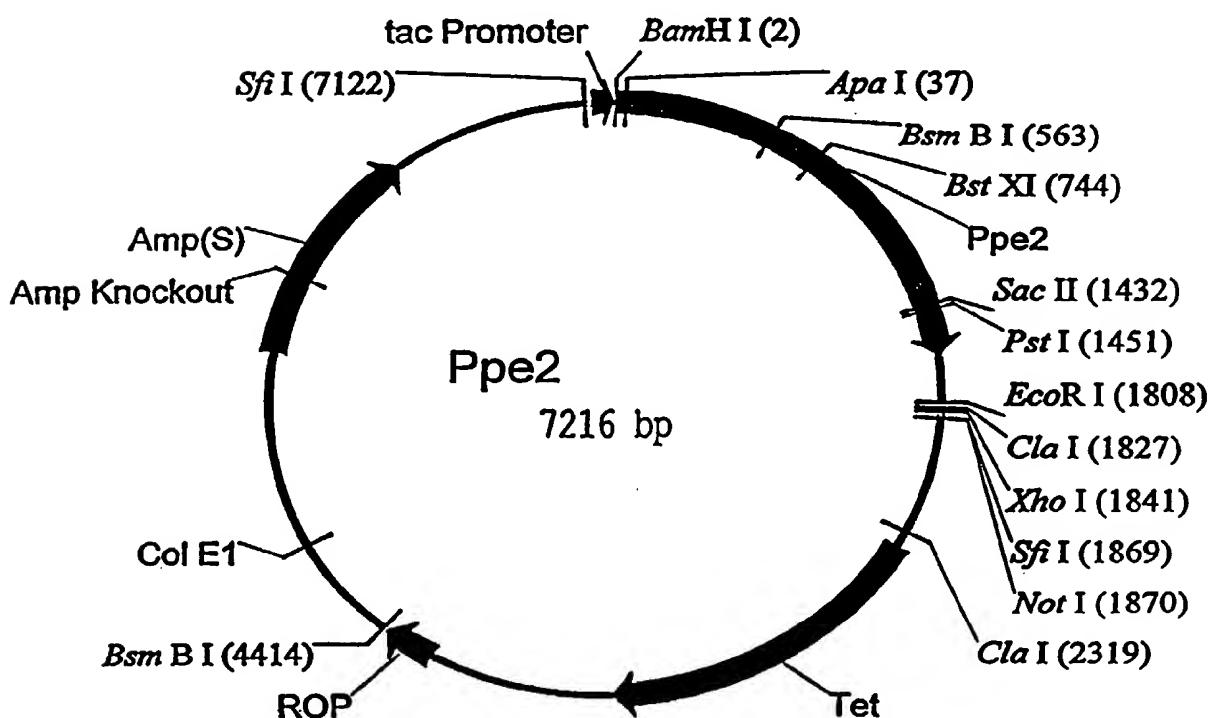
LcrAKM
LlaAKM
LmiAKM
PmiSKL
Ppy	G....GKS KL
LnoSKL
Ppe1	GKS KSK KAKL
Ppe2KS KL
PhgAKL
YGSKL

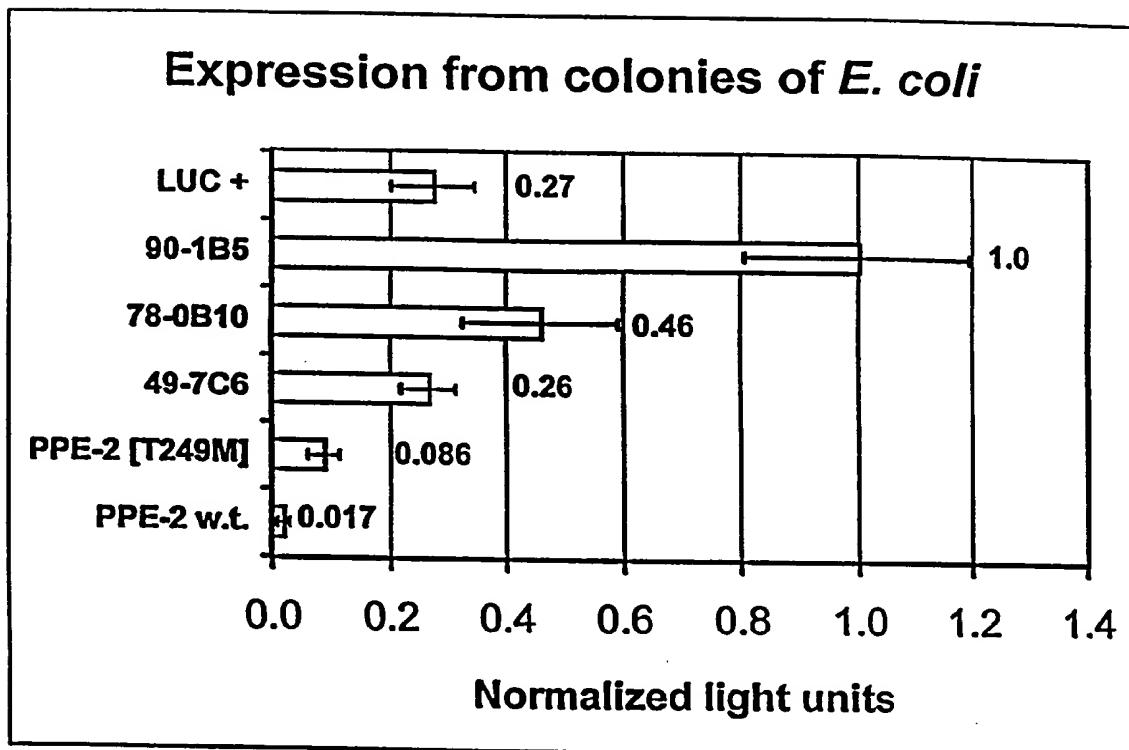
OOOOOOX-O

FIG. 19C (Continued)

Key to beetle Luciferase sequences:

Lcr	<i>Luciola cruciata</i>
Lla	<i>Luciola lateralis</i>
Lmi	<i>Luciola mingrellica</i>
Pmi	<i>Pyrocoelia miyako</i>
Ppy	<i>Photinus pyralis</i>
Lno	<i>Lampyris noctiluca</i>
Ppc1	<i>Photuris pennsylvanica</i> (1)
Ppc2	<i>Photuris pennsylvanica</i> (2)
Phg	<i>Phengodes</i> sp.
YG	<i>Pyrophorus plagiophthalmus</i> – yellow-green luminescence

FIG. 19D**FIG. 20**

**FIG. 21**

luc49-7C6 (SEQ ID NO:1)

GGATCCAATG	GCAGATAAAA	ATATTTATA	TGGGCCGAA	CCATTTATC	CCTGGCTGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTACGC	ATTATCTCGT	TATGCAGATA	TTTCAGGATG	120
CATAGCATTG	ACAATGCTC	ATACAAAAGA	AAATGTTTA	TATGAAGAGT	TTTAAAATT	180
GTCGTGTCGT	TTAGCGAAA	GTTTAAAAAA	GTATGGATTA	AAACAAAACG	ACACAATAGC	240
GGTGTGTCAG	AAAAATGGTT	TGCAATTTC	CCTTCCTATA	ATTGCATCAT	TGTATCTTGG	300
ATAATTGCA	GCACCTGTTA	GTGATAAATA	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGTAAAA	CCACGCATAA	TTTTTGTC	CAAGAATACT	TTTCAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATATG	TAGAAACTAT	TATTATATA	GACTTAAATG	AAGACTTAGG	480
AGGTTATCAA	TGCTCAACA	ACTTTATTC	TCAAAATTCC	GATATTAAATC	TGACGCTAAA	540
AAAATTAAA	CCATATTCTT	TTAATCGAGA	CGATCAGGTT	GCCTGGTAA	TGTTTCTTC	600
TGGTACAAC	GGTGTTCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGACGATT	660
TTCTCTGCA	AAAGATCCTA	CTTTGGTAA	CGCAATTAAAT	CCAACGACAG	CAATTAAAC	720
GGTAATACCT	TTCCACCATG	GTTTGGTAT	GATGACCACA	TTAGGATACT	TTACTTGTGG	780
ATTCCGAGTT	GTCTAATGC	ACACGTTGA	AGAAAAACTA	TTTCTACAAT	CATTACAAGA	840
TTATAAGTG	GAAGTACTT	TACTTGTACC	AACATTAATG	GCATTTCTTG	CAAAAGTGC	900
ATTAGTTGAA	AAAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	GC3CACCTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTAAAAAA	ACGGTTAAA	TTAAACTTTG	TCAGGCAGG	1020
GTATGGATTA	ACAGAAACCA	CTTCGGCTGT	TTTAATTACA	CCGAACAATG	ACGTCAGACC	1080
GGGATCAACT	GGTAAAATAG	TACCAATTCA	CGCTGTTAAA	GTTGTCGATC	CTACAACAGG	1140
AAAATTTTG	GGGCCAAATG	AACTGGAGA	ATTGTATT	AAAGGGGACA	TGATAATGAA	1200
AGGTTATTAT	AATAATGAAG	AAGCTACTAA	AGCAATTATT	AACAAAGACG	GATGGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATT	TATATTGTGG	ACAGGCTGAA	1320
GTCATTAAATT	AAATATAAAAG	GTTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGGGGCGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAATAT	CTAAACGAAC	AAATCGTACA	1500
AAATTGTTGTT	TCCAGTCAAG	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATT	1560
GGATGAAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAAACAC	<u>ACCAATGGG</u>					1639

FIG. 22

luc49-6C10 (SEQ ID NO: 2)

GGATCCAATG	GAAGATAAAA	ATATTTTATA	TGGACCTGAA	CCATTTTATC	CCTTGGCTGAA	60
TGGGACGGCT	GGAGAACAGA	TGTTTTACCG	ATTATCTCGT	TATGCAGATA	TTTCAGGATG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGC	CCCTGTTTA	TATGAAGAGT	TGTTAAAATT	180
GTCGTGTCGT	TTAGCGAAA	GTTTAAAAAA	GTATGGATTAA	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	GAAAATGGTT	TGCAATTTT	CCTTCCTATA	ATTGCATCAT	TGTATCTTGG	300
AATAATTGCA	GCACCTGTTA	GTGATAAAAATA	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGTAAAA	CCACGCATAA	TTTTTGTGCTC	CAAGAATACT	TTTCAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATATG	TAGAAAACATAT	TATTATATTA	GACTTAAATG	AAGACTTAGG	480
AGGTATCAA	TGCCCTCAACA	ACTTTATTC	TCAAAATTCC	GATATTAAATC	TGGACGTAAA	540
AAAATTTAAA	CCATATTCTT	TTAATCGAGA	CGATCAGGTT	GCGTTGGTAA	TGTTTTCTTC	600
TGGTACAAC	GGTGTTCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGCACGATT	660
TTCTCATGCA	AAAGATCCTA	CTTTTGGTAA	CGCAATTAAAT	CCAACGACAG	CAATTAAAC	720
GGTAATACCT	TTCCACCAGT	GTTTTGGTAT	GATGACCCACA	TTAGGATAACT	TTACTTGTGG	780
ATTCGGAGTT	GTTCTAATGC	ACACGTTTGA	AGAAAAAACTA	TTTCTACAAAT	CATTACAAGA	840
TTATAAAGTG	AAAAGTACTT	TACTTGTACC	AACATTAATG	GCATTTTTG	AAAAAGTGC	900
ATTAGTTGAA	AAAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	GCGCACCTTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTAAAAAA	ACGGTTAAA	TTAAACTTTG	TCAGGCAAGG	1020
GTATGGATTA	ACAGAAACCA	CTTCGGCTGT	TTTAATTACA	CCGAACAAATG	ACGTCAGACC	1080
GGGATCAACT	GGTAAAATAG	TACCATTCA	CGCTGTTAAA	GTGTCGATC	CTACAAACAGG	1140
AAAATTTTIG	GGGCCAAATG	AAACTGGAGA	ATTGTATTT	AAAGGCGACA	TGATAATGAA	1200
AGGTIAATTAT	ATAATGAAG	AAGCTACTAA	AGCAATTATT	ACAAAGACG	GATGGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATT	TATATTGTGG	ACAGGCTGAA	1320
GTCATTAATT	AAATATAAAG	GTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATAATTGTTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGGCGGCAG	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAAATAT	CTAAACGAAC	AAATCGTACA	1500
AAATTTTGT	TCCAGTCAG	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATTTT	1560
GGATGAAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAAACAC	<u>ACCAATGGG</u>					1639

FIG. 23

luc49-0G12 (SEQ ID NO:3)

GGATCCAATG	GAAGATAAAA	ATATTTTATA	TGGACCTGAA	CCATTTTATC	CCTTGGCTGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTTACGC	ATTATCTCGT	TATGCAGATA	TTTCAGGATG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGC	CCCTGTTTTA	TATGAAGAGT	TTTTAAAATT	180
GTCGTGTCGT	TTAGCGGAAA	GTTTTAAAAA	GTATGGATTA	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	GAAAATGGTT	TGCAATTTTT	CCTTCTATA	ATTGCATCAT	TGTATCTTGG	300
AATAATTGCA	GCACCTGTTA	GTGATAAATA	CATTGAAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGAAAAA	CCACGCATAA	TTTTTGCTC	CAAGAATACT	TTTCAAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATATG	TAGAAACTAT	TATTATATTA	GACTTAAATG	AAGACTTAGG	480
AGGTTATCAA	TGCCTCAACA	ACTTTATTTC	TCAAAATTCC	GATATTAATC	TTGACGTAAC	540
AAAATTTAAA	CCATATTCTT	TTAATCGAGA	CGATCAGGTT	GCGTTGGTAA	TGTTTCTTC	600
TGGTACAAC	GGTGTTCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGACGATT	660
TTCTTATGCA	AAAGATCCTA	CTTTGGTAA	CGCAATTAAAT	CCAACGACAG	CAATTAAAC	720
GGTAATACCT	TTCCACCATG	GTTTGGTAT	GATGACCACA	TTAGGATACT	TTACTTGTGG	780
ATTCCGAGTT	GTTCTAATGC	ACACGTTGA	AGAAAAACTA	TTTCTACAAT	CATTACAAGA	840
TTATAAAGTG	GAAAGTACTT	TACTTGTACC	AACATTAATG	GCATTTCTTG	CAAAAAGTGC	900
ATTAGTTGAA	AAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	GCGCACCTTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTGAAGAAA	ACGGTTTAAA	TTAAACTTTG	TCAGGCAAGG	1020
GTATGGATT	ACAGAAACCA	CTTCGGCTGT	TTTAATTACA	CCGAACAAATG	ACGTCAGACC	1080
GGGATCAACT	GGTAAAATAG	TACCATTCA	CGCTGTTAAA	GTTGTCGATC	CTACAAACAGG	1140
AAAAATTITG	GGGCCAAATG	AAACTGGAGA	ATTGTATTTT	AAAGGCGACA	TGATAATGAA	1200
AGTTTATTAT	ATAAATGAAG	AGCTACTAA	AGCAATTATT	ACAAAGACG	GATGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATTTT	TATATTGTGG	ACAGGCTGAA	1320
GTCATTAATT	AAATATAAAAG	GTTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGGGGCGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAATAT	CTAAACGAAC	AAATCGTACA	1500
AAATTTGTT	TCCAGTCAG	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATTTT	1560
GGATGAAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAAACAC	<u>ACCAATGGG</u>					1639

FIG. 24

luc49-7A5 (SEQ ID NO:4)

GGATCCAATG	GAAGATAAAA	ATATTTATA	TGGACCTGAA	CCATTTATC	CCTTGGCTGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTTACGC	ATTATCTCGT	TATGCAGATA	TTTCAGGATG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGC	CCCTGTTTA	TATGAAGAGT	TTTTAAAATT	180
GTCGTGTCGT	TTAGCGAAA	GTTTTAAAAA	GTATGGATT	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	GAAAATGGTT	TGCAATTTTT	CCTTCCTATA	ATTGCATCAT	TGTATCTTGG	300
AATAATTGCA	GCACCTGTTA	GTGATAAATA	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGTAAAA	CCACGCATAA	TTTTTGCTC	CAAGAATACT	TTTCAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATATG	TAGAAACTAT	TATTATATA	GACTTAAATG	AAGACTTAGG	480
AGGTTATCAA	TGCCTCAACA	ACTTTATTC	TCAAAATTCC	GATATTAATC	TTGACGTAAA	540
AAAATTAAA	CCATATTCTT	TTAATCGAGA	CGATCAGGTT	GCCTTGGTAA	TGTTTTCTTC	600
TGGTACAAC	GGTGTTCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGCACGATT	660
TTCTATTGCA	AAAGATCCTA	CTTTGGTAA	CGCAATTAAT	CCAACGACAG	CAATTTAAC	720
GGTAATACCT	TTCCACCATG	GTTTGGTAT	GATGACCACA	TTAGGATACT	TTACTTGTGG	780
ATTCGGAGTT	GTTCTAATGC	ACACGTTTGA	AGAAAAACTA	TTTCTACAT	CATTACAAGA	840
TTATAAAGTG	GAAAGTACTT	TACTTGTACC	AACATTAATG	GCATTTTTG	CAAAAAGTGC	900
ATTAAGTGTAA	AAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	GCGCACCTT	960
ATCAAAAGAA	ATTGGGGAGA	TGTTGAAAAA	ACGGTTAAA	TTAAACTTTG	TCAGGCAAGG	1020
GTATGGATTA	ACAGAAACCA	CTTCGGCTGT	TTTAATTACA	CCGAACAATG	ACGTCAGACC	1080
GGGATCAACT	GGTAAAATAG	TACCATTTC	CGCTGTTAAA	GTGTCGATC	CTACAAACAGG	1140
AAAATTTTG	GGGCCAATG	AAACTGGAGA	ATTGTATTT	AAAGGCAGA	TGATAATGAA	1200
AGGTTATTAT	AATAATGAAAG	AAGCTACTAA	AGCAATTATT	ACAAAGACG	GATGGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATT	TATATTGTGG	ACAGGCTGAA	1320
GTCATTAATT	AAATATAAAG	GTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGGGGCGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAAATAT	CTAACCGAAC	AAATCGTACA	1500
AAATTTGTT	TCCAGTCAG	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATTTTT	1560
GGATGAAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAAACAC	<u>ACCAATGGG</u>					1639

FIG. 25

luc49-4G11 (SEQ ID NO:5)

GGATCCAATG	GAAGATAAAA	ATATTTATA	TGGACCTGAA	CCATTTTATC	CCTTGGCTGA	60
TGGGACGGCT	GGAGAACAGA	TGTTT G ACGC	ATTATCTCGT	TATGCAGATA	TTTCAGGATG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGC	CCCTGTTTA	TATGAAGAGT	T G TTAAAATT	180
GTCGTGTCGT	TTAGCGAAA	GT T TTAAAAAA	GTATGGATT	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	GAAAATGGTT	TGCAATT	CCTTCCTATA	ATTGCATCAT	TGTATCTTGG	300
AATAATTGCA	GCACCTGTTA	GTGATAAATA	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGTAAAA	CCACCGATAA	TTTTTGCTC	CAAGAATACT	TTTCAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATATG	TAGAAACTAT	TATTATATTA	GACTTAAATG	AAGACTT G GG	480
AGGTTATCAA	TGCCTCAACA	ACTTTATTTC	TCAAAATTCC	GATATTAATC	TTGACGTAAA	540
AAAATTAAA	CCATATTCTT	TTAATCGAGA	CGATCAGGTT	CGGTTGGTAA	TGTTTCTTC	600
TGGTACAAC	GGTGTTCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGACGATT	660
TTCTCAT <u>GCA</u>	AAAGATCCTA	CTTTGGTAA	CGCAATTAAAT	CCAACGACAG	CAATTAAAC	720
GGTAATACCT	TTCCACCATG	GT T TTGGTAT	GATGACCACA	TTAGGATACT	TTACTTGTGG	780
ATTCCGAGTT	GTTCTAATGC	ACACGTTGA	AGAAAAAACTA	TTTCTACAAAT	CATTACAAGA	840
TTATAAAGTG	GAAAGTACTT	TACTTGTACC	AACATTAATG	GCATTTTTG	CAAAAGTGC	900
ATTAGTTGAA	AAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	GCGCACCTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTGA AAA	ACGGTTTAAA	TTAAACTTTG	TCAGGCAAGG	1020
GTATGGATTA	ACAGAAACCA	CTTCGGCTGT	TTTAATTACA	CCGAACAAATG	ACGTCAGACC	1080
GGGATCAACT	GGTAAAATAG	TACCAATTCA	CGCTGTTAAA	GTGTCGATC	CTACAAACAGG	1140
AAAAATT TT TG	GGGCCAAATG	AAACTGGAGA	ATTGTATTTT	AAAGGCGACA	TGATAATGAA	1200
AGGTTATTAT	AATAATGAAG	AAGCTACTAA	AGCAATTAAIT	AACAAAGACG	GATGGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATT	TATATTGTGG	ACAGGCTGAA	1320
GTCATTAATT	AAATATAAAG	GT T ATCAGGT	TGCACCTGCT	GAAT T GAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGGGGCGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAAATAT	CTAAACGAAC	AAATCGTACA	1500
AAATT TT TGTT	TCCAGTCAAG	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATT	1560
GGATGAAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAACAC	<u>ACCAATGGG</u>					1639

FIG. 26

luc49-7C6 (SEQ ID NO:14)

G	T	A	M	A	D	K	N	I	L	Y	G	P	E	P	F	Y	P	L	A	D	20
I	A	L	T	T	N	A	H	T	K	E	N	V	R	Y	A	D	I	S	G	C	40
S	C	R	L	A	E	S	F	K	K	Y	G	L	Y	E	E	F	L	K	L	60	
V	C	S	E	N	G	L	Q	F	F	L	P	I	K	Q	N	D	T	I	A	80	
I	I	A	A	P	V	S	D	K	Y	I	E	R	E	L	A	S	L	Y	L	100	
I	V	K	P	R	I	I	F	C	S	K	N	T	F	Q	K	I	H	S	L	G	120
K	S	K	L	K	Y	V	E	T	I	I	L	D	L	I	N	E	D	D	L	G	140
G	Y	Q	C	L	N	N	F	T	I	S	Q	N	S	D	I	N	L	D	V	K	160
K	F	K	P	Y	S	F	N	R	D	D	Q	V	A	L	N	V	M	F	S	S	180
G	T	T	G	V	S	K	G	V	M	L	T	H	K	N	I	V	A	R	F	200	
S	L	A	K	D	P	T	F	G	N	A	I	N	P	T	T	A	I	L	T	220	
V	I	P	F	H	H	G	F	G	M	M	T	L	F	G	L	Y	F	T	C	G	240
F	R	V	V	L	M	H	T	F	E	E	K	L	F	Q	S	A	L	Q	D	260	
Y	K	V	E	S	T	L	L	V	P	T	L	M	A	F	L	G	A	K	S	280	
L	V	E	K	Y	D	L	L	S	H	L	K	E	I	A	S	N	F	G	P	300	
S	K	E	I	G	E	M	V	K	K	R	F	K	L	N	N	D	V	R	Q	320	
Y	G	L	T	E	T	T	S	A	V	L	I	T	P	V	N	N	D	P	P	340	
G	S	T	G	K	I	V	P	F	H	A	V	K	V	V	V	D	D	P	T	T	360
K	I	L	G	P	N	E	E	A	T	K	A	F	K	G	D	M	I	M	K	380	
G	Y	Y	N	N	E	E	A	T	K	A	I	I	N	K	D	G	W	L	R	400	
S	G	D	I	A	Y	Y	D	N	D	G	H	F	Y	I	V	D	R	L	K	420	
S	L	I	K	Y	K	G	Y	Q	V	A	P	A	E	E	G	I	I	L	L	440	
Q	H	P	Y	I	V	D	A	G	V	T	G	K	Y	I	P	D	E	A	A	460	
L	P	A	A	G	V	V	V	Q	T	G	K	Y	L	N	E	Q	I	V	Q	480	
N	F	V	S	S	Q	V	S	T	A	K	W	R	K	V	G	V	K	F	L	500	
D	E	I	P	K	G	S	T	G	K	I	D	R	K	V	L	R	Q	M	F	520	
E	K	H	T	N	G																540
																					546

FIG. 27

Luc49-6C10 (SEQ ID NO:15)

M	E	D	K	N	I	L	Y	G	P	E	P	F	Y	P	L	A	D	20		
G	T	A	G	E	Q	M	F	Y	A	L	S	R	Y	A	D	I	S	G	C	40
I	A	L	T	N	A	H	T	K	E	N	V	L	Y	E	E	L	L	K	L	60
S	C	R	L	A	E	S	F	K	K	Y	G	L	K	Q	N	D	T	I	A	80
V	C	S	E	N	G	L	Q	F	F	L	P	I	I	A	S	L	Y	L	G	100
I	I	A	A	P	V	S	D	K	Y	I	E	R	E	L	I	H	S	L	G	120
I	V	K	P	R	I	I	F	C	S	K	N	T	F	Q	K	V	L	N	V	140
K	S	K	L	K	Y	V	E	T	I	I	I	L	D	L	N	E	D	L	G	160
G	Y	Q	C	L	N	N	F	I	S	Q	N	S	D	I	N	L	D	V	K	180
K	F	K	P	Y	S	F	N	R	D	D	Q	V	A	L	V	M	F	S	S	200
G	T	T	G	V	S	K	G	V	M	L	T	H	K	N	I	V	A	R	F	220
S	H	A	K	D	P	T	F	G	N	A	I	N	P	T	T	A	I	L	T	240
V	I	P	F	H	H	G	F	G	M	M	T	T	L	G	Y	F	T	C	G	260
F	R	V	V	L	M	H	T	F	E	E	K	L	F	L	Q	S	L	Q	D	280
Y	K	V	E	S	T	L	L	V	P	T	L	M	A	F	F	A	K	S	A	300
L	V	E	K	Y	D	L	S	H	L	K	E	I	A	S	G	G	A	P	L	320
S	K	E	I	G	E	M	V	K	K	R	F	K	L	N	F	V	R	Q	G	340
Y	G	L	T	E	T	T	S	A	V	L	I	T	P	N	N	D	V	R	P	360
G	S	T	G	K	I	V	P	F	H	A	V	K	V	V	D	P	T	T	G	380
K	I	L	G	P	N	E	T	G	E	L	Y	F	K	G	D	M	I	M	K	400
G	Y	Y	N	N	E	E	A	T	K	A	I	I	N	K	D	G	W	L	R	420
S	G	D	I	A	Y	Y	D	N	D	G	H	F	Y	I	V	D	R	L	K	440
S	L	I	K	Y	K	Y	Q	V	A	P	A	E	I	E	G	I	L	L	460	
Q	H	P	Y	I	V	D	A	G	V	T	G	I	P	D	E	A	A	G	E	480
L	P	A	A	G	V	V	V	Q	T	G	K	Y	L	N	E	Q	I	V	Q	500
N	F	V	S	S	Q	V	S	T	A	K	W	L	R	G	G	V	K	F	L	520
D	E	I	P	K	G	S	T	G	K	I	D	R	K	V	L	R	Q	M	F	540
E	K	H	T	N	G															546

FIG. 28

Luc49-0G12 (SEQ ID NO:16)

M E D K N I L Y G P E P F Y P L A D 20
G T A G E Q M F Y A L S R Y A D I S G C 40
I A L T N A H T K E N V L Y E E F L K L 60
S C R L A E S F K K Y G L K Q N D T I A 80
V C S E N G L Q F F L P I I A S L Y L G 100
I I A A P V S D K Y I E R E L I H S L G 120
I V K P R I I F C S K N T F Q K V L N V 140
K S K L K Y V E T I I I L D L N E D L G 160
G Y Q C L N N F I S Q N S D I N L D V K 180
K F K P Y S F N R D D Q V A L V M F S S 200
G T T G V S K G V M L T H K N I V V R F 220
S L A K D P T F G N A I N P T T A I L T 240
V I P F H H G F G M M T T L G Y F T C G 260
F R V V L M H T F E E K L F L Q S L Q D 280
Y K V E S T L L V P T L M A F F A K S A 300
L V E K Y D L S H L K E I A S G G A P L 320
S K E I G E M V K K R F K L N F V R Q G 340
Y G L T E T T S A V L I T P N N D V R P 360
G S T G K I V P F H A V K V V D P T T G 380
K I L G P N E T G E L Y F K G D M I M K 400
G Y Y N N E E A T K A I I T K D G W L R 420
S G D I A Y Y D N D G H F Y I V D R L K 440
S L I K Y K G Y Q V A P A E I E G I L L 460
Q H P Y I V D A G V T G I P D E A A G E 480
L P A A G V V V Q T G K Y L N E Q I V Q 500
N F V S S Q V S T A K W L R G G V K F L 520
D E I P K G S T G K I D R K V L R Q M F 540
E K H T N G 546

FIG. 29

Luc49-7A5 (SEQ ID NO:17)

M	E	D	K	N	I	L	Y	G	P	E	P	F	Y	P	L	A	D	20			
G	T	A	G	E	Q	M	F	Y	A	L	S	R	Y	A	D	I	S	G	C	40	
I	A	L	T	N	A	H	T	K	E	N	V	L	Y	E	E	F	L	K	L	60	
S	C	R	L	A	E	S	F	K	K	Y	G	L	K	Q	N	D	T	I	A	80	
V	C	S	E	N	G	L	Q	F	F	L	P	I	I	A	S	L	Y	L	G	100	
I	I	A	A	P	V	S	D	K	Y	I	E	R	E	L	I	H	S	L	G	120	
I	V	K	P	R	I	I	F	C	S	K	N	T	F	Q	K	V	L	N	V	140	
K	S	K	L	K	Y	V	E	T	I	I	I	L	D	L	N	E	D	L	G	160	
G	Y	Q	C	L	N	N	F	I	S	Q	N	S	D	I	N	L	D	V	K	180	
K	F	K	P	<u>Y</u>	S	F	N	R	D	D	Q	V	A	L	V	M	F	S	S	200	
G	T	T	G	V	S	K	G	V	M	L	T	H	K	N	I	V	A	R	F	220	
S	<u>I</u>	A	K	D	P	T	F	G	N	A	I	N	P	T	T	A	I	L	T	240	
V	I	P	F	H	H	G	F	G	M	M	T	T	L	G	Y	F	T	C	G	260	
F	R	V	<u>V</u>	L	M	H	T	F	E	E	K	L	F	L	Q	S	L	Q	D	280	
Y	K	V	E	S	T	L	L	V	P	T	L	M	A	F	<u>L</u>	A	K	S	A	300	
L	V	E	K	Y	D	L	S	H	L	K	E	I	A	S	G	G	A	P	L	320	
S	K	E	I	G	E	M	V	K	K	R	F	K	L	N	F	V	R	Q	G	340	
Y	G	L	T	E	T	T	S	A	V	L	I	T	P	<u>N</u>	N	D	V	R	P	360	
G	S	T	G	K	I	V	P	F	H	A	V	K	V	V	D	P	T	T	G	380	
K	I	L	G	P	N	E	T	G	E	L	Y	F	K	G	D	M	I	M	K	400	
G	Y	Y	N	N	E	E	A	T	K	A	I	I	N	K	D	G	W	L	R	420	
S	G	D	I	A	Y	Y	D	N	D	G	H	F	Y	I	V	D	R	L	K	440	
S	L	I	K	Y	K	G	Y	Q	V	A	P	A	E	I	E	G	I	L	L	460	
Q	H	P	Y	I	V	D	A	G	V	T	G	I	P	D	E	A	A	G	E	480	
L	P	A	A	G	V	V	V	Q	T	G	K	Y	L	N	E	Q	I	V	Q	500	
N	F	V	S	S	Q	V	V	S	T	A	K	W	L	R	G	G	V	K	F	L	520
D	E	I	P	K	G	S	T	G	K	I	D	R	K	V	L	R	Q	M	F	540	
E	K	H	<u>T</u>	<u>N</u>	<u>G</u>															546	

FIG. 30

Luc49-4G11 (SEQ ID NO:18)

M E D K N I L Y G P E P F Y P L A D 20
G T A G E Q M F D A L S R Y A D I S G C 40
I A L T N A H T K E N V L Y E E F L K L 60
S C R L A E S F K K Y G L K Q N D T I A 80
V C S E N G L Q F F L P I I A S L Y L G 100
I I A A P V S D K Y I E R E L I H S L G 120
I V K P R I I F C S K N T F Q K V L N V 140
K S K L K Y V E T I I I I L D L N E D L G 160
G Y Q C L N N F I S Q N S D I N L D V K 180
K F K P Y S F N R D D Q V A L V M F S S 200
G T T G V S K G V M L T H K N I V A R F 220
S H A K D P T F G N A I N P T T A I L T 240
V I P F H H G F G M M T T L G Y F T C G 260
F R V V L M H T F E E K L F L Q S L Q D 280
Y K V E S T L L V P T L M A F F A K S A 300
L V E K Y D L S H L K E I A S G G A P L 320
S K E I G E M V K K R F K L N F V R Q G 340
Y G L T E T T S A V L I T P N N D V R P 360
G S T G K I V P F H A V K V V D P T T G 380
K I L G P N E T G E L Y F K G D M I M K 400
G Y Y N N E E A T K A I I N K D G W L R 420
S G D I A Y Y D N D G H F Y I V D R L K 440
S L I K Y K G Y Q V A P A E I E G I L L 460
Q H P Y I V D A G V T G I P D E A A G E 480
L P A A G V V V Q T G K Y L N E Q I V Q 500
N F V S S Q V S T A K W L R G G V K F L 520
D E I P K G S T G K I D R K V L R Q M F 540
E K H T N G 546

FIG. 31

luc78-0B10 (SEQ ID NO:6)

GGATCCAATG	<u>G</u> CAGATAAGA	ATATTTTATA	TGGGCC <u>C</u> GA	CCATTTTATC	CCTTGGCTGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTG <u>AC</u> GC	ATTATCTCGT	TATGCAGATA	TTTC <u>CG</u> GGATG	120
CATAGCATTG	ACAA <u>AT</u> GCTC	ATACAAAAGA	AAATGTTTTA	TATGAAGAGT	TTTTAAAATT	180
GTCGTGTCGT	TTAGCGAAA	GTTTTAAAAA	GTATGGATT	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	AAAATGGTT	TGCAATT <u>TTT</u> T	CCTTC <u>CT</u> GTA	ATTGCATCAT	TGTATCTTGG	300
AAA <u>AT</u> TTGCA	GCACCTGTTA	GTGATAAAATA	CATTGAACGT	GAATTAATAAC	ACAGTCTTGG	360
TATTGAAAAA	CCACGCATAA	TTTTTG <u>CT</u> C	CAAGAATACT	TTTCAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAAT <u>CT</u> G	TAGAAACTAT	TATTATATT	GA <u>CT</u> TAATG	AAGACTT ^{TT} AGG	480
AGGTTATCAA	TGCCCAAC	ACTTTAT <u>TT</u> C	TCAAAATTCC	GATAG <u>TA</u> ATC	TGGACGTAAA	540
AAA <u>AT</u> TTAAA	CC <u>AT</u> TTCTT	TTAATCGAGA	CGATCAGGTT	CGC <u>TT</u> GGTAA	TGTTTTCTTC	600
TGGTACA <u>AC</u> T	GGTGT <u>TC</u> GA	AGGGAGTCAT	GCTA <u>AC</u> TCAC	AAGAATATTG	TTGCACGATT	660
TTCT <u>CT</u> TC <u>GA</u>	AAAGATCCTA	CTTTGGTAA	CGCAATTAA	CC <u>AC</u> GACAG	CAATTTAAC	720
GGTA <u>AT</u> ACCT	TTCCACC <u>AT</u> G	GTTTTGGTAT	GATGACCACA	TTAGGATACT	TTACTTTGTGG	780
ATTCCGAGTT	<u>G</u> TTCTAATGC	ACACGTTGA	AGAAAAACTA	TTTCTACAA	CATTACAAGA	840
TTAT <u>AA</u> AGTG	GAAGTACTT	TACTTGACC	AA <u>CT</u> TAATG	GCATTT <u>CT</u> TG	AAAAAGTGC	900
ATTAGTTGAA	AAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	GCGCACCTT	960
AT <u>CA</u> AAAAGAA	ATTGGGGAGA	TGGTGA <u>AAA</u> A	ACGGTTTAAA	TTAA <u>AC</u> TTTG	TCAGGCAAGG	1020
GTATGGATTA	ACAGAACCA	CTTCGGCTGT	TTTAATTACA	CCG <u>AA</u> AGGTG	ACG <u>CC</u> AGACC	1080
GGGATCA <u>AC</u> T	GGTAAAATAG	TACCATTCA	CGCTGTTAAA	GTTGTCGATC	CTACAA <u>AC</u> AGG	1140
AAA <u>AT</u> TTTG	GGGCCAAATG	AA <u>CT</u> GGAGA	ATTGTATTTT	AAAGGCG <u>CC</u> A	TGATAATGAA	1200
<u>GG</u> GT <u>AT</u> TAT	AATAATGAAG	AAGCTACTAA	AGCAATTATT	GAT <u>AA</u> ATGACG	GATGGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATT <u>TT</u> T	TATATTGTGG	ACAGGCTGAA	1320
GTCATTAATT	AA <u>AT</u> ATAAAAG	GTTATCAGGT	TGCA <u>CC</u> TGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGC <u>GGG</u> CGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAAATAT	CTAAACGAAC	AAATCGTACA	1500
AGATTTGTT	TCCAGTCAAG	TTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATTTT	1560
GGATGAAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAAAACAC	<u>AC</u> CA <u>AT</u> GGG					1639

FIG. 32

luc78-0G8 (SEQ ID NO:7)

GGATCCAATG	GCAGATAAAA	ATATTTATA	TGGGCCGAA	CCATTTTATC	CCTTGGCTGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTTACGC	ATTATCTCGT	TATGCAGATA	TTTCAGGATG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGC	CCCTGTTTA	TATGAAGAGT	TTTTAAAATT	180
GTCGTGTCTG	TTAGCGGAAA	GTTTTAAA	GTATGGATTA	AAACAAAACG	ACACAATAGC	240
GTCGTGTAGC	GAAAATGGTT	TGCAATT	CCTTCCCTGTA	ATTGCATCAT	TGTATCTTGG	300
ATAATTGCA	GCACCTGTTA	GTGATAAATA	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGTA	CCACGCATAA	TTTTTGCTC	CAAGAATACT	TTTCAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATATG	TAGAAAAC	TATTATATTA	GACTTAAATG	AAGACTTAGG	480
AGGTTATCAA	TGCCTCAACA	ACTTTTATTC	TCAAAATTCC	GATATTAATC	TTGACGTAAA	540
AAAATTAAA	CCATATTCTT	TTAATCGAGA	CGATCAGGTT	GCGTTGGTAA	TGTTTCTTC	600
TGGTACAAC	GGTGGTTCGGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGCACGATT	660
TTCTCTTGC	AAAGATCCTA	CTTTGGTAA	CGCAATTAAAT	CCAACGACAG	CAATTAAAC	720
GTTAACTACT	TTCCACCAATG	GTTTTGGTAT	GATGACCAACA	TTAGGATACT	TTACTTGTGG	780
ATTCCGAGTT	GTTCTAATGC	ACACGTTGA	AGAAAAACTA	TTTCTACAAT	CATTACAAGA	840
TTATAAAGTG	GAAAGTACTT	TACTTGTACC	AACATTAAATG	GCATTTCTTG	CAAAAAGTGC	900
ATTAGTTGAA	AAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	GCGCACCTTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTGAAAAA	ACGGTTTAAA	TTAAACTTTG	TCAGGCAAGG	1020
GTATGGATTA	ACAGAAACCA	CTTCGGCTGT	TTTAATTACA	CCGAAAXXXX	XXGTCAGACC	1080
GGGATCAACT	GGTAAAATAG	TACCAATTCA	CGCTGTTAAA	GTTGTCGATC	CTACAACAGG	1140
AAAATTGTG	GGGCCAAATG	AACTGGAGA	ATTGTATTTT	AAAGGCGACA	TGATAATGAA	1200
AGGTTATTAT	ATAATGAAG	AAGCTACTAA	AGCAATTATT	GATAAAAGACG	GATGGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATT	TATATTGTGG	ACAGGCTGAA	1320
GTCATTAATT	AAATATAAAG	GTTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGGGGCGA	1440
GCTTCCAGCT	GCAGGTGTG	TAGTACAGAC	TGGAAAATAT	CTAAACGAAC	AAATCGTACA	1500
AAATTGTTG	TCCAGTCAAG	TTTCAACAGC	CAAATGGCTA	CGGGGTGGGG	TGAAATTGTT	1560
GGATGAAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAAACAC	<u>ACCAATGGG</u>					1639

FIG. 33

Luc78-1E1 (SEQ ID NO: 8)

GGATCCAATG	G <u>C</u> AGATAAAA	ATATTTATA	TGGGCCGAA	CCATTTATC	CCTGGCTGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTACGC	ATTATCTCGT	TATGAGATA	TTTCAGGATG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGC	CCCTGTTTA	TATGAAGAGT	T <u>G</u> TTAAAATT	180
GTCGTGTCGT	TTAGCGAAA	GTTTTAAAAA	GTATGGATTA	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	GAAAATGGTT	TGCAATATT	CCTTCCTGTA	ATTGACATCAT	TGTATCTTGG	300
ATAATTGCA	GCACCTGTTA	GTGATAAATA	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGTA	AAAATCTAAA	CCACGCATAA	TTTTTGCTC	CAAGAATACT	TTTCAAAAG	420
TTAAATATG	TTAAATATG	TAGAAACTAT	TATTATATA	GA <u>C</u> TTAAATG	AAGACTT ^{AGG}	480
AGGTTATCAA	TGCCTCAACA	ACTTTATTC	TCAAAATTCC	GATATTAATC	TTGACGTAAA	540
AAAATTAAA	CC <u>A</u> TATTCTT	TAAATCGAGA	CGATCAGGTT	GC <u>G</u> TTGGTAA	TGTTTCTTC	600
TGGTACAAC	GGTGT <u>CC</u> GA	AGGGAGTCAT	GCTAAC <u>TC</u> CAC	A <u>G</u> AATATTG	TTGACGATT	660
TTCTATTGCA	AAAGATCCTA	CTTTGGTAA	CGCAAA <u>TA</u> AT	CCAACGACAG	CAATTTAAC	720
GGTAATACCT	TTCCACCATG	GT <u>TTT</u> GGTAT	GATGACCACA	TTAGGATACT	TTACTTG ^{GG}	780
ATTCCGAGTT	G <u>T</u> CTAA <u>AT</u> GC	ACACGTTGA	AGAAAAACTA	TTTCTACAAAT	CA <u>T</u> ACAAGA	840
TTATAAAGTG	GAAAGTACTT	TACTTGACC	AACATTAATG	GCATTT <u>CT</u> TG	CAAAAAGTGC	900
ATTAGTTGAA	AAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	CGC ^{AC} CTTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTGA <u>AAA</u> AA	ACGGTTTAA	TTAA <u>ACT</u> TTG	TCAGGCAAGG	1020
GTATGGATTA	ACAGAAACCA	CTCGGCTGT	TTTAATTACA	CG <u>AA</u> xx	xxG <u>C</u> AGACC	1080
GGGATCAACT	GGTAAAATAG	TACCA <u>TT</u> CA	CGCTGTTAA	GTGTCGATC	CTACAA <u>AC</u> AGG	1140
AAAAATTTG	GGGCCAAATG	A <u>Q</u> CTGGAGA	ATTGTTTTT	AAAGGCG <u>CA</u>	TGATAATGAA	1200
GGTTTATTAT	AATAATGAAG	AGCTACTAA	AGCAAA <u>TT</u> TT	ACAA <u>AG</u> ACG	GATGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATT	TATATTG ^{GG}	ACAGGCTGAA	1320
GTCATTAATT	AAATATAAAG	GT <u>T</u> ATCAGGT	TGCACCTGCT	GAA <u>AT</u> TGAGG	GAAT <u>ACT</u> CTT	1380
ACAACATCCG	TATATTGTTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGGGGCGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAA <u>AT</u> AT	CTAA <u>AC</u> GAAC	AAATCGTACA	1500
AAATTTGTT	TCCAGTCAAG	TTCAACAGC	CAA <u>AT</u> GGCTA	CGTGGTGGGG	TGAA <u>AT</u> TTT	1560
GGATGAAATT	CCCAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAA <u>AT</u> GTT	1620
TGAAAAACAC	<u>ACCA</u> ATGGG					1639

FIG. 34

luc78-2B4 (SEQ ID NO: 9)

GGATCCAATG	G <u>CA</u> GATAAAAA	ATATTTATA	TGG <u>CCCG</u> AA	CCATTTATC	CCTTGGCTGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTGACGC	ATTATCTCGT	TATGCAGATA	TTTCAGGATG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGC	CCCTGTTTTA	TATGAAGAGT	TGTTAAAATT	180
GTCGTGTCGT	TTAGCGAAA	GTTTTAAAAA	GTATGGATTA	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	GAAAATGGTT	TGCAATTTTT	CCTTCCTGTA	ATTGCATCAT	TGTATCTTGG	300
ATAATTGCA	GCACCTGTTA	GTGATAAATA	C <u>GT</u> TGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGTAAAA	CCACGCAATAA	TTTTTGCTC	CAAGAATACT	TTTCAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATATG	TAGAAACTAT	TATTATATTA	GACTTAAATG	AAGACTTAGG	480
AGGTTATCAA	TGCCTCAACA	ACTTTATTC	TCAAAATTCC	GATAGTAATC	T <u>GA</u> CGTAAA	540
AAAATTAAA	CCAAATTCTT	TTAACGAGA	CGATCAGGTT	GC <u>GT</u> GGTAA	TGTTTTCTTC	600
TGGTACAAC	GGTGGTTCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGCACGATT	660
TTCTCTT <u>GC</u> A	AAAGATCCTA	CTTTGGTAA	CGCAATTAA	CCAACGACAG	CAATTAAAC	720
GTAATACCT	TTCCACCAATG	TTTTGGTAT	GATGACCACA	TTAGGATACT	TTACTTGTGG	780
ATTCCGAGTT	GTTCTAATGC	ACACGTTGA	AGAAAAACTA	TTTCTACAAT	CATTACAAGA	840
TTATAAAAGTG	GAAAGTACTT	TACTTGTACC	AACATTAATG	GCATTT <u>CT</u> TG	CAAAAAGTGC	900
ATTAGTTGAA	AAGTACGATT	TATCGCACTT	AAAAGAATT	GCATCTGGTG	GGCACCCTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTGA	ACGTTTAAA	TTAAACTTG	TCAGGCAAGG	1020
GTATGGATTA	ACAGAAACCA	CTTCGGCTGT	TTTAATTACA	CCG <u>AC</u> CCCC	xxGGAGACC	1080
GGGATCAACT	GGTAAAATAG	TACCATTCA	CGCTGTTAAA	GTTGT <u>CG</u> GATC	CTACAAACAGG	1140
AAAATTTTG	GGGCCAAAATG	AA <u>CT</u> GGAGA	ATTGTATTT	AAAGG <u>CC</u> CA	TGATAATGAA	1200
<u>GG</u> TTTATTAT	AATAATGAAG	AAGCTACTAA	AGCAATTATT	GA <u>TA</u> AAAGACG	GATGGTTGCG	1260
CTCTGGTGT	ATTGCTTATT	ATGACAATGA	TGGCCATT	TATATTGTGG	ACAGGCTGAA	1320
GTCATTAAAT	AAATATAAAG	GTTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTG	ATGCGGGCGT	TACTGGTATA	CCGGATGAAG	CCGGGGCGA	1440
GCTTCCAGCT	GCAGGTGTG	TAGTACAGAC	TGGAAAATAT	CTAAACGAAC	AAATCGTACA	1500
AAATTTGTT	TCCAGTCAAG	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATTTT	1560
GGATGAAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAACAC	<u>ACCA</u> ATGGG					1639

FIG. 35

luc78-0B10 (SEQ ID NO:19)

G	T	M	A	D	K	N	I	L	Y	G	P	E	P	F	Y	P	L	A	D	20	
I	A	A	G	E	Q	M	F	D	A	L	S	R	Y	A	D	I	S	G	C	40	
S	C	R	L	A	E	S	F	K	K	Y	G	L	Y	E	E	F	L	K	L	60	
V	C	S	E	N	G	L	Q	F	F	L	P	V	I	A	S	L	Y	L	G	80	
I	I	A	A	P	V	S	D	K	Y	I	E	R	E	L	I	H	S	L	G	100	
I	V	K	P	R	I	I	F	C	S	K	N	T	F	Q	K	V	L	N	V	120	
K	S	K	L	K	S	V	E	T	I	I	I	L	D	L	N	E	D	L	G	140	
G	Y	Q	C	L	N	N	F	I	S	Q	N	S	D	S	N	L	D	V	K	160	
K	F	K	P	Y	S	F	N	R	D	D	Q	V	A	L	V	M	F	S	S	180	
G	T	T	G	V	P	K	G	V	M	L	T	H	K	N	I	V	A	R	F	200	
S	L	A	K	D	P	P	T	F	G	N	A	I	N	P	T	T	A	I	L	220	
V	I	P	F	H	H	G	F	G	M	M	T	T	L	G	Y	F	T	C	G	240	
F	R	V	Y	L	M	H	T	F	E	E	K	L	F	Q	S	L	Q	D	260		
Y	K	V	E	S	T	L	L	V	P	T	L	M	A	F	L	A	K	S	A	280	
L	V	E	K	Y	D	L	S	H	L	K	E	I	A	S	G	G	A	P	L	300	
S	K	E	I	G	E	M	V	K	K	R	F	K	L	N	F	V	R	Q	G	320	
Y	G	L	T	E	E	T	T	S	A	V	L	I	T	P	K	G	D	A	R	340	
G	S	T	G	K	I	V	P	F	H	A	V	K	V	V	D	P	T	T	G	360	
K	I	L	G	P	N	E	E	P	G	E	L	Y	F	K	G	A	M	I	M	380	
G	Y	Y	N	N	A	Y	Y	D	N	D	G	H	F	Y	I	V	D	R	L	K	400
S	G	D	I	K	Y	K	G	Y	Q	V	A	P	A	E	I	E	G	I	L	L	420
S	L	I	P	Y	I	V	D	A	G	V	T	G	I	P	D	E	A	A	G	E	440
Q	H	P	Y	I	V	V	V	Q	T	G	K	Y	L	N	E	Q	I	V	Q	460	
L	P	A	A	G	V	V	Q	T	G	K	Y	L	R	G	G	V	K	F	L	480	
D	F	V	S	S	Q	V	S	T	A	K	W	L	R	G	G	V	K	F	L	500	
D	E	I	P	K	G	S	T	G	K	I	D	R	K	V	L	R	Q	M	F	520	
E	K	H	T	N	G															540	
																				546	

FIG. 36

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Luc78-0G8 (SEQ ID NO:20)

M A D K N I L Y G P E P F Y P L A D 20
G T A G E Q M F Y A L S R Y A D I S G C 40
I A L T N A H T K E N V L Y E E F L K L 60
S C R L A E S F K K Y G L K Q N D T I A 80
V C S E N G L Q F F L P V I A S L Y L G 100
I I A A P V S D K Y I E R E L I H S L G 120
I V K P R I I F C S K N T F Q K V L N V 140
K S K L K Y V E T I I I I L D L N E D L G 160
G Y Q C L N N F I S Q N S D I N L D V K 180
K F K P Y S F N R D D Q V A L V M F S S 200
G T T G V P K G V M L T H K N I V A R F 220
S L A K D P T F G N A I N P T T A I L T 240
V I P F H H G F G M M T T L G Y F T C G 260
F R V V L M H T F E E K L F L Q S L Q D 280
Y K V E S T L L V P T L M A F L A K S A 300
L V E K Y D L S H L K E I A S G G A P L 320
S K E I G E M V K K R F K L N F V R Q G 340
Y G L T E T T S A V L I T P K X X V R P 360
G S T G K I V P F H A V K V V D P T T G 380
K I L G P N E P G E L Y F K G D M I M K 400
G Y Y N N E E A T K A I I D K D G W L R 420
S G D I A Y Y D N D G H F Y I V D R L K 440
S L I K Y K G Y Q V A P A E I E G I L L 460
Q H P Y I V D A G V T G I P D E A A G E 480
L P A A G V V V Q T G K Y L N E Q I V Q 500
N F V S S Q V S T A K W L R G G V K F L 520
D E I P K G S T G K I D R K V L R Q M F 540
E K H T N G 546

FIG. 37

Luc78-1E1 (SEQ ID NO:21)

M A D K N I L Y G P E P F Y P L A D 20
G T A G E Q M F D A L S R Y A D I P G C 40
I A L T N A H T K E N V L Y E E F L K L 60
S C R L A E S F K K Y G L K Q N D T I A 80
V C S E N G L Q Y F L P V I A S L Y L G 100
I I A A P V S D K Y I E R E L I H S L G 120
I V K P R I I F C S K N T F Q K V L N V 140
K S K L K Y V E T I I I L D L N E D L G 160
G Y Q C L N N F I S Q N S D I N L D V K 180
K F K P N S F N R D D Q V A L V M F S S 200
G T T G V P K G V M L T H K N I V A R F 220
S I A K D P T F G N A I N P T T A I L T 240
V I P F H H G F G M M T T L G Y F T C G 260
F R V V L M H T F E E K L F L Q S L Q D 280
Y K V E S T L L V P T L M A F L A K S A 300
L V E K Y D L S H L K E I A S G G A P L 320
S K E I G E M V K K R F K L N F V R Q G 340
Y G L T E T T S A V L I T P K X X A R P 360
G S T G K I V P F H A V K V V D P T T G 380
K I L G P N E P G E L Y F K G A M I M K 400
G Y Y N N E E A T K A I I D K D G W L R 420
S G D I A Y Y D N D G H F Y I V D R L K 440
S L I K Y K G Y Q V A P A E I E G I L L 460
Q H P Y I V D A G V T G I P D E A A G E 480
L P A A G V V V Q T G K Y L N E Q I V Q 500
N F V S S Q V S T A K W L R G G V K F L 520
D E I P K G S T G K I D R K V L R Q M F 540
E K H T N G 546

FIG. 38

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Luc78-2B4 (SEQ ID NO:22)

M	A	D	K	N	I	L	Y	G	P	E	P	F	Y	P	L	A	D	20			
G	T	A	G	E	Q	M	F	<u>D</u>	A	L	S	R	Y	A	D	I	<u>P</u>	G	C	40	
I	A	L	T	N	A	H	T	K	E	N	V	L	Y	E	E	F	L	K	L	60	
S	C	R	L	A	E	S	F	K	K	Y	G	L	K	Q	N	D	T	I	A	80	
V	C	S	E	N	G	L	Q	F	F	L	P	<u>V</u>	I	A	S	L	Y	L	G	100	
I	I	A	A	P	V	S	D	K	Y	<u>V</u>	E	R	E	L	I	H	S	L	G	120	
I	V	K	P	R	I	I	F	C	S	K	N	T	F	Q	K	V	L	N	V	140	
K	S	K	L	K	Y	V	E	T	I	I	I	L	D	L	N	E	D	L	G	160	
G	Y	Q	C	L	N	N	F	I	S	Q	N	S	D	<u>S</u>	N	L	D	V	K	180	
K	F	K	P	N	S	F	N	R	D	D	Q	V	A	L	V	M	F	S	S	200	
G	T	T	G	V	<u>P</u>	K	G	V	M	L	T	H	K	N	I	V	A	R	F	220	
S	L	A	K	D	P	T	F	G	N	A	I	N	P	T	T	A	I	L	T	240	
V	I	P	F	H	H	G	F	G	M	M	T	T	L	G	Y	F	T	C	G	260	
F	R	V	<u>V</u>	L	M	H	T	F	E	E	K	L	F	L	Q	S	L	Q	D	280	
Y	K	V	E	S	T	L	L	V	P	T	L	M	A	F	<u>L</u>	A	K	S	A	300	
L	V	E	K	Y	D	L	S	H	L	K	E	I	A	S	G	G	A	P	L	320	
S	K	E	I	G	E	M	V	K	K	R	F	K	L	N	F	V	R	Q	G	340	
Y	G	L	T	E	T	T	S	A	V	L	I	T	P	K	X	X	<u>A</u>	R	P	360	
G	S	T	G	K	I	V	P	F	H	A	V	K	V	V	D	P	T	T	G	380	
K	I	L	G	P	N	E	T	G	E	L	Y	F	K	G	<u>A</u>	M	I	M	K	400	
G	Y	Y	N	N	E	E	A	T	K	A	I	I	<u>D</u>	K	D	G	W	L	R	420	
S	G	D	I	A	Y	Y	D	N	D	G	H	F	Y	I	V	D	R	L	K	440	
S	L	I	K	Y	K	Y	Q	V	A	P	A	E	I	E	G	I	L	L	460		
Q	H	P	Y	I	V	D	A	G	V	T	G	I	P	D	E	A	A	G	E	480	
L	P	A	A	G	V	V	V	Q	T	G	K	Y	L	N	E	Q	I	V	Q	500	
N	F	V	S	S	Q	V	V	S	T	A	K	W	L	R	G	G	V	K	F	L	520
D	E	I	P	K	G	S	T	G	K	I	D	R	K	V	L	R	Q	M	F	540	
E	K	H	T	N	G															546	

FIG. 39

1uc85-4F12 (SEQ ID NO:10)

GGATCCAATG	GCAGATAAAA	ATATTTATA	TGGGCCGGAA	CCATTTATC	CCTTGGCTGA	60
TGGGACGGCT	GGAGAACAGA	TGTGTTACGC	ATTATCTCGT	TATGCAGATA	TTCCGGCTG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGC	CCCTGTTTTA	TATGAAGAGT	TTTTAAAATT	180
GTCGTGTCGT	TTAGCGGAAA	GTTTTAAAAA	GTATGGATTA	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	AAAAATGGTT	TGCAATTTC	CCTTCCTGTA	ATTGCATCAT	TGTATCTTGG	300
AATAATTGTC	GCACCTGTTA	ACGATAAAATA	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGTAAAAA	CCACGCATAG	TTTTTTGCTC	CAAGAACTACT	TTTCAAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATCTG	TAGAAACTAT	TATTATATTA	GACTTAAATG	AAGACTTAGG	480
AGGTTATCAA	TGCCTCAACA	ACTTTATTTC	TCAAAATTCC	GATATTAATC	TTGACGTAAA	540
AAAATTTAAA	CCATATTCTT	TTAATCGAGA	CGATCAGGTT	GCGTTGATTA	TGTTTCTTC	600
TGGTACAAC	GGTCTGCCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGACCGATT	660
TTCTCTTCCA	AAAGATCCTA	CTTTTGGTAA	CGCAATTAAAT	CCGACGACAG	CAATTAAAC	720
GGTAATACCT	TTCCACCATG	TTTTGGTAT	GATGACCACA	TTAGGATACT	TTACTTGTGG	780
ATTCGGAGTT	GTTCTAATGC	ACACGTTTGA	AGAAAAACTA	TTTCTACAAT	CATTACAAGA	840
TTATAAAGTG	GAAAGTACTT	TACTTGTACC	AACATTAATG	GCATTTCTTG	CAAAAAGTGC	900
ATTAGTTGAA	AACTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	GCGCACCTTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTAAAAAA	ACGGTTAAA	TTAAACTTTG	TCAGGCAAGG	1020
GTATGGATTA	ACAGAAACCA	CTTGGCTGT	TTTAATTACA	CCGAAACGCC	CGGCCAGACC	1080
GGGATCAACT	GGTAAAATAG	TACCATTTCA	CGCTGTTAAA	GTTGTCGATC	CTACAAACAGG	1140
AAAATTTG	GGGCCAAATG	AACTGGAGA	ATTGTATTTT	AAAGGCCGA	TGATAATGAA	1200
GGGTTATTAT	AATAATGAAG	AAGCTACTAA	AGCAATTATT	GATAATGACG	GATGGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCATTTC	TATATTGTGG	ACAGGCTGAA	1320
GTCATTAATT	AAATATAAAG	GTTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTTG	ATGCCGGCT	TACTGGTATT	CCGGATGAAG	CCGGGGCGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAAATAT	CTAAACGAAC	AAATCGTACA	1500
AAATTTGTT	TCCAGTCAAG	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATTTTT	1560
GGATGAAATT	CCCAAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAAACAC	<u>ACCAATGGG</u>					1639

FIG. 40

Luc85-4F12 (SEQ ID NO:23)

M A D K N I L Y G P E P F Y P L A D 20
G T A G E Q M F D A L S R Y A D I P G C 40
I A L T N A H T K E N V L Y E E F L K L 60
S C R L A E S F K K Y G L K Q N D T I A 80
V C S E N G L Q F F L P V I A S L Y L G 100
I I Y A P V N D K Y I E R E L I H S L G 120
I V K P R I I F C S K N T F Q K V L N V 140
K S K L K S V E T I I I L D L N E D L G 160
G Y Q C L N N F I S Q N S D I N L D V K 180
K F K P Y S F N R D D Q V A L I M F S S 200
G T T G L P K G V M L T H K N I V A R F 220
S L A K D P T F G N A I N P T T A I L T 240
V I P F H H G F G M M T T L G Y F T C G 260
F R V V L M H T F E E K L F L Q S L Q D 280
Y K V E S T L L V P T L M A F L A K S A 300
L V E K Y D L S H L K E I A S G G A P L 320
S K E I G E M V K K R F K L N F V R Q G 340
Y G L T E T T S A V L I T P K X X A R P 360
G S T G K I V P F H A V K V V D P T T G 380
K I L G P N E P G E L Y F K G P M I M K 400
G Y Y N N E E A T K A I I D N D G W L R 420
S G D I A Y Y D N D G H F Y I V D R L K 440
S L I K Y K G Y Q V A P A E I E G I L L 460
Q H P Y I V D A G V T G I P D E A A G E 480
L P A A G V V V Q T G K Y L N E Q I V Q 500
D F V S S Q V S T A K W L R G G V K F L 520
D E I P K G S T G K I D R K V L R Q M F 540
E K H T N G 546

FIG. 41

Luc90-1B5 (SEQ ID NO:11)

GGATCCAATG	GCAGATAAGA	ATATTTATA	TGGGCCCGAA	CCATTTATC	CCTTGGAAAGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTGACGC	ATTATCTCGT	TATGCAGATA	TTCCGGGCTG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGA	AAATGTTTTA	TATGAAGAGT	TTCTGAAACT	180
GTCGTGTCGT	TTAGCGGAAA	GTTTTAAAAA	GTATGGATTA	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	GAAAATGGTC	TGCAATTTC	CCTTCCTGTA	ATTGCATCAT	TGTATCTTGG	300
AATAATTGTC	GCACCTGTTA	ACGATAAATA	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATGTAAAGA	CCACGCATAG	TTTTTGCTC	CAAGAATACT	TTTCAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATCTA	TGAAAACATAT	TATTATATTA	GACTTAAATG	AAGACTTACG	480
AGGTTATCAA	TGCCTCAACA	ACTTTATTC	TCAAAATTCC	GATACTAATC	TGGACGTAAA	540
AAAATTAAAC	CCATATTCTT	TTAATCGAGA	CGATCAGGTT	GCGTGTGATTA	TGTTTTCTTC	600
TGGTACAACG	GGTCTGCCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGCACGATT	660
TTCTCTTGCA	AAAGATCCTA	CTTTGGTAA	CGCAATTAAT	CCGACGACAG	CAATTAAAC	720
GGTAATACCT	TTCCACCATG	TTTTGGTAT	GATGACCAACA	TTAGGAACT	TTACTGTGG	780
ATTCCGAGTT	GTTCTAATGC	ACACGTTTGA	AGAAAAACTA	TTTCTACAAAT	CATTACAAGA	840
TTATAAAGTG	GAAAGTACTT	TACTTGTACC	AAACATTAATG	GCATTTCTG	CAAAAAGTGC	900
ATTAGTTGAA	AAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	GCGCACCTTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTAAAAAA	ACGGTTAAA	TTAAACTTG	TCAGGCAAGG	1020
GTATGGATTA	ACAGAAACCA	CTTCGGCTGT	TTTAATTACA	CCGAAAGGTG	ACGCCAAACC	1080
GGGATCAACT	GGTAAAATAG	TACCATTCA	CGCTGTTAAA	GTTGTCGATC	CTACAAACAGG	1140
AAAATTTTG	GGGCCAAATG	AACTCTGGAGA	ATTGTATTT	AAAGGCCCGA	TGATAATGAA	1200
GGGTTATTAT	AATAATGAAG	AAGCTACTAA	AGCAATTATT	GATAATGACG	GATGGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATTTC	TATATTGTGG	ACAGGCTGAA	1320
GTCACTGATT	AAATATAAAG	GTTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGCGGGCGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAAATAT	CTAAACGAAC	AAATCGTACA	1500
AGATTATGTT	GCCAGTCAG	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATTTTT	1560
GGATGAAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAAACAC	ACCAATGGG					1639

FIG. 42

Luc90-1B5 (SEQ ID NO:24)

M	A	D	K	N	I	L	Y	G	P	E	P	F	Y	P	L	E	D	20	
G	T	A	G	E	Q	M	F	D	A	L	S	R	Y	A	D	I	P	E	40
I	A	L	T	N	A	H	T	K	E	N	V	L	Y	E	E	F	L	K	60
S	C	R	L	A	E	S	F	K	K	Y	G	L	K	Q	N	D	T	I	80
V	C	S	E	N	G	L	Q	F	F	L	P	V	I	A	S	L	Y	L	100
I	I	Y	A	P	V	N	D	K	Y	I	E	R	E	L	I	H	S	L	120
I	V	K	P	R	I	V	F	C	S	K	N	T	F	Q	K	V	L	N	140
K	S	K	L	K	S	I	E	T	I	I	I	L	D	L	N	E	D	L	160
G	Y	Q	C	L	N	N	F	I	S	Q	N	S	D	S	N	L	D	V	180
K	F	K	P	Y	S	F	N	R	D	D	Q	V	A	L	I	M	F	S	200
G	T	T	G	L	P	K	G	V	M	L	T	H	K	N	I	V	A	R	220
S	L	A	K	D	P	T	F	G	N	A	I	N	P	T	T	A	I	L	240
V	I	P	F	H	H	G	F	G	M	M	T	T	L	G	Y	F	T	C	260
F	R	V	V	L	M	H	T	F	E	E	K	L	F	G	Q	S	L	Q	280
Y	K	V	E	S	T	L	L	V	P	T	L	M	A	F	L	A	K	S	300
L	V	E	K	Y	D	L	S	H	L	K	E	I	A	S	G	G	A	P	320
S	K	E	I	G	E	M	V	K	K	R	F	K	L	N	F	V	R	Q	340
Y	G	L	T	E	T	T	S	A	V	L	I	T	P	K	G	D	A	K	360
G	S	T	G	K	I	V	P	F	H	A	V	K	V	V	D	P	T	T	380
K	I	L	G	P	N	E	E	A	T	K	A	I	I	D	N	D	G	I	400
G	Y	Y	N	N	E	E	A	T	K	A	I	I	D	N	D	G	W	L	420
S	G	D	I	A	Y	Y	D	N	D	G	H	F	Y	I	I	V	D	R	440
S	L	I	K	Y	K	G	Y	Q	V	A	P	A	E	I	E	G	I	L	460
Q	H	P	Y	I	V	D	A	G	V	T	G	I	I	P	D	E	A	A	480
L	P	A	A	G	V	V	V	Q	T	G	K	Y	L	N	E	Q	I	V	500
D	Y	V	A	S	Q	V	S	T	A	K	W	L	R	G	G	V	K	F	520
D	E	I	P	K	G	S	T	G	K	I	D	R	K	V	L	R	Q	M	540
E	K	H	T	N	G														546

FIG. 43

lucPpe2 [T249M] (SEQ ID NO:12)

GGATCCAATG	GAAGATAAAA	ATATTTATA	TGGACCTGAA	CCATTTATC	CCTGGCTGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTACGC	ATTATCTCGT	TATGCAGATA	TTTCAGGATG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGA	AAATGTTTA	TATGAAGAGT	TTTTAAAATT	180
GTCGTGTCGT	TTAGCGGAAA	TTTTAAAAAA	GTATGGATTA	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	AAAAATGGTT	TGCAATTTC	CCTTCCTTTA	ATTGCATCAT	TGTATCTTGG	300
AATAATTGCA	GCACCTGTTA	GTGATAAATA	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGAAAAA	CCACGCATAA	TTTTTGTTC	CAAGAATACT	TTTCAAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATATG	TAGAAACTAT	TATTATATTA	GACTTAAATG	AAGACTTAGG	480
AGGTTATCAA	TGCCTCAACA	ACTTTATTC	TCAAAATTCC	GATATTAATC	TTGACGTAAA	540
AAAATTTAAA	CCAAATTCTT	TTAATCGAGA	CGATCAGGTT	GCGTTGGTAA	TGTTTCTTC	600
TGGTACAAC	GGTGTTCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGCACGATT	660
TTCTCATTGC	AAAGATCCTA	CTTTGGTAA	CGCAATTAAT	CCAACGACAG	CAATTAAAC	720
GGTAATACCT	TTCCACCATG	TTTTGGTAT	GATGACCACA	TTAGGATACT	TTACTTGTGG	780
ATTCCGAGTT	GCTCTAATGC	ACACGTTGA	AGAAAAAACTA	TTTCTACAAT	CATTACAAGA	840
TTATAAAGTG	GAAAGTACTT	TACTTGTACC	AACATTAATG	GCATTTTTG	CAAAAGTGC	900
ATTAGTTGAA	AAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGTG	GCGCACCTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTGAAAAA	ACGGTTAAA	TTAAACTTTG	TCAGGCAAGG	1020
GTATGGATT	ACAGAAACCA	CTTCGGCTGT	TTTAATTACA	CCGGACACTG	ACGTCAGACC	1080
GGGATCAACT	GGTAAAATAG	TACCATTTCA	CGCTGTTAAA	GTTGTCGATC	CTACAAACAGG	1140
AAAATTTTG	GGGCCAAATG	AAACTGGAGA	ATTGTATTTT	AAAGGCGACA	TGATAATGAA	1200
AAGTTATTAT	AATAATGAAG	AAGCTACTAA	AGCAATTATT	AAACAAAGACG	GATGGTTGCG	1260
CTCTGGTGT	ATTGCTTATT	ATGACAATGA	TGGCATTTC	TATATTGTGG	ACAGGCTGAA	1320
GTCATTAATT	AAATATAAAG	GTTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGCAGGCGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAAATAT	CTAAACGAAC	AAATCGTACA	1500
AAATTTGTT	TCCAGTCAAG	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATTTT	1560
GGATGAAATT	CCCAAAGGAT	CAAATGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
TGAAAAACAC	AAATCTAAGC	TG				1642

FIG. 44

LucPpe2 [T249M] (SEQ ID NO:25)

M E D K N I L Y G P E P F Y P L A D 20
G T A G E Q M F Y A L S R Y A D I S G C 40
I A L T N A H T K E N V L Y E E F L K L 60
S C R L A E S F K K Y G L K Q N D T I A 80
V C S E N G L Q F F L P L I A S L Y L G 100
I I A A P V S D K Y I E R E L I H S L G 120
I V K P R I I F C S K N T F Q K V L N V 140
K S K L K Y V E T I I I L D L N E D L G 160
G Y Q C L N N F I S Q N S D I N L D V K 180
K F K P N S F N R D D Q V A L V M F S S 200
G T T G V S K G V M L T H K N I V A R F 220
S H C K D P T F G N A I N P T T A I L T 240
V I P F H H G F G M M T T L G Y F T C G 260
F R V A L M H T F E E K L F L Q S L Q D 280
Y K V E S T L L V P T L M A F F A K S A 300
L V E K Y D L S H L K E I A S G G A P L 320
S K E I G E M V K K R F K L N F V R Q G 340
Y G L T E T T S A V L I T P D T D V R P 360
G S T G K I V P F H A V K V V D P T T G 380
K I L G P N E T G E L Y F K G D M I M K 400
G Y Y N N E E A T K A I I N K D G W L R 420
S G D I A Y Y D N D G H F Y I V D R L K 440
S L I K Y K G Y Q V A P A E I E G I L L 460
Q H P Y I V D A G V T G I P D E A A G E 480
L P A A G V V V Q T G K Y L N E Q I V Q 500
N F V S S Q V S T A K W L R G G V K F L 520
D E I P K G S T G K I D R K V L R Q M F 540
E K H K S K L 547

FIG. 45

LucPpL 81-6G1 (SEQ ID NO:26)

M M K R E K N V I Y G P E P L H P L E D 20
L T A G E M L F R A L R K H S H L P Q A 40
L V D V V G D E S L S Y K E F F E A T V 60
L L A Q S L H N C G Y K M N D V V S I C 80
A E N N T R F F I P V I A A W Y I G M I 100
V A P V N E S Y I P D E L C K V M G I S 120
K P Q I V F T T K N I L N K V L E V Q S 140
R T N F I K R I I I L D T V E N I H G C 160
E S L P N G I S R Y S D G N I A N F K P 180
L H F D P V E Q V A A I L C S S G T T G 200
L P K G V M Q T H Q N I C V R L I H A L 220
D P R A G T Q L I P G V T V L V Y L P F 240
F H A F G F S I T L G Y F M V G L R V I 260
M E R R F D Q E A F L K A I Q D Y E V R 280
S V I N V P S V I L F L S K S P L V D K 300
Y D L S S L R E L C C G A A P L A K E V 320
A E V A A K R L N L P G I R C G F G L T 340
E S T S A N I H S L R D E F K S G S L G 360
R V T P L M A A K I A D R E T G K A L G 380
P N Q V G E L C I K G P M V S K G Y V N 400
N V E A T K E A I D D D G W L H S G D F 420
G Y Y D E D E H F Y V V D R Y K E L I K 440
Y K G S Q V A P A E L E E I L L K N P C 460
I R D V A V V G I P D L E A G E L P S A 480
F V V K Q P G K E I T A K E V Y D Y L A 500
E R V S H T K Y L R G G V R F V D S I P 520
R N V T G K I T R K E L L K Q L L E K A 540
G G 542

FIG. 46

LucPpl 81-6G1

ATGATGAAGC GAGAGAAAAA TGTTATATAT GGACCCGAAC CCCTACACCC CTTGGAAGAC
 TTAACAGCTG GAGAAATGCT CTTCCGTGCC CTTCGAAAAC ATTCTCATTT ACCGCAGGC
 TTAGTAGATG TGGTTGGCGA CGAATCGCTT TCCTATAAAG AGTTTTTGAG AGCQACAGTC
 CTCCTAGCGC AAAGTCTCCA CAATTGTGGA TACAAGATGA ATGATGTAGT GTCGATCTGC
 GCCGAGAATA ATACAAGATT TTTTATTCCC GTTATTGAG CTTGGTATAT TGGTATGATT
 GTAGCACCTG TTAATGAAAG TTACATCCC GATGAACCTCT GTAAGGTQAT GGGTATATCG
 AAACCACAAA TAGTTTTAC QACAAAGAAC ATTTTAAATA AGGTATTGGA GGTACAGAGC
 AGAACTAATT TCATAAAAAG GATCATCATA CTTGATACTG TAGAAAACAT ACACGGTTGT
 GAAAGTCTTC CCAATTTTAT TTCTCGTTAT TCGGATGGAA ATATTGCCAA CTTCAAACCT
 TTACATTTCG ATCTGTGTA GCAAGTGGCA GCTATCTTAT GTTCGTCAGG CACTACTGGA
 TTACCGAAAG GTGTAATGCA AACTCACCA AATATTTGTG TCCGACTTAT ACATGCTTTA
 GACCCCAGGG CAGGAACGCA ACTTATTCCCT GGTGTGACAG TCTTAGTATA TCTGCCCTTT
 TTCCATGCTT TTGGGTTCTC TATAACCTTG GGATACTTCA TGGTGGGTCT TCGTGTATC
 ATGTTQAGAC GATTGATCA AGAAGCATTCTT CTTAAAGCTA TTCAGGATTA TGAAGTTCGA
 AGTGTAAATT ACGTTCCATC AGTAATATTG TTCTTATCGA AAAGTCCTTT GGTTGACAAA
 TACGATTITAT CAAGTTTAAG GGAATTGTGT TGCGGTGCGG CACCATTAGC AAAAGAAGTT
 GCTGAGGTTG CAGCAAAACG ATTAAACCTTG CCAGGAATTG GCTGTGGATT TGGTTGACA
 GAATCTACTT CAGCTAATAT ACACAGTCCTT AGGGATGAAT TAAATCAGG ATCACTTGGA
 AGAGTTACTC CTTTAATGGC AGCTAAAATA GCAGATAGGG AAACCTGGTAA AGCATTGGGA
 CCAAATCAAG TTGGTGAATT ATGCQATTAAA GGTCCCCATGG TATCGAAAGG TTACGTGAAC
 AATGTAGAAG CTACCAAAGA AGCTATTGAT GATGATGGTT GGCTTCACTC TGGAGACTTT
 GGATACTATG ATGAGGATGA GCATTCTAT GTGGTGGACCC GTTACAAGGA ATTGATTAAA
 TATAAGGGCT CTCAGGTAGC ACCTGCAGAA CTAGAAGAGA TTTTATTGAA AAATCCATGT
 ATCAGAGATG TTGCTGTGGT TGGTATTCCCT GATCTAGAAG CTGGAGAACT GCCATCTGCG
 TTTGTGGTTA AACAGCCCGG AAAGGAGATT ACAGCTAAAG AAGTGTACGA TTATCTTGCC
 GAGAGGGTCT CCCATACAAA GTATTIGCGT GGAGGGGTTC GATTGTTGA TAGCATACCA
 AGGAATGTTA CAGGTAAAAT TACAAGAAAG GAACTTCTGA AGCAGTTGCT GGAGAAGGGCG
GGAGGT

FIG. 47

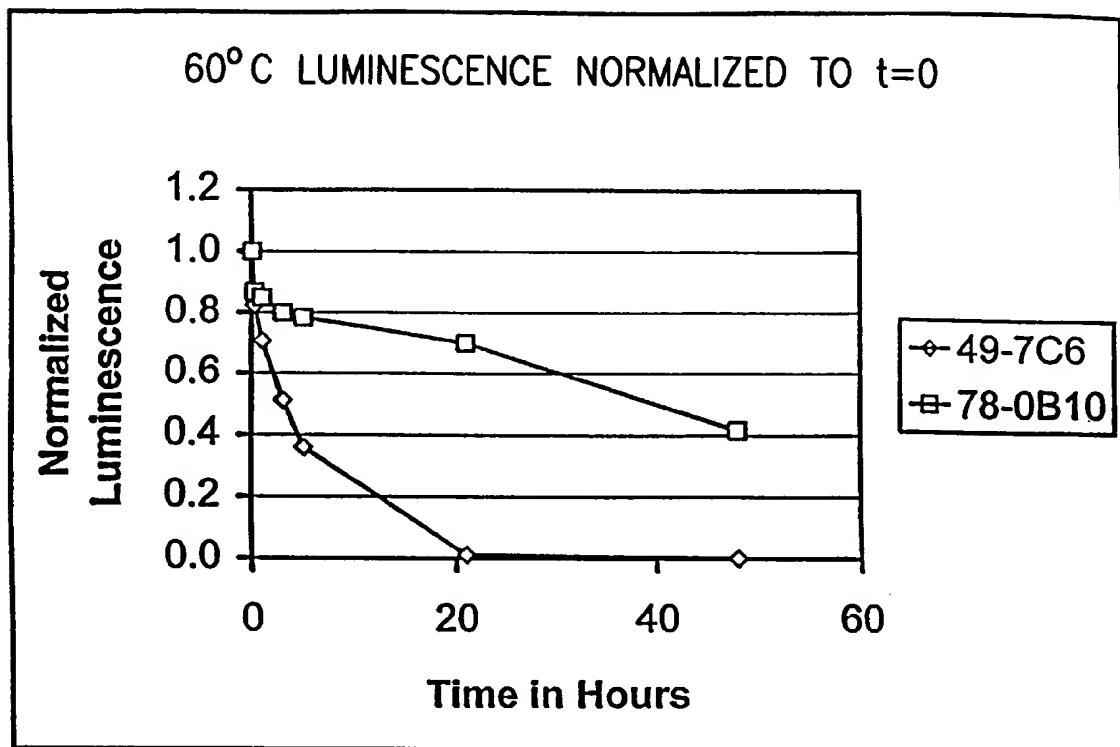


FIG. 48

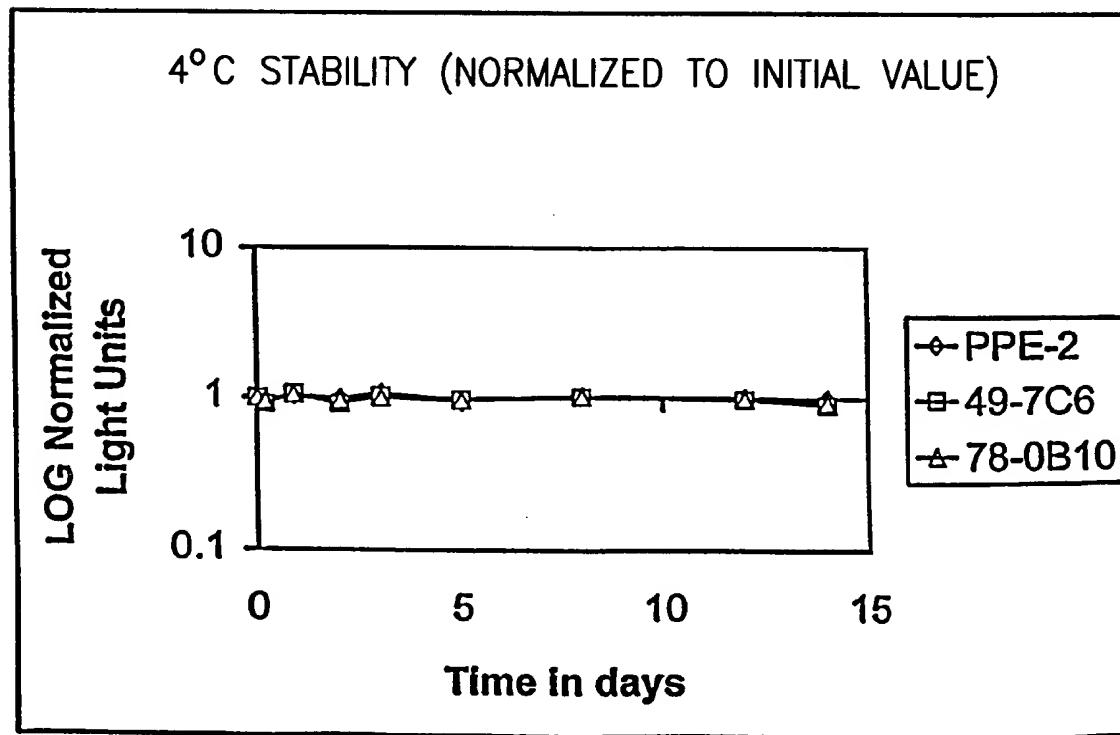


FIG. 49

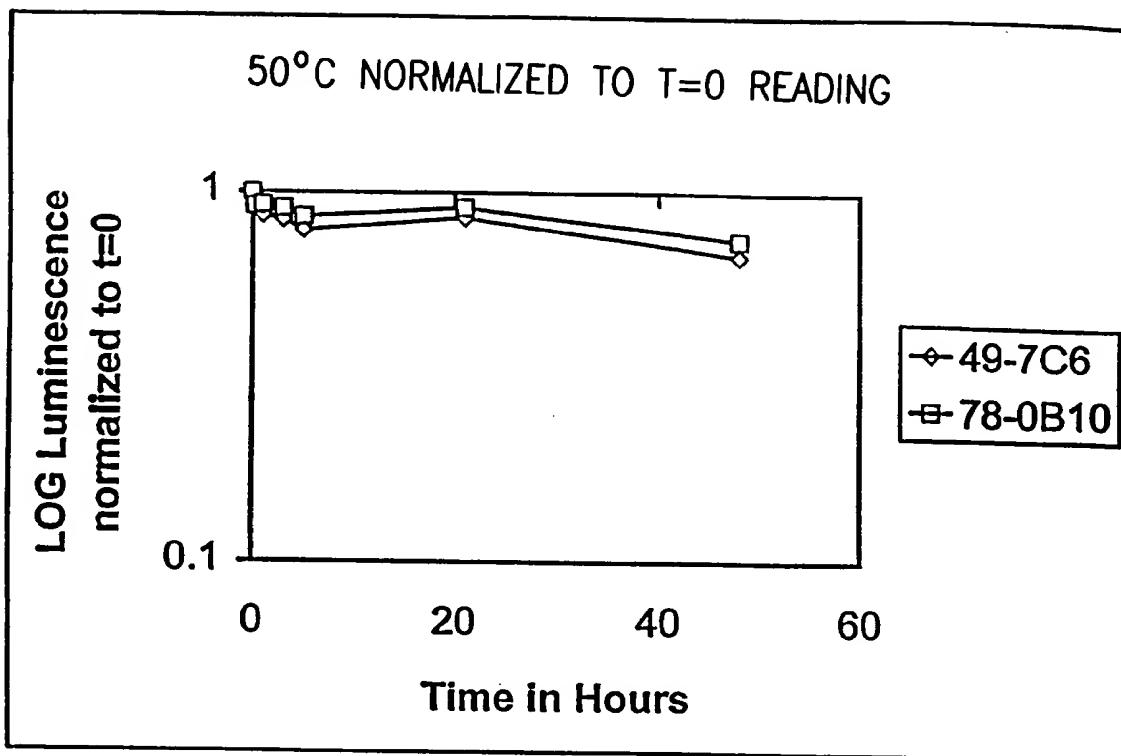


FIG. 50

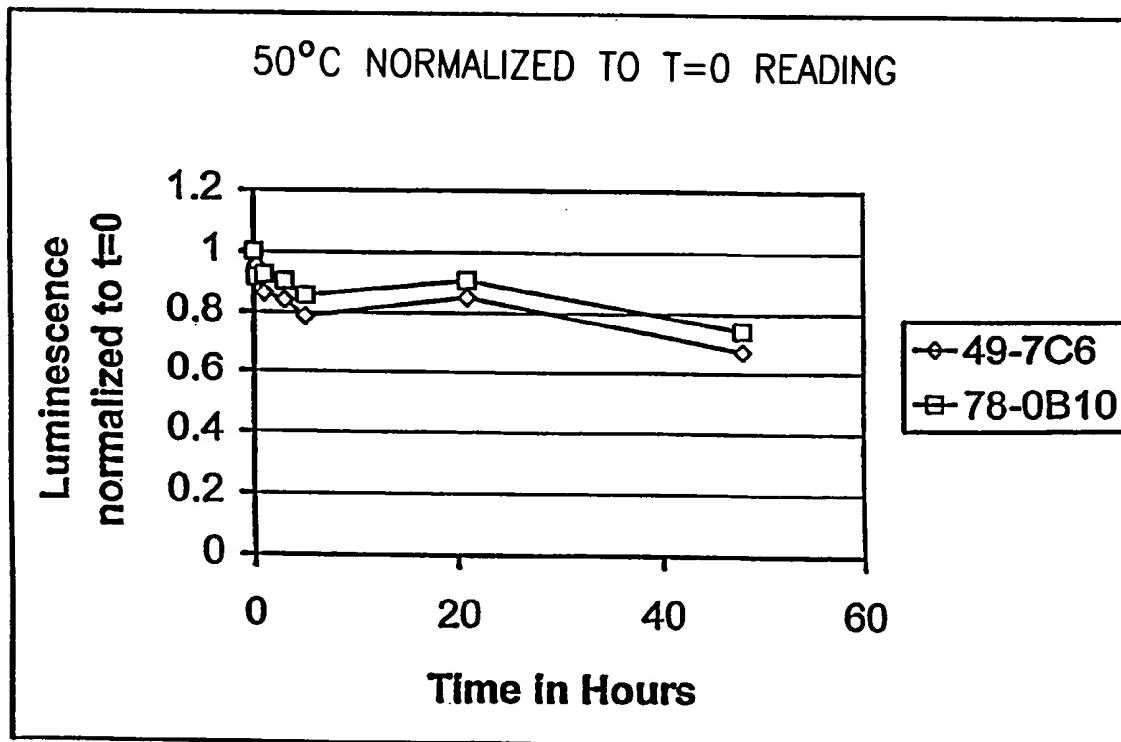


FIG. 51

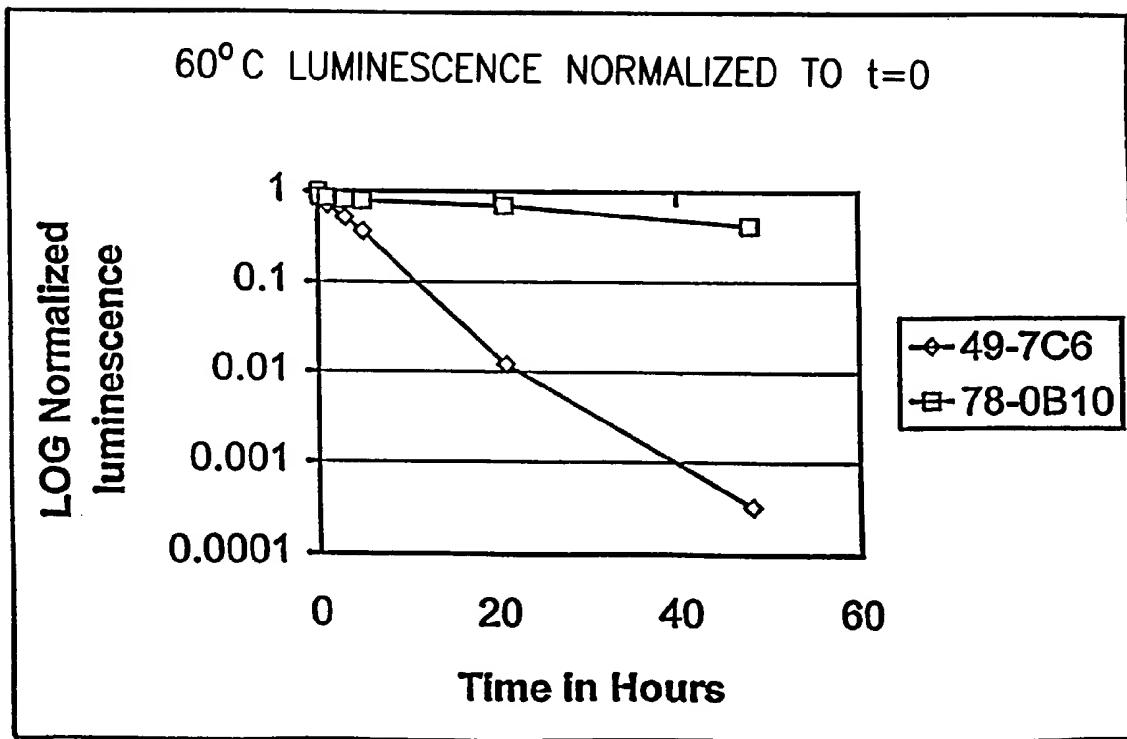


FIG. 52

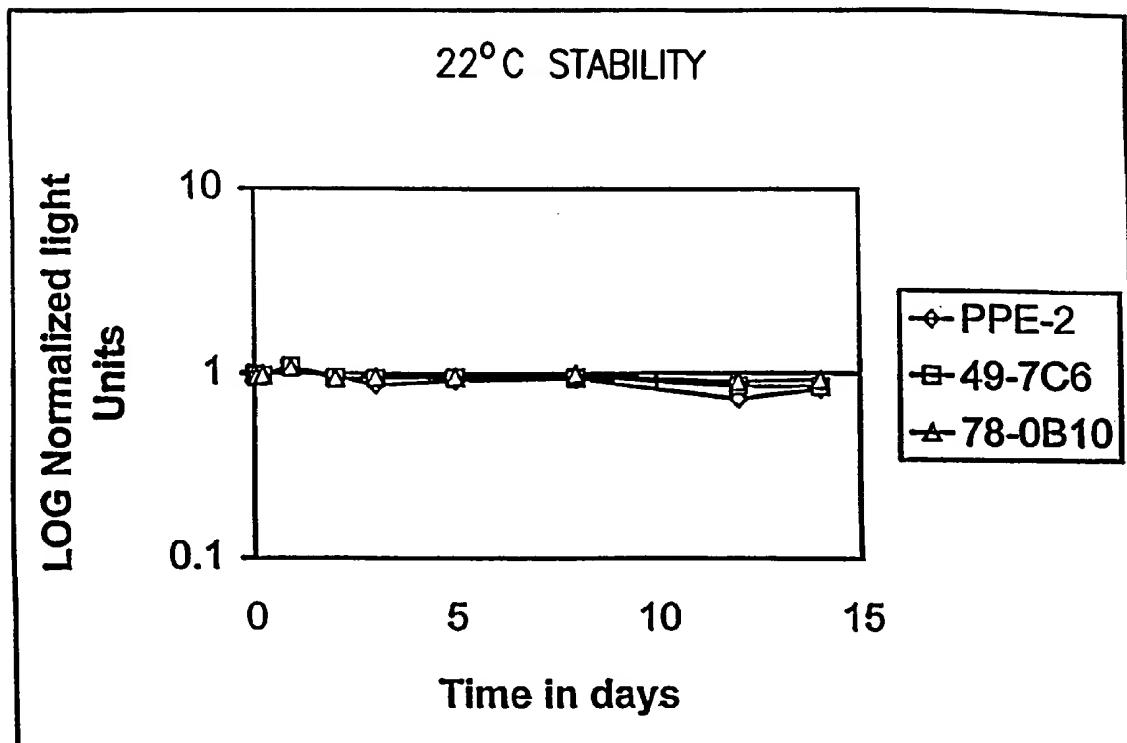


FIG. 53

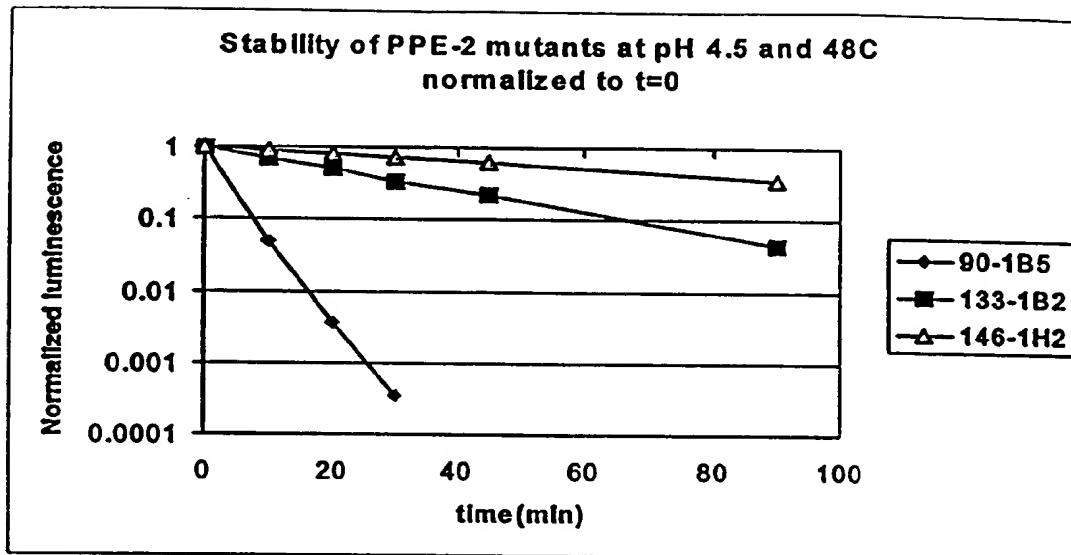


FIG. 54A

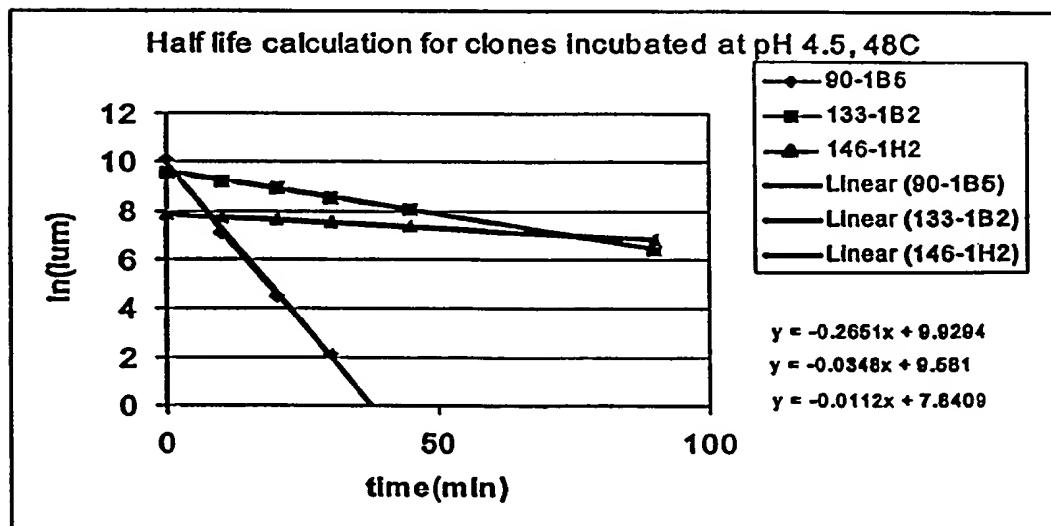


FIG. 54B

Luc133-1B2 (SEQ ID NO:42)

AGATCCAATG	GCAGATAAGA	ATATTTATA	TGGGCCGAA	CCATTTTATC	CCTTGGAGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTQACGC	ATTATCTCGT	TATGCAGATA	TTTGGGCTG	120
CATAGCATG	ACAAATGCTC	ATACAAAAGA	AAATGTTTA	TATGAAGAGT	TTCTGAAACT	180
GTCGTGTCGT	TTAGCGGAAA	GTTTTAAAAA	GTATGGATT	AAACAAAACG	ACACAATAGC	240
GGTGTGTAAC	GAAAATAGTC	TGCAATT	CCTTCCCTGTA	ATTGCATCAT	TGTATCTGG	300
AATAATTGTG	GCACCTGTTA	<u>ACGATAAATA</u>	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGTAAAA	CCACGCATAG	TTTTTGTC	CAAGAATACT	TTTCAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATGTA	TQGAAACTAT	TATTATATA	GACTTAAATG	ATGACTTAGG	480
AGGTTATCAA	TGCCTCAACA	ACTTTATTC	TCAAAATTCC	GATAQTAATC	TQGACGTAAA	540
AAAATTTAAA	CCATATTCTT	TTAATCGAGA	CGATCAGGT	GGGTTGATTA	TGTTTCTTC	600
TGGTACAAC	GGTCTGCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGCACGATT	660
TTCTATTGCA	AAAGATCCTA	CTTTGGTAA	CGCAATTAA	CCACGTCAG	CAATTAAAC	720
GGTAATACT	TTCCACCATG	TTTTGGTAT	GATGACCA	TTAGGATACT	TTACTTGTGG	780
ATTCCGAGTT	GTCTAATGC	ACACGTTGA	AGAAAAACTA	TTTCTACAAT	CATTACAAGA	840
TTATAAAGTG	GAAGTACTT	TACTTGTACC	AACATTAATG	GCATTTCTTG	CAAAAAGTGC	900
ATTAGTTGAA	AAGTACGATT	TATCGCACTT	AAAAGAAATT	GCATCTGGT	GCGCACCTTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTGA	ACGGTTAAA	TTAAACTTG	TCAGGCAAGG	1020
GTATGGATT	ACAGAAACCA	CTTCGGCTGT	TTAATTACA	CGQAAAGGTG	ACGCCAACACC	1080
GGGATCAACT	GGTAAAATAG	TACCATTCA	CGCTGTTAAA	GTTGTCGATC	CTACAACAGG	1140
AAAATTTG	GGGCCAAATG	<u>AAQCTGGAGA</u>	ATTGTATTT	AAAGGCCCGA	TGATAATGAA	1200
GGGTTATTAT	AATAATGAAG	AAGCTACTAA	AGCAATTATT	<u>GATAATGACG</u>	GATGGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATT	TATATTGTGG	ACAGGGCTGAA	1320
GTCACTGATT	AAATATAAAAG	GTTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCG	TATATTGTTG	ATGCCGGCGT	TACTGGTATA	CCGGATGAAG	CCGGGGCGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAATAT	CTAAACGAAC	AAATCGTACA	1500
AGATTATGTT	GCCAGTCAAG	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGATATT	1560
GGATGAAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
AGAAAAACAC	<u>ACCAATCGG</u>					1639

FIG. 55

Luc146-1H2 (SEQ ID NO:43)

GGATCCAATG	GGAGATAAGA	ATATTTATA	TGGGCCGAA	CCATTTATC	CCTTGGAAAGA	60
TGGGACGGCT	GGAGAACAGA	TGTTTGACGC	ATTATCTCGT	TATGCAGCTA	TTCCGGGCTG	120
CATAGCATTG	ACAAATGCTC	ATACAAAAGA	AAATGTTTA	TATGAAGAGT	TTCTGAAACT	180
GTCGTGTCGT	TTAGCGGAAA	GTTTAAAAAA	GTATGGATT	AAACAAAACG	ACACAATAGC	240
GGTGTGTAGC	GAAAATAGTC	TGCAATT	CCTTCCTGTA	ATTGCACTCAT	TGTATCTTGG	300
AATAATTGTG	GCACCTGTTA	<u>ACGATAAATA</u>	CATTGAACGT	GAATTAATAC	ACAGTCTTGG	360
TATTGTAAAA	CCACGCATAG	TTTTTGCTC	CAAGAATACT	TTTCAAAAAG	TACTGAATGT	420
AAAATCTAAA	TTAAAATCTA	TTGAAACTAT	TATTATATT	GACTTAAATG	AAGACTTAGG	480
AGGTTATCAA	TGCCTCAACA	ACTTTATTC	TCAAAATTCC	GATAQTAATC	TGGACGTAAA	540
AAAATTTAAA	CCCTATTCTT	TTAACGAGA	CGATCAGGTT	GCGTQGATTA	TGTTTCTTC	600
TGGTACAAC	GGTCTGCCGA	AGGGAGTCAT	GCTAACTCAC	AAGAATATTG	TTGACGATT	660
TTCTATTGCA	AAAGATCCTA	CTTTGGTAA	CGCAATTAA	CCGACGTCA	CAATTAAAC	720
GGTAATACCT	TTCCACCATG	GTTTGGTAT	GATGACCACA	TTAGGATACT	TTACTTGTGG	780
ATTCCGAGTT	GTTCTAATGC	ACACGTTGA	AGAAAAACTA	TTTCTACAA	CATTACAAGA	840
TTATAAAGTG	GAAAGTACTT	TACTTGTACC	AACATTAATG	GCATTTCTTG	CAAAAAGTGC	900
ATTAGTTGAA	AAGTACGATT	TATCGCATT	AAAAGAAATT	GCATCTGGTG	GCGCACCTTT	960
ATCAAAAGAA	ATTGGGGAGA	TGGTGAAAAA	ACGGTTAAA	TTAAACTTTG	TCAGGCAAGG	1020
GTATGGATTA	ACAGAAACCA	CTTCGGCTGT	TTTAATTACA	CCGAAAGGTG	ACGGCAAAACC	1080
GGGATCAACT	GGTAAAATAG	TACCAATTCA	CGCTGTTAAA	GTTGTCGATC	CTACAACAGG	1140
AAAAATTG	GGGCCAAATG	AAQCTGGAGA	ATTGTATT	AAAGGCCGA	TGATAATGAA	1200
GGGTATTAT	AATAATGAAG	AAGCTACTAA	AGCAATTATT	GATAATGACG	GATGGTTGCG	1260
CTCTGGTGAT	ATTGCTTATT	ATGACAATGA	TGGCCATT	TATATTGTGG	ACAGGCTGAA	1320
GTCACTGATT	AAATATAAAAG	GTTATCAGGT	TGCACCTGCT	GAAATTGAGG	GAATACTCTT	1380
ACAACATCCG	TATATTGTTG	ATGCCGGGT	TACTGGTATA	CCGGATGAAG	CCGGGGCGA	1440
GCTTCCAGCT	GCAGGTGTTG	TAGTACAGAC	TGGAAAATAT	CTAACCGAAC	AAATCGTACA	1500
AGATTATGTT	<u>GCCAGTCAAG</u>	TTTCAACAGC	CAAATGGCTA	CGTGGTGGGG	TGAAATT	1560
GGATGAATT	CCCAAAGGAT	CAACTGGAAA	AATTGACAGA	AAAGTGTAA	GACAAATGTT	1620
AGAAAACAC	<u>ACCAATGGG</u>					1639

FIG. 56

Luc133-1B2 (SEQ ID NO:44)

G	T	A	G	E	Q	M	F	D	A	L	S	R	P	E	P	F	Y	P	I	P	L	E	D	20
I	A	L	T	N	A	H	T	K	E	N	V	L	Y	E	E	F	F	A	E	E	G	C	40	
S	C	R	L	A	E	S	F	K	K	Y	G	L	K	Q	N	D	T	I	A	K	L	60		
V	C	S	E	N	S	L	Q	F	F	L	P	V	I	A	S	L	Y	L	G	L	80			
I	I	Y	A	P	V	N	D	K	Y	I	E	R	E	L	I	H	S	S	L	G	100			
I	V	K	P	R	I	V	F	C	S	K	N	T	F	Q	K	V	L	N	V	N	V	120		
K	S	K	L	K	S	I	E	T	I	I	I	L	D	L	N	D	D	D	L	L	G	140		
G	Y	Q	C	L	N	N	F	I	S	Q	N	S	D	S	N	N	L	D	D	V	K	160		
K	F	K	P	Y	S	F	N	R	D	D	Q	V	A	L	I	M	F	S	S	F	180			
G	T	T	G	L	P	E	K	G	V	M	L	T	H	K	N	I	V	A	R	R	F	200		
S	I	A	K	D	P	T	F	G	N	A	I	N	P	T	S	E	A	I	L	C	T	220		
V	I	P	F	H	H	G	F	G	M	M	T	T	L	G	Y	F	T	T	C	G	240			
F	R	R	V	Y	L	M	H	T	F	E	E	K	L	F	L	Q	S	L	Q	D	260			
Y	K	V	E	S	T	L	L	V	P	T	L	M	A	F	L	E	A	K	S	A	280			
L	V	E	K	Y	D	L	L	S	H	L	K	E	I	A	S	G	G	A	P	L	300			
S	K	E	I	G	E	M	V	K	K	R	F	K	L	N	F	V	R	Q	G	320				
Y	G	L	T	E	T	T	S	A	V	L	I	T	P	K	E	D	A	K	P	340				
G	S	T	G	K	I	V	P	F	H	A	V	K	V	V	D	P	T	T	G	360				
K	I	L	G	P	N	E	E	P	G	E	L	Y	F	K	G	I	M	I	M	K	380			
G	Y	Y	N	N	E	E	A	T	K	A	I	I	D	N	D	D	G	W	L	R	400			
S	G	D	I	A	Y	Y	D	N	D	G	H	F	Y	I	E	E	A	A	G	E	420			
S	L	I	K	Y	K	G	Y	Q	V	A	P	A	E	I	E	G	I	L	L	L	440			
Q	H	P	Y	I	V	D	A	G	V	T	G	I	I	P	D	E	A	A	G	E	460			
L	P	A	A	G	V	V	V	Q	T	G	K	Y	L	N	E	Q	I	V	Q	500				
D	Y	V	A	S	Q	V	S	T	A	K	W	L	R	G	G	V	I	F	L	520				
D	E	I	P	K	G	S	T	G	K	I	D	R	K	V	L	R	Q	M	L	540				
E	K	H	T	N	G																			

FIG. 57

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Luc146-1H2 (SEQ ID NO:45)

G	T	A	D	K	N	I	L	Y	G	P	E	P	F	Y	P	L	E	D	20
I	A	L	E	Q	M	F	D	A	L	S	R	Y	A	A	P	G	C	40	
S	C	R	L	A	H	T	K	E	N	V	L	Y	E	F	P	L	K	60	
V	C	S	E	N	S	F	K	K	Y	G	L	K	Q	N	D	T	I	80	
I	I	V	A	P	V	N	Q	F	F	L	P	V	I	A	S	L	Y	100	
I	V	K	P	R	I	V	F	C	S	K	N	T	F	Q	K	V	L	120	
K	S	K	L	K	S	I	E	T	I	I	I	L	D	L	N	V	140		
G	Y	Q	C	L	N	N	F	I	S	Q	N	S	D	S	N	D	V	160	
K	F	K	P	Y	S	F	N	R	D	D	Q	V	A	S	I	M	F	180	
G	T	T	G	L	P	K	G	V	M	L	T	H	K	N	I	V	R	200	
S	I	A	K	D	P	T	F	G	N	A	I	N	P	T	S	A	I	220	
V	I	P	F	H	H	G	F	G	M	M	T	T	L	G	Y	F	T	240	
F	R	V	V	L	M	H	T	F	E	E	K	L	F	Q	S	L	Q	260	
Y	K	V	E	S	T	L	L	V	P	T	L	M	A	F	L	A	K	280	
L	V	E	K	Y	D	L	S	H	L	K	E	I	A	S	N	F	V	300	
S	K	E	I	G	E	M	V	K	K	R	F	K	L	G	D	A	P	320	
Y	G	L	T	E	T	T	S	A	V	L	I	T	P	K	G	D	A	340	
G	S	T	G	K	I	V	P	L	H	A	V	K	V	D	P	T	T	360	
K	I	L	G	P	N	E	E	P	G	E	L	Y	F	K	G	P	M	380	
G	Y	Y	N	N	E	E	P	A	T	K	A	I	I	D	N	D	W	400	
S	G	D	I	A	Y	Y	D	N	D	G	H	F	Y	I	V	D	R	420	
S	L	I	K	Y	K	G	Y	Q	V	A	P	A	E	I	E	G	I	440	
Q	H	P	Y	I	V	D	A	G	V	T	G	I	P	D	E	A	A	460	
L	P	A	A	G	V	V	Q	T	G	K	Y	L	N	E	Q	I	V	480	
D	Y	V	A	S	Q	V	S	T	A	K	W	L	R	G	G	V	K	500	
D	E	I	P	K	G	S	T	G	K	I	D	R	K	V	L	R	Q	520	
E	K	H	T	N	G												M	L	540
																			546

FIG. 58

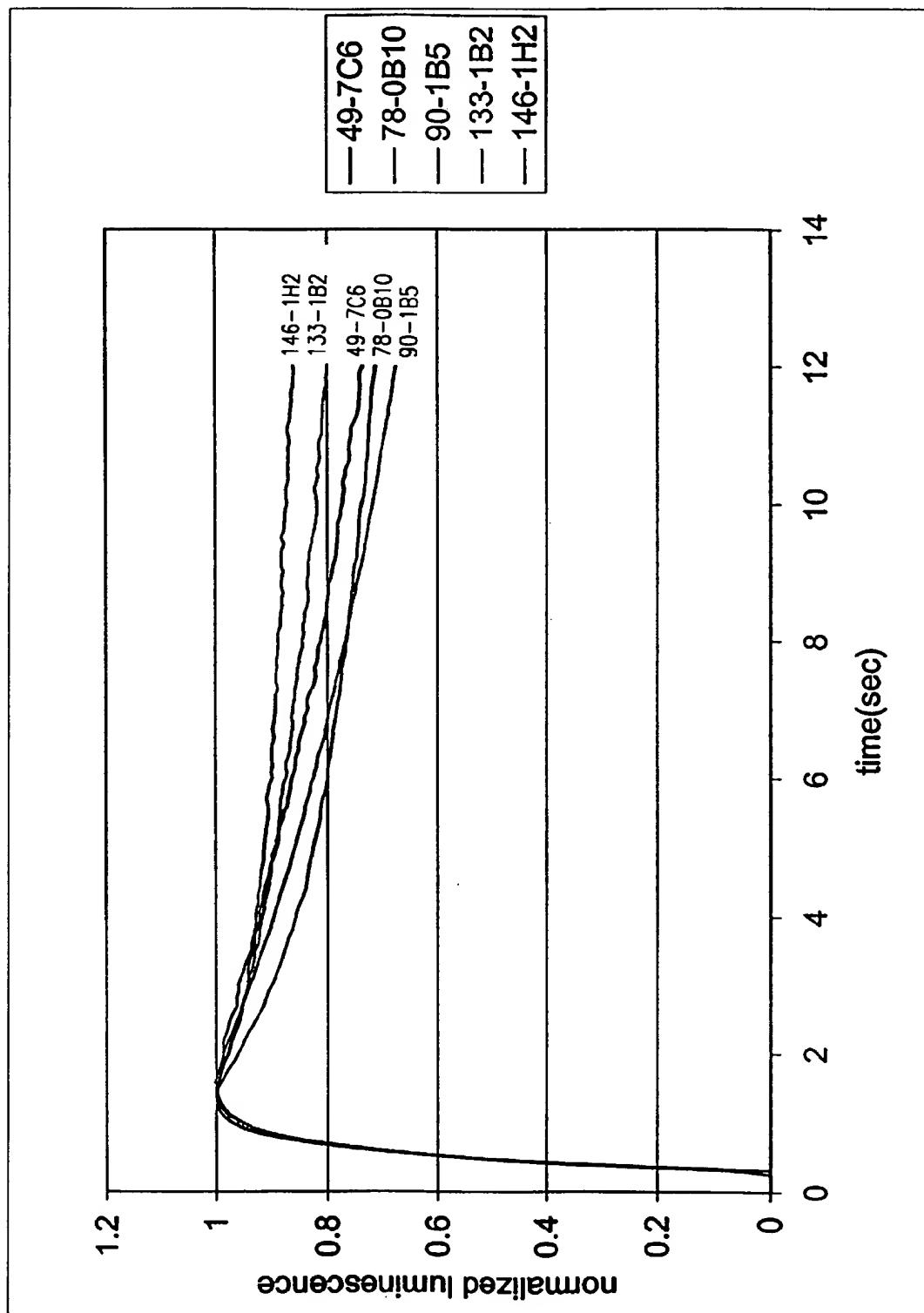


FIG. 59

65/70

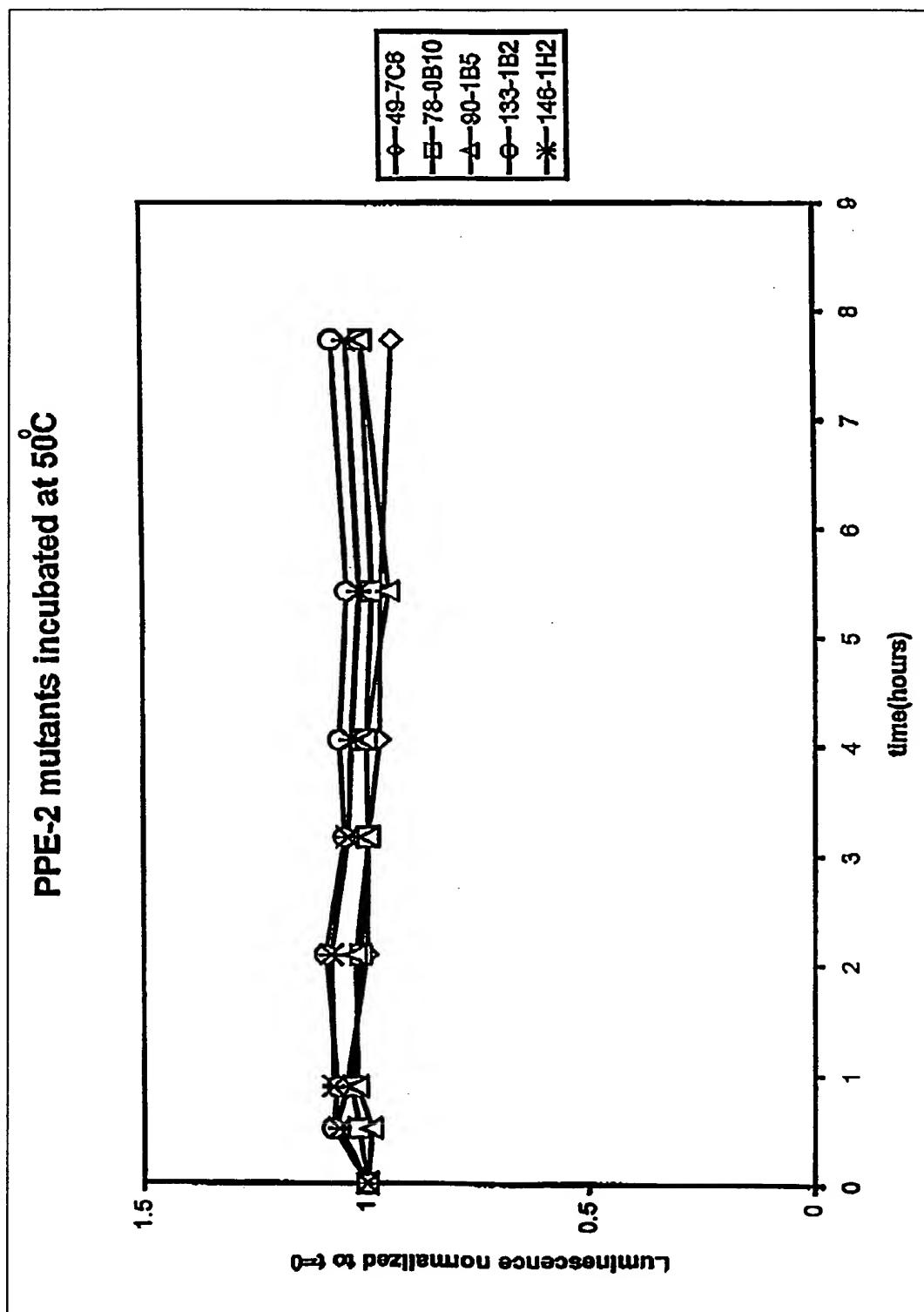


FIG. 60

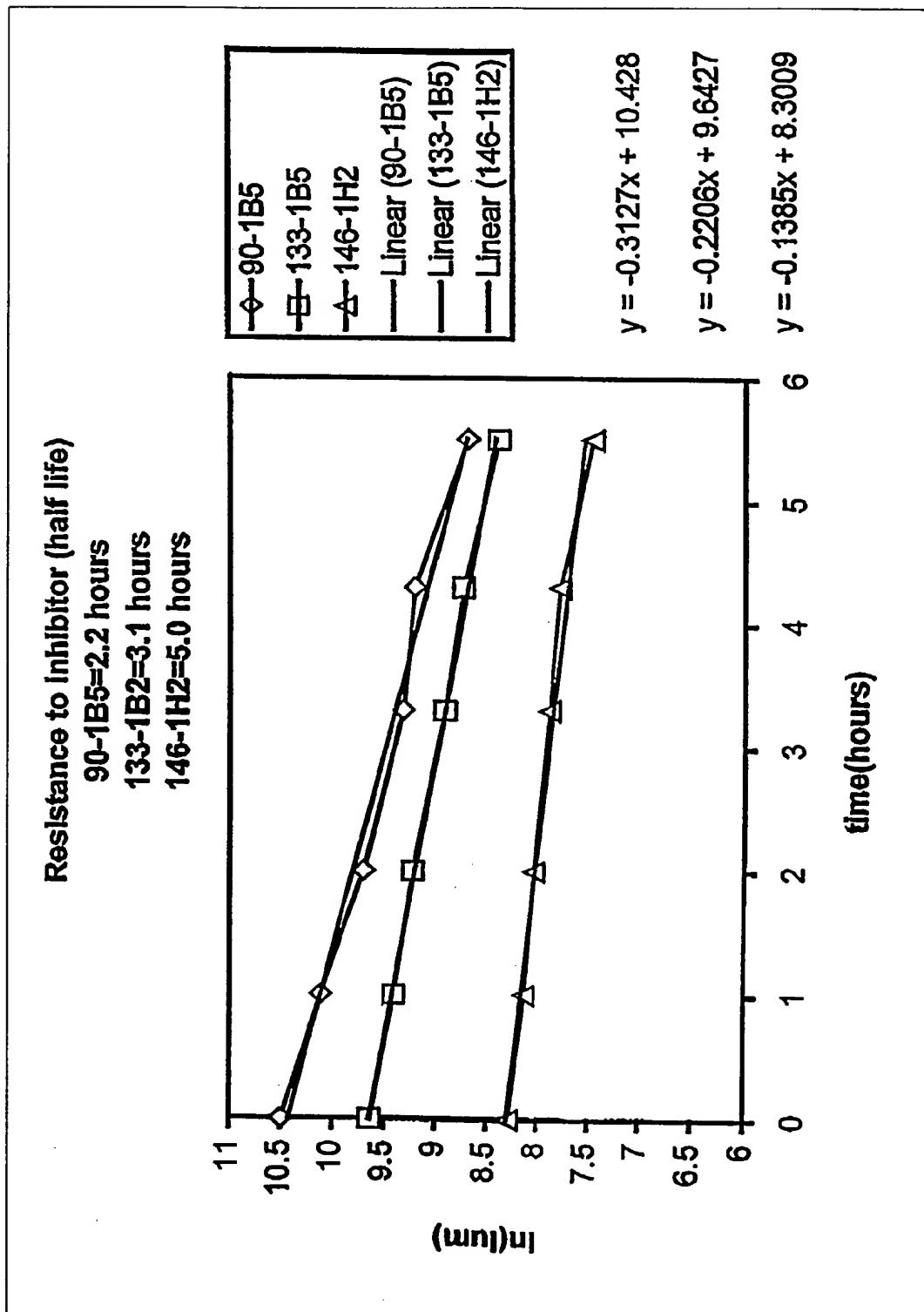


FIG. 61

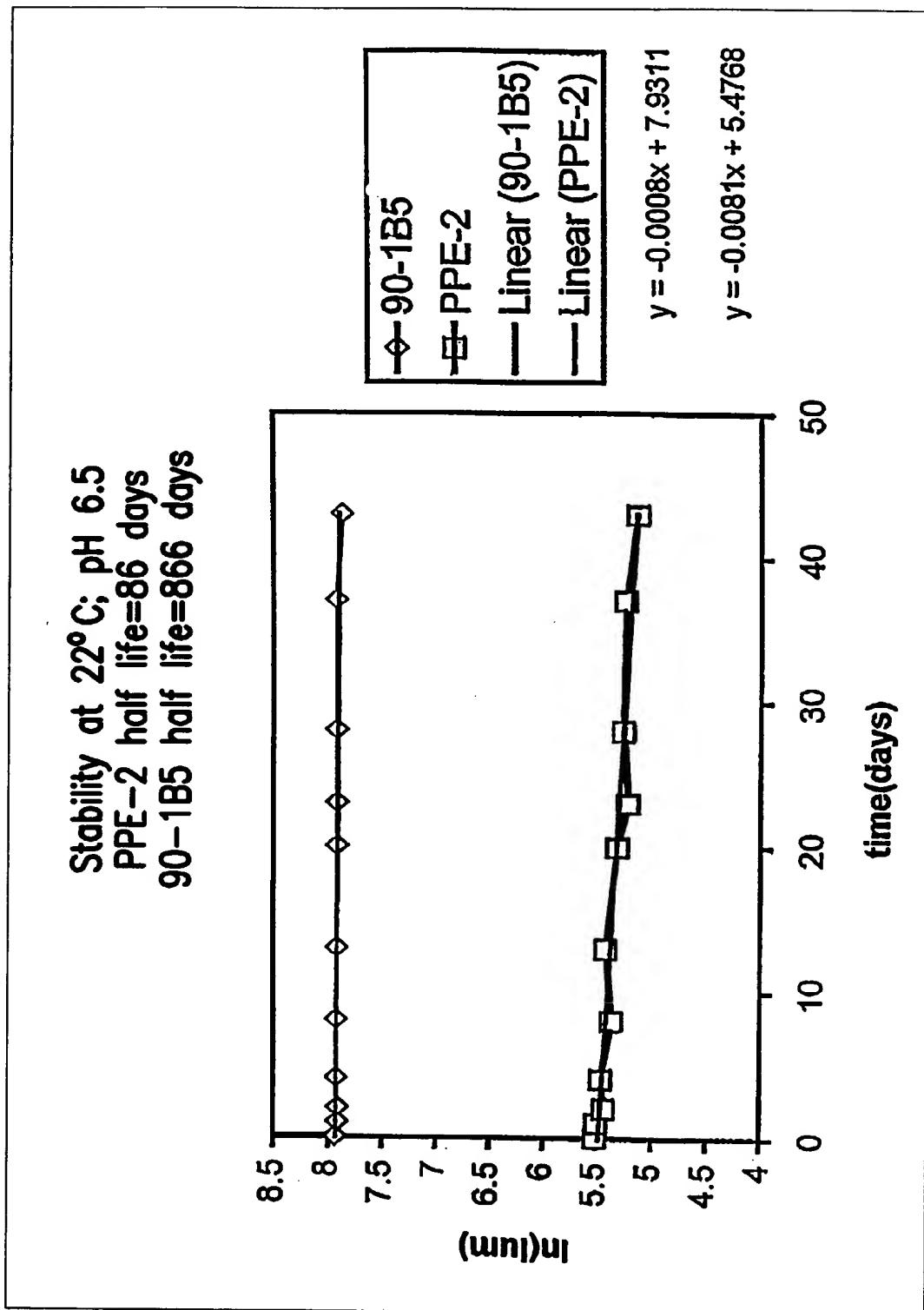


FIG. 62

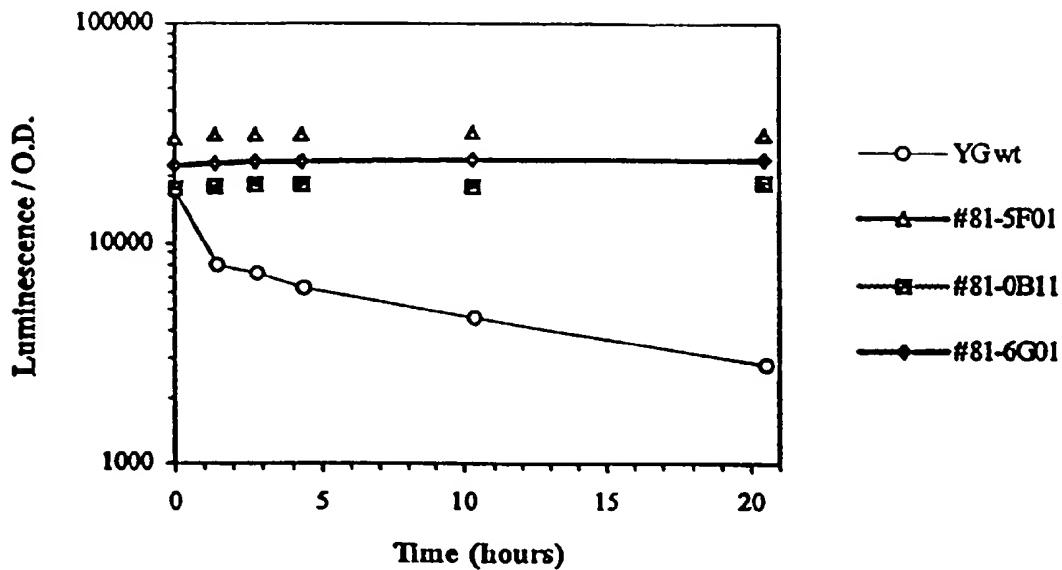


FIG. 63

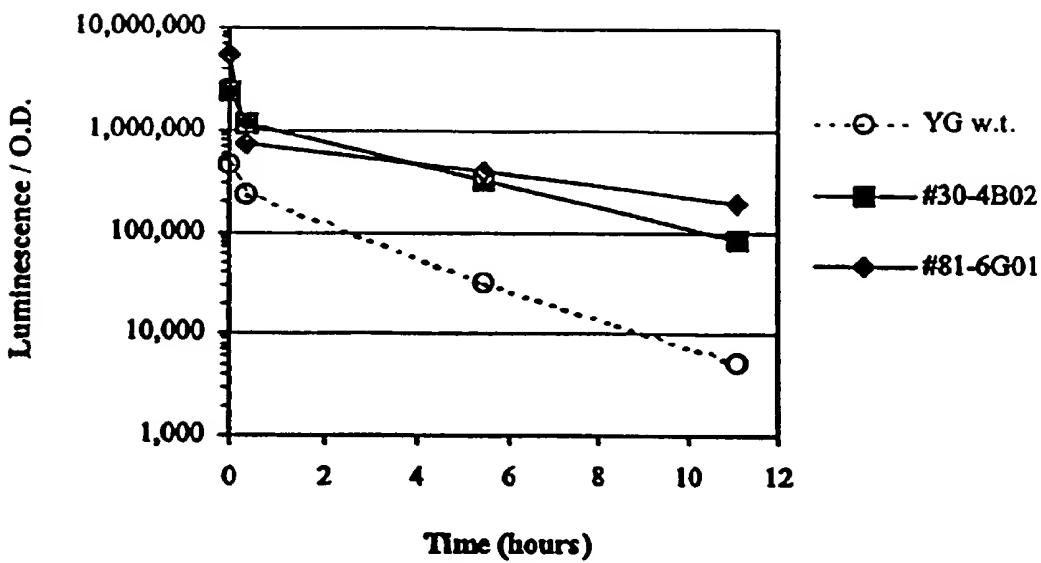


FIG. 64

LucPp181-0B11

GGATCCCAGT ATGAAGCGGAG AGAAAAATGT TATATATGGA CCCGAACCCC TACACCCCTT 60
 GGAAGACTTA ACAGCTGGAG AAATGCTCTT CGGTGCCCTT CGAAAACATT CTCATTTACC 120
 GCAGGGCTTTA GTAGATGTGG TTGGCGACGA ATCGCTTCC TATAAAGAGT TTTTTGAAGC 180
GACAGTCCCTC CTAGCCAAA GTCTCCACAA TTGTGGATAC AAGATGAATG ATGTAGTGTC 240
 GATCTGCGCC GAGAATAATA CAAGATTTT TATTCCCGTT ATTGCAGCTT GGTATATTGG 300
 TATGATTGTA GCACCTGTTA ATGAAAGTTA CATCCCAGAT GAACTCTGTA AGGTCAGGG 360
 TATATCGAA CCACAAATAG TTTTTACGAC AAAGAACATT TAAATAAGG TATTGGAGGT 420
 ACAGAGCAGA ACTAATTTCA TAAAAAGGAT CATCGTACTT GATACTGTAG AAAACATACA 480
 CGGTTGTGAA AGTCTTCCC ATTTTATTTC TCGTTATTCG GATGGAAATA TTGCCAACT 540
 CAAACCTTTA CATTTCGATC CTGTAGGAC AGTGGCAGCT ATCTTATGTT CGTCAGGCAC 600
 TACTGGATT ACGAAAGGTG TAATGCAAAC TCACCAAAAT ATTTGTGTCC GACTTATACA 660
 TGCTTTAGAC CCCAGGGCAG GAACGCAACT TATTCCCTGGT GTGACAGTCT TAGTATATCT 720
 GCCTTTTTTC CATGCTTTTG GGTTCCTCTAT AACCTTGGGA TACTTCATGG TGGGTCCTCG 780
 TGTTATCATG TCAAGACGAT TTGATCGAGA AGCATTCTA AAAGCTATTTC AGGATTATGA 840
 AGTTCGAAAGT GTAATTAAACG TTCCATCAGT AATATTGTT TTATCGAAAAA GTCCTTGGT 900
 TGACAAATAC GATTTATCAA GTTTAAGGGA ATTGTGTTGC GGTGCGGCAC CATTAGCAAA 960
 AGAAGTTGCT GAGGTTGCAG CAAAACGATT AAACTTGCCA GGAATTCGCT GTGGATTTTGG 1020
 TTTGACAGAA TCTACTTCAG CTAATATACA CAGTCTTAGG GATGAATTTTA ACCCAGGATC 1080
 ACTTGGAAAGA GTTACTCCTT TAATGGCAGC TAAAATAGCA GATAGGGAAA CTGGTAAAGC 1140
 ATTGGGACCA AATCAAGTTG GTGAATTATG CATTAAAGGT CCCATGGTAT CGAAAGGTTA 1200
 CGTGAACAAT GTAGAAGCTA CCAAAGAAGC TATTGATGAT GATGGTTTGC TTCACTCTGG 1260
 AGACTTTGGA TACTATGATG AGGATGAGCA TTTCTATGTG GTGGACCGTT ACAAGGAATT 1320
 GATTAAATAT AAGGGCTCTC AGGTAGCACC TGCAGAACTA GAAGAGATT TATTGAAAAAA 1380
 TCCATGTATC AGAGATGTTG CTGTGGTTGG TATTCCCTGAT CTAGAAGCTG GAGAACTGCC 1440
 ATCTGCGTTT GTGGTTAAAC AGCCCGGAAA GGAGATTACA GCTAAAGAAG TGTACGATTA 1500
 TCTTGCCGAG AGGGTCTCCC ATACAAAGTA TTTCGGTGGA GGGGTTCGAT TCGTTGATAG 1560
 CATACCCAGG AATGTTACAG GTAAAATTAC AAGAAAGGAA CTTCTGAAGC AGTTGCTGG 1620
GAAGGCGGGA GGT

FIG. 65

LucPpl 81-0B11

E	D	M	M	K	R	E	K	N	V	I	Y	G	P	E	P	L	H	P	L	18	
Q	A	L	V	D	G	E	M	L	F	R	A	L	R	K	H	S	H	L	P	38	
T	V	L	L	A	Y	S	G	D	E	S	L	S	Y	K	M	F	F	E	A	58	
I	C	A	E	N	N	T	R	H	N	C	G	Y	K	M	N	D	V	V	S	78	
M	I	V	A	P	V	N	E	F	F	I	P	P	D	E	L	C	K	Y	I	G	98
I	S	K	P	Q	I	V	F	T	T	K	N	I	L	N	K	V	L	E	V	138	
Q	S	R	T	N	F	I	K	R	I	I	V	L	D	T	V	E	N	I	H	158	
G	C	E	S	L	P	N	F	I	S	R	Y	S	D	G	N	I	A	N	F	178	
K	P	L	H	E	D	P	V	E	Q	V	A	A	I	L	C	S	S	G	T	198	
T	G	L	P	K	G	V	M	Q	T	H	Q	N	I	C	V	R	L	I	H	218	
A	L	D	P	R	A	G	G	T	Q	L	I	P	G	V	T	V	L	V	Y	238	
P	F	F	H	A	F	F	F	S	I	T	L	G	Y	F	M	V	G	L	R	258	
V	I	M	S	R	R	F	D	P	E	A	F	L	K	A	I	Q	D	Y	E	278	
V	R	S	V	I	N	V	P	S	V	I	L	F	L	S	K	S	P	L	V	298	
D	K	Y	D	L	S	S	L	R	E	L	C	C	G	A	A	P	L	A	K	318	
E	V	A	E	V	A	A	K	R	L	N	L	P	G	I	R	C	G	F	G	338	
L	T	E	S	T	S	A	A	N	I	H	S	L	R	D	E	F	K	P	G	358	
L	G	R	V	T	P	L	M	A	A	K	I	A	D	R	E	T	G	K	A	378	
L	G	P	N	Q	V	G	E	L	C	I	K	G	P	M	V	S	K	G	Y	398	
V	N	N	V	E	A	T	K	E	A	I	D	D	G	D	R	Y	H	S	G	418	
D	F	G	Y	Y	D	E	D	E	H	F	Y	V	V	D	R	Y	K	E	L	438	
I	K	Y	K	G	S	Q	V	A	P	A	E	L	E	E	I	L	L	K	N	458	
P	C	I	R	D	V	A	V	V	G	I	P	D	L	E	A	G	E	L	P	478	
S	A	F	V	V	K	Q	P	G	K	E	I	T	A	K	E	V	Y	D	Y	498	
L	A	E	R	V	S	H	T	K	Y	L	R	G	G	V	R	F	V	D	S	518	
I	P	R	N	V	T	G	K	I	T	R	K	E	L	L	K	Q	L	L	E	538	
K	A	G	G																	542	

FIG. 66

SEQUENCE LISTING

<110> Promega Corporation

<120> Thermostable luciferases and methods of production.

<130> 341.012WO1

<150> US 09/396,154

<151> 1999-09-15

<150> US 09/156,946

<151> 1998-09-18

<150> PCT/US98/19494

<151> 1998-09-18

<150> US 60/059,379

<151> 1997-09-19

<160> 93

<170> FastSEQ for Windows Version 3.0

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<211> 1639

<212> DNA

<213> Artificial Sequence

<220>

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catagcattg acaaattgttc atacaaaaga aaatgttttata tatgaagagt ttttaaaatt	180
gtcgtgtcgat tttagcgaaa gttttaaaaaa gtatggatta aaacaaaacg acacaatagc	240
gggtgttagc gaaaatggtt tgcaattttt ctttccata attgcattat tttatcttgg	300
aataattgca gcacctgtta gtgataaata cattgaacgt gaattaatac acagtcttgg	360
tattgtaaaa ccacgcataa ttttttgctc caagaataact tttcaaaaag tactgaatgt	420
aaaatctaaa ttaaaatatg tagaaactat tattatatta gacttaaatg aagacttagg	480

aggttatcaa tgcctcaaca actttatttc tcaaattcc gatattaatc tggacgtaaa	540
aaaatttaaa ccatattctt ttaatcgaga cgatcagggtt gcgttggtaa tgtttcttc	600
tggtacaact ggtgttcga agggagtcac gctaactcac aagaatattg ttgcacgatt	660
ttctcttgca aaagatccta cttttggtaa cgcaattaat ccaacgacag caatttaac	720
ggtaatacct ttccaccatg gttttggat gatgaccaca ttaggatact ttacttgtgg	780
attccgagtt gttctaattgc acacgtttga agaaaaacta tttctacaat cattacaaga	840
ttataaaagtg gaaagtactt tacttgtacc aacattaatg gcatttcttg caaaaagtgc	900
attagttgaa aagtacgatt tatcgcactt aaaagaaaatt gcatctggtg gcgcacctt	960
atcaaaaagaa attggggaga tggtaaaaa acggttaaa ttaaactttg tcagggcaagg	1020
gtatggatta acagaaacca cttcggctgt tttaattaca ccaaacaatg acgtcagacc	1080
gggatcaact ggtaaaatag taccatttca cgctgttaaa gttgtcgatc ctacaacagg	1140
aaaaattttg gggccaaatg aacctggaga attgtatttt aaaggcgaca tgataatgaa	1200
aggttattat aataatgaag aagctactaa agcaattatt aacaaagacg gatggttgcg	1260
ctctggtgat attgcttatt atgacaatga tggccatttt tatattgtgg acaggctgaa	1320
gtcattaatt aaatataaag gttatcaggt tgcacctgct gaaattgagg gaatacttt	1380
acaacatccg tatattgttg atgcggcggt tactggata ccggatgaag ccgcggcgaa	1440
gctccagct gcaggtgttg tagtacagac tggaaaatat ctaaacgaac aaatcgata	1500
aaattttgtt tccagtcaag ttcaacacg caaatggcta cgtgggggg tgaaaatttt	1560
ggatgaaatt cccaaaggat caactggaaa aattgacaga aaagtgttaa gacaaatgtt	1620
tggaaaacac accaatggg	1639

<210> 2

<211> 1639

<212> DNA

<213> Artificial Sequence

<220>

<223> note = "mutant luciferase"

<400> 2

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tgggacggct ggagaacaga tggggatcgat attatctcgat tatgcagata tttcaggatg	120
catagcattt gacaaatgctc atacaaaatc ccctgttttata tatgaagagt tgtaaaaaatt	180
gtcgtgtcgat ttagcggaaa gttttaaaaaa gtatggatta aaacaaaacg acacaatagc	240
ggtgtgttagc gaaaatggtt tgcattttttt ccttcctata attgcattat tgcattttgg	300
aataatttgcgat gcacctgtta gtgataaata cattgaacgt gaattaatac acgttgg	360
tattgtaaaa ccacgcataa tttttgtctc caagaataact tttcaaaaaag tactgaatgt	420
aaaatctaaa ttaaaatatg tagaaaactat tattatatta gactttaaatg aagacttgg	480
aggttatcaa tgcctcaaca actttatttc tcaaattcc gatattaatc tggacgtaaa	540
aaaatttaaa ccatattctt ttaatcgaga cgatcagggtt gcgttggtaa tgtttcttc	600
tggtacaact ggtgttcga agggagtcac gctaactcac aagaatattg ttgcacgatt	660
ttctcttgca aaagatccta cttttggtaa cgcaattaat ccaacgacag caatttaac	720

ggtaataacct ttccaccatg gttttggtat gatgaccaca ttaggatact ttacttgtgg 780
attccgagtt gttctaatgc acacgttga agaaaaacta tttctacaat cattacaaga 840
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acaacatccg tatattgttg atgcccgcgt tactggtata ccggatgaag ccgcggcga 1440
gcttccagct gcaggtgtt tagtacagac tggaaaatat ctaaacgaac aaatcgtaca 1500
aaatTTTgtt tccagtcaag tttcaacagc caaaatggcta cgtgggggg tgaaaatttt 1560
ggatgaaatt cccaaaggat caactggaaa aattgacaga aaagtgttaa gacaaatgtt 1620
tqaaaaaacac accaatqgg 1639

<210> 3

<211> 1639

<212> DNA

<213> Artificial Sequence

<220>

<223> note = "mutant luciferase"

<400> 3

ggatccaatg gaagataaaaa atattttata tggacctgaa ccattttatc ctttggctga 60
tgggacggct ggagaacaga tgaaaaacgc attatctcg tatacgatata ttccaggatg 120
catagcattg acaaatgctc atacaaaagc ccctgttttata tataagagt tttaaaatt 180
gtcggtcg ttagcggaaa gttttaaaaa gtatggatta aaacaaaacg acacaatagc 240
ggtgtgttagc gaaaatggtt tgcaattttt ccttcctata attgcatcat tttatcttgg 300
aataattgca gcacctgtta gtgataaata cattgaacgt gaattaatac acagtcttgg 360
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acaacatccg tatattgtt atgcggcgt tactggtata ccggatgaag ccgcggcga	1440
gctccagct gcaggggttg tagtacagac tggaaaatat ctaaacgaac aaatcgata	1500
aaattttgtt tccagtcag tttcaacacg caaatggcta cgtggtgggg tgaaaatTTT	1560
ggatgaaatt cccaaaggat caactggaaa aattgacaga aaagtgttaa gacaaatgtt	1620
tgaaaaacac accaatggg	1639

<210> 4

<211> 1639

<212> DNA

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 4

ggatccaaatg gaagataaaa atattttata tggacctgaa ccattttatc cttggctga	60
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catagcattg acaaATGCTC atacaaaAGC ccctgttttA tatgaagagt tttaaaatt	180
gtcgtgtcgT tttagcgaaa gttttaaaaa gtatggatta aaacaaaacg acacaatagc	240
ggtgtgtagc gaaaatggTT tgcaattttt cttccata attgcatacat tgcatactgg	300
aataattgca gcacctgttA gtgataaata cattgaacgt gaattaatac acagtcttgg	360
tattgtaaaa ccacgcataa tttttgctc caagaatact tttcaaaaag tactgaatgt	420
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atcaaaaagaa attggggaga tggtaaaaa acggttaaa ttaaactttg tcaggcaagg	1020
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aggttattat aataatgaag aagctactaa agcaattatt aacaaagacg gatggttgcg	1260
ctctggtgat attgcttatt atgacaatga tggccatttt tatattgtgg acaggctgaa	1320
gtcattaatt aaatataaag gttatcaggt tgcacctgct gaaattgagg gaatacttt	1380
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aaattttgtt tccagtcag tttcaacagc caaatggcta cgtggtgggg tgaaattttt	1560
gatgaaatt cccaaaggat caactggaaa aattgacaga aaagtgttaa gacaaatgtt	1620
tgaaaaacac accaatggg	1639

<210> 5

<211> 1639

<212> DNA

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 5

ggatccaatg gaagataaaa atatttata tggacctgaa ccattttatc ccttggctga	60
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catagcattg acaaattgtc atacaaaagc ccctgttttata tatgaagagt tgtaaaatt	180
gtcgtgtcgat ttagcggaaa gttttaaaaa gtatggatta aaacaaaacg acacaatagc	240
ggtgtgttagc gaaaatggtt tgcaattttt ctttcctata attgcattcat tgtatcttgg	300
aataattgtca gcacctgttata gtgataaata cattgaacgt gaattaatac acagtcttgg	360
tattgtaaaa ccacgcataa tttttgtctt caagaataact tttcaaaaag tactgaatgt	420
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aggttatcaa tgcctcaaca actttatttc tcaaaaattcc gatattaatc ttgcacgtaaa	540
aaaattttaaa ccatattctt ttaatcgaga cgatcagggtt gcgttggtaa tgtttcttc	600
tggtacaact ggtgttcga agggagtcat gctaactcac aagaatattt ttgcacgatt	660
ttctcatgca aaagatccta cttttggtaa cgcaattaat ccaacgacag caattttaac	720
ggtaataacct ttccaccatg gttttggat gatgaccaca tttaggataact ttacttgtgg	780
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<210> 6

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<212> DNA

<213> Artificial Sequence

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<223> /note = "mutant luciferase"

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<211> 1639
<212> DNA
<213> Artificial Sequence

<220>
<221> unsure
<222> (1067) ... (1072)
<223> /note = "unknown nucleotides"

<223> /note = "mutant luciferase"

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<210> 8
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<212> DNA
<213> Artificial Sequence

<220>
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<223> /note = "unknown nucleotides"

<223> note = " Mutant luciferase

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<211> 1639
<212> DNA
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<220>
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<223> /note = "unknown nucleotides"

<223> note = "Mutant luciferase

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<210> 10
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<223> /note = "unknown nucleotides"

<223> note = " Mutant luciferase

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<210> 11
<211> 1639
<212> DNA
<213> Artificial Sequence

<220>
<223> /note = "mutant luciferase"

<400> 11

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<212> DNA
<213> Artificial Sequence

<220>
 <223> /note = "mutant luciferase"

<400> 12

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<210> 13
 <211> 1626
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> /note = "mutant luciferase"

<400> 13

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ccaggaattt gctgtggatt tggtttgcac	
aatctactt cagctaatat acacagtctt	
agggatgaat ttaaatcagg atcacttgc	
agagttactc cttaatggc agctaaaata	
gcagataggg aaactggtaa agcattggg	
ccaaatcaag ttggtaattt atgcattaaa	
ggtccatgg tataatggc	
ttacgtgaac aatgttagaaat	
ctaccaaaga agctattgtat gatgtggtt	
ggcttcactc tggagacttt	
ggatactatg atgaggatga	
gttacaaggaa attgatttttttata	
ataaagggtt ctcaggtagc acctgcagaa	
ctagaagaga ttttattgaa aaatccatgt	
atcagagatg ttgctgttgt tggattttctt	
gatctagaag ctggagaact gccatctgc	
tttgggttta aacagccccgg	
aaaggagatt acagctaaag aagtgtacga	
ttatcttgc	
gagagggtct cccataaaaaa gtatgtcgt	
ggaggggttc gattcgttga tagcatacca	
aggatgtt caggtaaaat tacaagaaag	
gaacttctga agcagttgt ggagaaggcg	
ggaggtt	

<210> 14

<211> 544

<212> PRT

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 14

Met Ala Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu

1

5

10

15

Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Tyr Ala Leu Ser Arg Tyr

20

25

30

Ala Asp Ile Ser Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu
 35 40 45
 Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu
 50 55 60
 Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys
 65 70 75 80
 Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Ile Ile Ala Ser Leu Tyr
 85 90 95
 Leu Gly Ile Ile Ala Ala Pro Val Ser Asp Lys Tyr Ile Glu Arg Glu
 100 105 110
 Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser
 115 120 125
 Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Tyr
 130 135 140
 Val Glu Thr Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr
 145 150 155 160
 Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ile Asn Leu Asp
 165 170 175
 Val Lys Lys Phe Lys Pro Tyr Ser Phe Asn Arg Asp Asp Gln Val Ala
 180 185 190
 Leu Val Met Phe Ser Ser Gly Thr Thr Gly Val Ser Lys Gly Val Met
 195 200 205
 Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser Leu Ala Lys Asp Pro
 210 215 220
 Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile
 225 230 235 240
 Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr
 245 250 255
 Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe
 260 265 270
 Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro
 275 280 285
 Thr Leu Met Ala Phe Leu Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp
 290 295 300
 Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys
 305 310 315 320
 Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg
 325 330 335
 Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro
 340 345 350
 Asn Asn Asp Val Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His
 355 360 365

Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn
 370 375 380
 Glu Pro Gly Glu Leu Tyr Phe Lys Gly Asp Met Ile Met Lys Gly Tyr
 385 390 395 400
 Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asn Lys Asp Gly Trp
 405 410 415
 Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr
 420 425 430
 Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val
 435 440 445
 Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val
 450 455 460
 Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro
 465 470 475 480
 Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile
 485 490 495
 Val Gln Asn Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg
 500 505 510
 Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys
 515 520 525
 Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly
 530 535 540

<210> 15
 <211> 544
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> /note = "mutant luciferase"

<400> 15
 Met Glu Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu
 1 5 10 15
 Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Tyr Ala Leu Ser Arg Tyr
 20 25 30
 Ala Asp Ile Ser Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu
 35 40 45
 Asn Val Leu Tyr Glu Glu Leu Leu Lys Leu Ser Cys Arg Leu Ala Glu
 50 55 60
 Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys
 65 70 75 80

Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Ile Ile Ala Ser Leu Tyr
 85 90 95
 Leu Gly Ile Ile Ala Ala Pro Val Ser Asp Lys Tyr Ile Glu Arg Glu
 100 105 110
 Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser
 115 120 125
 Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Tyr
 130 135 140
 Val Glu Thr Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr
 145 150 155 160
 Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ile Asn Leu Asp
 165 170 175
 Val Lys Lys Phe Lys Pro Tyr Ser Phe Asn Arg Asp Asp Gln Val Ala
 180 185 190
 Leu Val Met Phe Ser Ser Gly Thr Thr Gly Val Ser Lys Gly Val Met
 195 200 205
 Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser His Ala Lys Asp Pro
 210 215 220
 Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile
 225 230 235 240
 Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr
 245 250 255
 Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe
 260 265 270
 Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro
 275 280 285
 Thr Leu Met Ala Phe Phe Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp
 290 295 300
 Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys
 305 310 315 320
 Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg
 325 330 335
 Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro
 340 345 350
 Asn Asn Asp Val Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His
 355 360 365
 Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn
 370 375 380
 Glu Thr Gly Glu Leu Tyr Phe Lys Gly Asp Met Ile Met Lys Gly Tyr
 385 390 395 400
 Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asn Lys Asp Gly Trp
 405 410 415

Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr
 420 425 430
 Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val
 435 440 445
 Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val
 450 455 460
 Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro
 465 470 475 480
 Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile
 485 490 495
 Val Gln Asn Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg
 500 505 510
 Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys
 515 520 525
 Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly
 530 535 540

<210> 16

<211> 544

<212> PRT

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 16

Met Glu Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu
 1 5 10 15
 Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Tyr Ala Leu Ser Arg Tyr
 20 25 30
 Ala Asp Ile Ser Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu
 35 40 45
 Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu
 50 55 60
 Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys
 65 70 75 80
 Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Ile Ile Ala Ser Leu Tyr
 85 90 95
 Leu Gly Ile Ile Ala Ala Pro Val Ser Asp Lys Tyr Ile Glu Arg Glu
 100 105 110
 Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser
 115 120 125

Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Tyr
130 135 140
Val Glu Thr Ile Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr
145 150 155 160
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ile Asn Leu Asp
165 170 175
Val Lys Lys Phe Lys Pro Tyr Ser Phe Asn Arg Asp Asp Gln Val Ala
180 185 190
Leu Val Met Phe Ser Ser Gly Thr Thr Gly Val Ser Lys Gly Val Met
195 200 205
Leu Thr His Lys Asn Ile Val Val Arg Phe Ser Leu Ala Lys Asp Pro
210 215 220
Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile
225 230 235 240
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr
245 250 255
Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe
260 265 270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro
275 280 285
Thr Leu Met Ala Phe Phe Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp
290 295 300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys
305 310 315 320
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg
325 330 335
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro
340 345 350
Asn Asn Asp Val Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His
355 360 365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn
370 375 380
Glu Thr Gly Glu Leu Tyr Phe Lys Gly Asp Met Ile Met Lys Gly Tyr
385 390 395 400
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Thr Lys Asp Gly Trp
405 410 415
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr
420 425 430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val
435 440 445
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val
450 455 460

Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro
 465 470 475 480
 Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile
 485 490 495
 Val Gln Asn Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg
 500 505 510
 Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys
 515 520 525
 Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly
 530 535 540

<210> 17

<211> 544

<212> PRT

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 17

Met Glu Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu
 1 5 10 15
 Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Tyr Ala Leu Ser Arg Tyr
 20 25 30
 Ala Asp Ile Ser Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu
 35 40 45
 Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu
 50 55 60
 Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys
 65 70 75 80
 Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Ile Ile Ala Ser Leu Tyr
 85 90 95
 Leu Gly Ile Ile Ala Ala Pro Val Ser Asp Lys Tyr Ile Glu Arg Glu
 100 105 110
 Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser
 115 120 125
 Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Tyr
 130 135 140
 Val Glu Thr Ile Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr
 145 150 155 160
 Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ile Asn Leu Asp
 165 170 175

Val Lys Lys Phe Lys Pro Tyr Ser Phe Asn Arg Asp Asp Gln Val Ala
180 185 190
Leu Val Met Phe Ser Ser Gly Thr Thr Gly Val Ser Lys Gly Val Met
195 200 205
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser Ile Ala Lys Asp Pro
210 215 220
Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile
225 230 235 240
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr
245 250 255
Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe
260 265 270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro
275 280 285
Thr Leu Met Ala Phe Leu Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp
290 295 300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys
305 310 315 320
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg
325 330 335
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro
340 345 350
Asn Asn Asp Val Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His
355 360 365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn
370 375 380
Glu Thr Gly Glu Leu Tyr Phe Lys Gly Asp Met Ile Met Lys Gly Tyr
385 390 395 400
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asn Lys Asp Gly Trp
405 410 415
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr
420 425 430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val
435 440 445
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val
450 455 460
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro
465 470 475 480
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile
485 490 495
Val Gln Asn Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg
500 505 510

Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys
515 520 525
Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly
530 535 540

<210> 18
<211> 544
<212> PRT
<213> Artificial Sequence

<220>
<223> /note = "mutant luciferase"

<400> 18

Met Glu Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu
1 5 10 15
Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Asp Ala Leu Ser Arg Tyr
20 25 30
Ala Asp Ile Ser Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu
35 40 45
Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu
50 55 60
Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys
65 70 75 80
Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Ile Ile Ala Ser Leu Tyr
85 90 95
Leu Gly Ile Ile Ala Ala Pro Val Ser Asp Lys Tyr Ile Glu Arg Glu
100 105 110
Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser
115 120 125
Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Tyr
130 135 140
Val Glu Thr Ile Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr
145 150 155 160
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ile Asn Leu Asp
165 170 175
Val Lys Lys Phe Lys Pro Tyr Ser Phe Asn Arg Asp Asp Gln Val Ala
180 185 190
Leu Val Met Phe Ser Ser Gly Thr Thr Gly Val Ser Lys Gly Val Met
195 200 205
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser His Ala Lys Asp Pro
210 215 220

Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile
225 230 235 240
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr
245 250 255
Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe
260 265 270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro
275 280 285
Thr Leu Met Ala Phe Phe Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp
290 295 300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys
305 310 315 320
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg
325 330 335
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro
340 345 350
Asn Asn Asp Val Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His
355 360 365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn
370 375 380
Glu Thr Gly Glu Leu Tyr Phe Lys Gly Asp Met Ile Met Lys Gly Tyr
385 390 395 400
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asn Lys Asp Gly Trp
405 410 415
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr
420 425 430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val
435 440 445
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val
450 455 460
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro
465 470 475 480
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile
485 490 495
Val Gln Asn Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg
500 505 510
Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys
515 520 525
Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly
530 535 540

<211> 544
<212> PRT
<213> Artificial Sequence

<220>
<223> /note = "mutant luciferase"

<400> 19

Met	Ala	Asp	Lys	Asn	Ile	Leu	Tyr	Gly	Pro	Glu	Pro	Phe	Tyr	Pro	Leu
1					5				10					15	
Ala	Asp	Gly	Thr	Ala	Gly	Glu	Gln	Met	Phe	Asp	Ala	Leu	Ser	Arg	Tyr
			20					25					30		
Ala	Asp	Ile	Ser	Gly	Cys	Ile	Ala	Leu	Thr	Asn	Ala	His	Thr	Lys	Glu
			35				40					45			
Asn	Val	Leu	Tyr	Glu	Glu	Phe	Leu	Lys	Leu	Ser	Cys	Arg	Leu	Ala	Glu
			50			55			60						
Ser	Phe	Lys	Lys	Tyr	Gly	Leu	Lys	Gln	Asn	Asp	Thr	Ile	Ala	Val	Cys
			65			70			75				80		
Ser	Glu	Asn	Gly	Leu	Gln	Phe	Phe	Leu	Pro	Val	Ile	Ala	Ser	Leu	Tyr
				85				90					95		
Leu	Gly	Ile	Ile	Ala	Ala	Pro	Val	Ser	Asp	Lys	Tyr	Ile	Glu	Arg	Glu
			100				105					110			
Leu	Ile	His	Ser	Leu	Gly	Ile	Val	Lys	Pro	Arg	Ile	Ile	Phe	Cys	Ser
			115				120					125			
Lys	Asn	Thr	Phe	Gln	Lys	Val	Leu	Asn	Val	Lys	Ser	Lys	Leu	Lys	Ser
			130			135				140					
Val	Glu	Thr	Ile	Ile	Ile	Leu	Asp	Leu	Asn	Glu	Asp	Leu	Gly	Gly	Tyr
			145			150				155			160		
Gln	Cys	Leu	Asn	Asn	Phe	Ile	Ser	Gln	Asn	Ser	Asp	Ser	Asn	Leu	Asp
				165				170				175			
Val	Lys	Lys	Phe	Lys	Pro	Tyr	Ser	Phe	Asn	Arg	Asp	Asp	Gln	Val	Ala
			180				185				190				
Leu	Val	Met	Phe	Ser	Ser	Gly	Thr	Thr	Gly	Val	Pro	Lys	Gly	Val	Met
			195				200				205				
Leu	Thr	His	Lys	Asn	Ile	Val	Ala	Arg	Phe	Ser	Leu	Ala	Lys	Asp	Pro
			210			215				220					
Thr	Phe	Gly	Asn	Ala	Ile	Asn	Pro	Thr	Thr	Ala	Ile	Leu	Thr	Val	Ile
			225			230				235			240		
Pro	Phe	His	His	Gly	Phe	Gly	Met	Met	Thr	Thr	Leu	Gly	Tyr	Phe	Thr
				245				250				255			
Cys	Gly	Phe	Arg	Val	Val	Leu	Met	His	Thr	Phe	Glu	Glu	Lys	Leu	Phe
				260				265			270				

Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro
 275 280 285
 Thr Leu Met Ala Phe Leu Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp
 290 295 300
 Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys
 305 310 315 320
 Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg
 325 330 335
 Gln Gly Tyr Gly Leu Thr Glu Thr Ser Ala Val Leu Ile Thr Pro
 340 345 350
 Lys Gly Asp Ala Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His
 355 360 365
 Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn
 370 375 380
 Glu Pro Gly Glu Leu Tyr Phe Lys Gly Ala Met Ile Met Lys Gly Tyr
 385 390 395 400
 Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asp Asn Asp Gly Trp
 405 410 415
 Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr
 420 425 430
 Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val
 435 440 445
 Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val
 450 455 460
 Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro
 465 470 475 480
 Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile
 485 490 495
 Val Gln Asp Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg
 500 505 510
 Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys
 515 520 525
 Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly
 530 535 540

<210> 20

<211> 544

<212> PRT

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<221> unsure
<222> (354)...(355)
<223> /note = "unknown amino acids"

<400> 20

Met Ala Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu
1 5 10 15
Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Tyr Ala Leu Ser Arg Tyr
20 25 30
Ala Asp Ile Ser Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu
35 40 45
Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu
50 55 60
Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys
65 70 75 80
Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Val Ile Ala Ser Leu Tyr
85 90 95
Leu Gly Ile Ile Ala Ala Pro Val Ser Asp Lys Tyr Ile Glu Arg Glu
100 105 110
Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser
115 120 125
Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Tyr
130 135 140
Val Glu Thr Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr
145 150 155 160
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ile Asn Leu Asp
165 170 175
Val Lys Lys Phe Lys Pro Tyr Ser Phe Asn Arg Asp Asp Gln Val Ala
180 185 190
Leu Val Met Phe Ser Ser Gly Thr Thr Gly Val Pro Lys Gly Val Met
195 200 205
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser Leu Ala Lys Asp Pro
210 215 220
Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile
225 230 235 240
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr
245 250 255
Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe
260 265 270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro
275 280 285
Thr Leu Met Ala Phe Leu Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp

290	295	300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys		
305	310	315
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg		
325	330	335
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro		
340	345	350
Lys Xaa Xaa Val Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His		
355	360	365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn		
370	375	380
Glu Pro Gly Glu Leu Tyr Phe Lys Gly Asp Met Ile Met Lys Gly Tyr		
385	390	395
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asp Lys Asp Gly Trp		
405	410	415
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr		
420	425	430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val		
435	440	445
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val		
450	455	460
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro		
465	470	475
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile		
485	490	495
Val Gln Asn Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg		
500	505	510
Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys		
515	520	525
Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly		
530	535	540

<210> 21

<211> 544

<212> PRT

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<221> unsure

<222> (354)...(355)

<223> /note = "unknown amino acids"

<400> 21

Met Ala Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu
1 5 10 15
Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Asp Ala Leu Ser Arg Tyr
20 25 30
Ala Asp Ile Pro Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu
35 40 45
Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu
50 55 60
Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys
65 70 75 80
Ser Glu Asn Gly Leu Gln Tyr Phe Leu Pro Val Ile Ala Ser Leu Tyr
85 90 95
Leu Gly Ile Ile Ala Ala Pro Val Ser Asp Lys Tyr Ile Glu Arg Glu
100 105 110
Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser
115 120 125
Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Tyr
130 135 140
Val Glu Thr Ile Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr
145 150 155 160
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ile Asn Leu Asp
165 170 175
Val Lys Lys Phe Lys Pro Asn Ser Phe Asn Arg Asp Asp Gln Val Ala
180 185 190
Leu Val Met Phe Ser Ser Gly Thr Thr Gly Val Pro Lys Gly Val Met
195 200 205
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser Ile Ala Lys Asp Pro
210 215 220
Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile
225 230 235 240
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr
245 250 255
Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe
260 265 270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro
275 280 285
Thr Leu Met Ala Phe Leu Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp
290 295 300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys

305	310	315	320
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg			
325	330	335	
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro			
340	345	350	
Lys Xaa Xaa Ala Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His			
355	360	365	
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn			
370	375	380	
Glu Pro Gly Glu Leu Tyr Phe Lys Gly Ala Met Ile Met Lys Gly Tyr			
385	390	395	400
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asp Lys Asp Gly Trp			
405	410	415	
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr			
420	425	430	
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val			
435	440	445	
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val			
450	455	460	
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro			
465	470	475	480
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile			
485	490	495	
Val Gln Asn Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg			
500	505	510	
Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys			
515	520	525	
Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly			
530	535	540	

<210> 22
<211> 544
<212> PRT
<213> Artificial Sequence

<220>
<223> /note = "mutant luciferase"

<221> unsure
<222> (354)...(355)
<223> /note = "unknown amino acids"

<400> 22

Met Ala Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu
1 5 10 15
Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Asp Ala Leu Ser Arg Tyr
20 25 30
Ala Asp Ile Pro Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu
35 40 45
Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu
50 55 60
Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys
65 70 75 80
Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Val Ile Ala Ser Leu Tyr
85 90 95
Leu Gly Ile Ile Ala Ala Pro Val Ser Asp Lys Tyr Val Glu Arg Glu
100 105 110
Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser
115 120 125
Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Tyr
130 135 140
Val Glu Thr Ile Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr
145 150 155 160
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ser Asn Leu Asp
165 170 175
Val Lys Lys Phe Lys Pro Asn Ser Phe Asn Arg Asp Asp Gln Val Ala
180 185 190
Leu Val Met Phe Ser Ser Gly Thr Thr Gly Val Pro Lys Gly Val Met
195 200 205
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser Leu Ala Lys Asp Pro
210 215 220
Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile
225 230 235 240
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr
245 250 255
Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe
260 265 270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro
275 280 285
Thr Leu Met Ala Phe Leu Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp
290 295 300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys
305 310 315 320
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg

325	330	335
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro		
340	345	350
Lys Xaa Xaa Ala Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His		
355	360	365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn		
370	375	380
Glu Thr Gly Glu Leu Tyr Phe Lys Gly Ala Met Ile Met Lys Gly Tyr		
385	390	395
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asp Lys Asp Gly Trp		
405	410	415
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr		
420	425	430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val		
435	440	445
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val		
450	455	460
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro		
465	470	475
480		
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile		
485	490	495
Val Gln Asn Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg		
500	505	510
Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys		
515	520	525
Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly		
530	535	540

<210> 23

<211> 544

<212> PRT

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<221> unsure

<222> (354) . . . (355)

<223> /note = "unknown amino acids"

<400> 23

Met Ala Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu

1	5	10	15
Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Asp Ala Leu Ser Arg Tyr			
20	25	30	
Ala Asp Ile Pro Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu			
35	40	45	
Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu			
50	55	60	
Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys			
65	70	75	80
Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Val Ile Ala Ser Leu Tyr			
85	90	95	
Leu Gly Ile Ile Val Ala Pro Val Asn Asp Lys Tyr Ile Glu Arg Glu			
100	105	110	
Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser			
115	120	125	
Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Ser			
130	135	140	
Val Glu Thr Ile Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr			
145	150	155	160
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ile Asn Leu Asp			
165	170	175	
Val Lys Lys Phe Lys Pro Tyr Ser Phe Asn Arg Asp Asp Gln Val Ala			
180	185	190	
Leu Ile Met Phe Ser Ser Gly Thr Thr Gly Leu Pro Lys Gly Val Met			
195	200	205	
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser Leu Ala Lys Asp Pro			
210	215	220	
Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile			
225	230	235	240
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr			
245	250	255	
Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe			
260	265	270	
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro			
275	280	285	
Thr Leu Met Ala Phe Leu Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp			
290	295	300	
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys			
305	310	315	320
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg			
325	330	335	
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro			

340	345	350
Lys Xaa Xaa Ala Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His		
355	360	365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn		
370	375	380
Glu Pro Gly Glu Leu Tyr Phe Lys Gly Pro Met Ile Met Lys Gly Tyr		
385	390	395
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asp Asn Asp Gly Trp		
405	410	415
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr		
420	425	430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val		
435	440	445
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val		
450	455	460
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro		
465	470	475
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile		
485	490	495
Val Gln Asp Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg		
500	505	510
Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys		
515	520	525
Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly		
530	535	540

<210> 24

<211> 544

<212> PRT

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 24

Met Ala Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu			
1	5	10	15
Glu Asp Gly Thr Ala Gly Glu Gln Met Phe Asp Ala Leu Ser Arg Tyr			
20	25	30	
Ala Asp Ile Pro Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu			
35	40	45	
Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu			

50	55	60
Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys		
65	70	75
Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Val Ile Ala Ser Leu Tyr		
85	90	95
Leu Gly Ile Ile Val Ala Pro Val Asn Asp Lys Tyr Ile Glu Arg Glu		
100	105	110
Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Val Phe Cys Ser		
115	120	125
Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Ser		
130	135	140
Ile Glu Thr Ile Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr		
145	150	155
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ser Asn Leu Asp		
165	170	175
Val Lys Lys Phe Lys Pro Tyr Ser Phe Asn Arg Asp Asp Gln Val Ala		
180	185	190
Leu Ile Met Phe Ser Ser Gly Thr Thr Gly Leu Pro Lys Gly Val Met		
195	200	205
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser Leu Ala Lys Asp Pro		
210	215	220
Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile		
225	230	235
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr		
245	250	255
Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe		
260	265	270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro		
275	280	285
Thr Leu Met Ala Phe Leu Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp		
290	295	300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys		
305	310	315
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg		
325	330	335
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro		
340	345	350
Lys Gly Asp Ala Lys Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His		
355	360	365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn		
370	375	380
Glu Pro Gly Glu Leu Tyr Phe Lys Gly Pro Met Ile Met Lys Gly Tyr		

385	390	395	400
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asp Asn Asp Gly Trp			
405	410	415	
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr			
420	425	430	
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val			
435	440	445	
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val			
450	455	460	
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro			
465	470	475	480
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile			
485	490	495	
Val Gln Asp Tyr Val Ala Ser Gln Val Ser Thr Ala Lys Trp Leu Arg			
500	505	510	
Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys			
515	520	525	
Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Thr Asn Gly			
530	535	540	

<210> 25

<211> 545

<212> PRT

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 25

1	5	10	15
Met Glu Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu			
20	25	30	
Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Tyr Ala Leu Ser Arg Tyr			
35	40	45	
Ala Asp Ile Ser Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu			
50	55	60	
Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu			
65	70	75	80
Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys			
85	90	95	
Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Leu Ile Ala Ser Leu Tyr			
Leu Gly Ile Ile Ala Ala Pro Val Ser Asp Lys Tyr Ile Glu Arg Glu			

100	105	110
Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser		
115	120	125
Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Tyr		
130	135	140
Val Glu Thr Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr		
145	150	155
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ile Asn Leu Asp		
165	170	175
Val Lys Lys Phe Lys Pro Asn Ser Phe Asn Arg Asp Asp Gln Val Ala		
180	185	190
Leu Val Met Phe Ser Ser Gly Thr Thr Gly Val Ser Lys Gly Val Met		
195	200	205
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser His Cys Lys Asp Pro		
210	215	220
Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile		
225	230	235
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr		
245	250	255
Cys Gly Phe Arg Val Ala Leu Met His Thr Phe Glu Glu Lys Leu Phe		
260	265	270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro		
275	280	285
Thr Leu Met Ala Phe Phe Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp		
290	295	300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys		
305	310	315
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg		
325	330	335
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro		
340	345	350
Asp Thr Asp Val Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His		
355	360	365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn		
370	375	380
Glu Thr Gly Glu Leu Tyr Phe Lys Gly Asp Met Ile Met Lys Gly Tyr		
385	390	395
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asn Lys Asp Gly Trp		
405	410	415
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr		
420	425	430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val		

435 440 445
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val
450 455 460
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro
465 470 475 480
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile
485 490 495
Val Gln Asn Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg
500 505 510
Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys
515 520 525
Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Lys Ser Lys
530 535 540
Leu
545

<210> 26
<211> 542
<212> PRT
<213> Artificial Sequence

<220>
<223> note = "mutant luciferase"

<400> 26
Met Met Lys Arg Glu Lys Asn Val Ile Tyr Gly Pro Glu Pro Leu His
1 5 10 15
Pro Leu Glu Asp Leu Thr Ala Gly Glu Met Leu Phe Arg Ala Leu Arg
20 25 30
Lys His Ser His Leu Pro Gln Ala Leu Val Asp Val Val Gly Asp Glu
35 40 45
Ser Leu Ser Tyr Lys Glu Phe Phe Glu Ala Thr Val Leu Leu Ala Gln
50 55 60
Ser Leu His Asn Cys Gly Tyr Lys Met Asn Asp Val Val Ser Ile Cys
65 70 75 80
Ala Glu Asn Asn Thr Arg Phe Phe Ile Pro Val Ile Ala Ala Trp Tyr
85 90 95
Ile Gly Met Ile Val Ala Pro Val Asn Glu Ser Tyr Ile Pro Asp Glu
100 105 110
Leu Cys Lys Val Met Gly Ile Ser Lys Pro Gln Ile Val Phe Thr Thr
115 120 125
Lys Asn Ile Leu Asn Lys Val Leu Glu Val Gln Ser Arg Thr Asn Phe

130 135 140
Ile Lys Arg Ile Ile Ile Leu Asp Thr Val Glu Asn Ile His Gly Cys
145 150 155 160
Glu Ser Leu Pro Asn Gly Ile Ser Arg Tyr Ser Asp Gly Asn Ile Ala
165 170 175
Asn Phe Lys Pro Leu His Phe Asp Pro Val Glu Gln Val Ala Ala Ile
180 185 190
Leu Cys Ser Ser Gly Thr Thr Gly Leu Pro Lys Gly Val Met Gln Thr
195 200 205
His Gln Asn Ile Cys Val Arg Leu Ile His Ala Leu Asp Pro Arg Ala
210 215 220
Gly Thr Gln Leu Ile Pro Gly Val Thr Val Leu Val Tyr Leu Pro Phe
225 230 235 240
Phe His Ala Phe Gly Phe Ser Ile Thr Leu Gly Tyr Phe Met Val Gly
245 250 255
Leu Arg Val Ile Met Phe Arg Arg Phe Asp Gln Glu Ala Phe Leu Lys
260 265 270
Ala Ile Gln Asp Tyr Glu Val Arg Ser Val Ile Asn Val Pro Ser Val
275 280 285
Ile Leu Phe Leu Ser Lys Ser Pro Leu Val Asp Lys Tyr Asp Leu Ser
290 295 300
Ser Leu Arg Glu Leu Cys Cys Gly Ala Ala Pro Leu Ala Lys Glu Val
305 310 315 320
Ala Glu Val Ala Ala Lys Arg Leu Asn Leu Pro Gly Ile Arg Cys Gly
325 330 335
Phe Gly Leu Thr Glu Ser Thr Ser Ala Asn Ile His Ser Leu Arg Asp
340 345 350
Glu Phe Lys Ser Gly Ser Leu Gly Arg Val Thr Pro Leu Met Ala Ala
355 360 365
Lys Ile Ala Asp Arg Glu Thr Gly Lys Ala Leu Gly Pro Asn Gln Val
370 375 380
Gly Glu Leu Cys Ile Lys Gly Pro Met Val Ser Lys Gly Tyr Val Asn
385 390 395 400
Asn Val Glu Ala Thr Lys Glu Ala Ile Asp Asp Asp Gly Trp Leu His
405 410 415
Ser Gly Asp Phe Gly Tyr Tyr Asp Glu Asp Glu His Phe Tyr Val Val
420 425 430
Asp Arg Tyr Lys Glu Leu Ile Lys Tyr Lys Gly Ser Gln Val Ala Pro
435 440 445
Ala Glu Leu Glu Glu Ile Leu Leu Lys Asn Pro Cys Ile Arg Asp Val
450 455 460
Ala Val Val Gly Ile Pro Asp Leu Glu Ala Gly Glu Leu Pro Ser Ala

465	470	475	480
Phe Val Val Lys Gln Pro Gly Lys Glu Ile Thr Ala Lys Glu Val Tyr			
485	490	495	
Asp Tyr Leu Ala Glu Arg Val Ser His Thr Lys Tyr Leu Arg Gly Gly			
500	505	510	
Val Arg Phe Val Asp Ser Ile Pro Arg Asn Val Thr Gly Lys Ile Thr			
515	520	525	
Arg Lys Glu Leu Leu Lys Gln Leu Leu Glu Lys Ala Gly Gly			
530	535	540	

<210> 27
 <211> 548
 <212> PRT
 <213> Luciola cruciata

<400> 27			
Met Glu Asn Met Glu Asn Asp Glu Asn Ile Val Val Gly Pro Lys Pro			
1	5	10	15
Phe Tyr Pro Ile Glu Glu Gly Ser Ala Gly Thr Gln Leu Arg Lys Tyr			
20	25	30	
Met Glu Arg Tyr Ala Lys Leu Gly Ala Ile Ala Phe Thr Asn Ala Val			
35	40	45	
Thr Gly Val Asp Tyr Ser Tyr Ala Glu Tyr Leu Glu Lys Ser Cys Cys			
50	55	60	
Leu Gly Lys Ala Leu Gln Asn Tyr Gly Leu Val Val Asp Gly Arg Ile			
65	70	75	80
Ala Leu Cys Ser Glu Asn Cys Glu Glu Phe Phe Ile Pro Val Ile Ala			
85	90	95	
Gly Leu Phe Ile Gly Val Gly Val Ala Pro Thr Asn Glu Ile Tyr Thr			
100	105	110	
Leu Arg Glu Leu Val His Ser Leu Gly Ile Ser Lys Pro Thr Ile Val			
115	120	125	
Phe Ser Ser Lys Lys Gly Leu Asp Lys Val Ile Thr Val Gln Lys Thr			
130	135	140	
Val Thr Thr Ile Lys Thr Ile Val Ile Leu Asp Ser Lys Val Asp Tyr			
145	150	155	160
Arg Gly Tyr Gln Cys Leu Asp Thr Phe Ile Lys Arg Asn Thr Pro Pro			
165	170	175	
Gly Phe Gln Ala Ser Ser Phe Lys Thr Val Glu Val Asp Arg Lys Glu			
180	185	190	
Gln Val Ala Leu Ile Met Asn Ser Ser Gly Ser Thr Gly Leu Pro Lys			
195	200	205	

Gly Val Gln Leu Thr His Glu Asn Thr Val Thr Arg Phe Ser His Ala
210 215 220
Arg Asp Pro Ile Tyr Gly Asn Gln Val Ser Pro Gly Thr Ala Val Leu
225 230 235 240
Thr Val Val Pro Phe His His Gly Phe Gly Met Phe Thr Thr Leu Gly
245 250 255
Tyr Leu Ile Cys Gly Phe Arg Val Val Met Leu Thr Lys Phe Asp Glu
260 265 270
Glu Thr Phe Leu Lys Thr Leu Gln Asp Tyr Lys Cys Thr Ser Val Ile
275 280 285
Leu Val Pro Thr Leu Phe Ala Ile Leu Asn Lys Ser Glu Leu Leu Asn
290 295 300
Lys Tyr Asp Leu Ser Asn Leu Val Glu Ile Ala Ser Gly Gly Ala Pro
305 310 315 320
Leu Ser Lys Glu Val Gly Glu Ala Val Ala Arg Arg Phe Asn Leu Pro
325 330 335
Gly Val Arg Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Ile Ile
340 345 350
Ile Thr Pro Glu Gly Asp Asp Lys Pro Gly Ala Ser Gly Lys Val Val
355 360 365
Pro Leu Phe Lys Ala Lys Val Ile Asp Leu Asp Thr Lys Lys Ser Leu
370 375 380
Gly Pro Asn Arg Arg Gly Glu Val Cys Val Lys Gly Pro Met Leu Met
385 390 395 400
Lys Gly Tyr Val Asn Asn Pro Glu Ala Thr Lys Glu Leu Ile Asp Glu
405 410 415
Glu Gly Trp Leu His Thr Gly Asp Ile Gly Tyr Tyr Asp Glu Glu Lys
420 425 430
His Phe Phe Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly
435 440 445
Tyr Gln Val Pro Pro Ala Glu Leu Glu Ser Val Leu Leu Gln His Pro
450 455 460
Ser Ile Phe Asp Ala Gly Val Ala Gly Val Pro Asp Pro Val Ala Gly
465 470 475 480
Glu Leu Pro Gly Ala Val Val Val Leu Glu Ser Gly Lys Asn Met Thr
485 490 495
Glu Lys Glu Val Met Asp Tyr Val Ala Ser Gln Val Ser Asn Ala Lys
500 505 510
Arg Leu Arg Gly Gly Val Arg Phe Val Asp Glu Val Pro Lys Gly Leu
515 520 525
Thr Gly Lys Ile Asp Gly Arg Ala Ile Arg Glu Ile Leu Lys Lys Pro
530 535 540

Val Ala Lys Met

545

<210> 28

<211> 548

<212> PRT

<213> Luciola lateralis

<400> 28

Met Glu Asn Met Glu Asn Asp Glu Asn Ile Val Tyr Gly Pro Glu Pro

1 5 10 15

Phe Tyr Pro Ile Glu Glu Gly Ser Ala Gly Ala Gln Leu Arg Lys Tyr

20 25 30

Met Asp Arg Tyr Ala Lys Leu Gly Ala Ile Ala Phe Thr Asn Ala Leu

35 40 45

Thr Gly Val Asp Tyr Thr Tyr Ala Glu Tyr Leu Glu Lys Ser Cys Cys

50 55 60

Leu Gly Glu Ala Leu Lys Asn Tyr Gly Leu Val Val Asp Gly Arg Ile

65 70 75 80

Ala Leu Cys Ser Glu Asn Cys Glu Glu Phe Phe Ile Pro Val Leu Ala

85 90 95

Gly Leu Phe Ile Gly Val Gly Val Ala Pro Thr Asn Glu Ile Tyr Thr

100 105 110

Leu Arg Glu Leu Val His Ser Leu Gly Ile Ser Lys Pro Thr Ile Val

115 120 125

Phe Ser Ser Lys Lys Gly Leu Asp Lys Val Ile Thr Val Gln Lys Thr

130 135 140

Val Ala Thr Ile Lys Thr Ile Val Ile Leu Asp Ser Lys Val Asp Tyr

145 150 155 160

Arg Gly Tyr Gln Ser Met Asp Asn Phe Ile Lys Lys Asn Thr Pro Gln

165 170 175

Gly Phe Lys Gly Ser Ser Phe Lys Thr Val Glu Val Asn Arg Lys Glu

180 185 190

Gln Val Ala Leu Ile Met Asn Ser Ser Gly Ser Thr Gly Leu Pro Lys

195 200 205

Gly Val Gln Leu Thr His Glu Asn Ala Val Thr Arg Phe Ser His Ala

210 215 220

Arg Asp Pro Ile Tyr Gly Asn Gln Val Ser Pro Gly Thr Ala Ile Leu

225 230 235 240

Thr Val Val Pro Phe His His Gly Phe Gly Met Phe Thr Thr Leu Gly

245 250 255

Tyr Leu Thr Cys Gly Phe Arg Ile Val Met Leu Thr Lys Phe Asp Glu

260	265	270
Glu Thr Phe Leu Lys Thr Leu Gln Asp Tyr Lys Cys Ser Ser Val Ile		
275	280	285
Leu Val Pro Thr Leu Phe Ala Ile Leu Asn Arg Ser Glu Leu Leu Asp		
290	295	300
Lys Tyr Asp Leu Ser Asn Leu Val Glu Ile Ala Ser Gly Gly Ala Pro		
305	310	315
Leu Ser Lys Glu Ile Gly Glu Ala Val Ala Arg Arg Phe Asn Leu Pro		
325	330	335
Gly Val Arg Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Ile Ile		
340	345	350
Ile Thr Pro Glu Gly Asp Asp Lys Pro Gly Ala Ser Gly Lys Val Val		
355	360	365
Pro Leu Phe Lys Ala Lys Val Ile Asp Leu Asp Thr Lys Lys Thr Leu		
370	375	380
Gly Pro Asn Arg Arg Gly Glu Val Cys Val Lys Gly Pro Met Leu Met		
385	390	395
Lys Gly Tyr Val Asp Asn Pro Glu Ala Thr Arg Glu Ile Ile Asp Glu		
405	410	415
Glu Gly Trp Leu His Thr Gly Asp Ile Gly Tyr Tyr Asp Glu Glu Lys		
420	425	430
His Phe Phe Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly		
435	440	445
Tyr Gln Val Pro Pro Ala Glu Leu Glu Ser Val Leu Leu Gln His Pro		
450	455	460
Asn Ile Phe Asp Ala Gly Val Ala Gly Val Pro Asp Pro Ile Ala Gly		
465	470	475
Glu Leu Pro Gly Ala Val Val Val Leu Glu Lys Gly Lys Ser Met Thr		
485	490	495
Glu Lys Glu Val Met Asp Tyr Val Ala Ser Gln Val Ser Asn Ala Lys		
500	505	510
Arg Leu Arg Gly Gly Val Arg Phe Val Asp Glu Val Pro Lys Gly Leu		
515	520	525
Thr Gly Lys Ile Asp Gly Lys Ala Ile Arg Glu Ile Leu Lys Lys Pro		
530	535	540
Val Ala Lys Met		
545		

<210> 29

<211> 548

<212> PRT

<213> Luciola mingrellica

<400> 29
 Met Glu Met Glu Lys Glu Glu Asn Val Val Tyr Gly Pro Leu Pro Phe
 1 5 10 15
 Tyr Pro Ile Glu Glu Gly Ser Ala Gly Ile Gln Leu His Lys Tyr Met
 20 25 30
 His Gln Tyr Ala Lys Leu Gly Ala Ile Ala Phe Ser Asn Ala Leu Thr
 35 40 45
 Gly Val Asp Ile Ser Tyr Gln Glu Tyr Phe Asp Ile Thr Cys Arg Leu
 50 55 60
 Ala Glu Ala Met Lys Asn Phe Gly Met Lys Pro Glu Glu His Ile Ala
 65 70 75 80
 Leu Cys Ser Glu Asn Cys Glu Glu Phe Phe Ile Pro Val Leu Ala Gly
 85 90 95
 Leu Tyr Ile Gly Val Ala Val Ala Pro Thr Asn Glu Ile Tyr Thr Leu
 100 105 110
 Arg Glu Leu Asn His Ser Leu Gly Ile Ala Gln Pro Thr Ile Val Phe
 115 120 125
 Ser Ser Arg Lys Gly Leu Pro Lys Val Leu Glu Val Gln Lys Thr Val
 130 135 140
 Thr Cys Ile Lys Lys Ile Val Ile Leu Asp Ser Lys Val Asn Phe Gly
 145 150 155 160
 Gly His Asp Cys Met Glu Thr Phe Ile Lys Lys His Val Glu Leu Gly
 165 170 175
 Phe Gln Pro Ser Ser Phe Val Pro Ile Asp Val Lys Asn Arg Lys Gln
 180 185 190
 His Val Ala Leu Leu Met Asn Ser Ser Gly Ser Thr Gly Leu Pro Lys
 195 200 205
 Gly Val Arg Ile Thr His Glu Gly Ala Val Thr Arg Phe Ser His Ala
 210 215 220
 Lys Asp Pro Ile Tyr Gly Asn Gln Val Ser Pro Gly Thr Ala Ile Leu
 225 230 235 240
 Thr Val Val Pro Phe His His Gly Phe Gly Met Phe Thr Thr Leu Gly
 245 250 255
 Tyr Phe Ala Cys Gly Tyr Arg Val Val Met Leu Thr Lys Phe Asp Glu
 260 265 270
 Glu Leu Phe Leu Arg Thr Leu Gln Asp Tyr Lys Cys Thr Ser Val Ile
 275 280 285
 Leu Val Pro Thr Leu Phe Ala Ile Leu Asn Lys Ser Glu Leu Ile Asp
 290 295 300
 Lys Phe Asp Leu Ser Asn Leu Thr Glu Ile Ala Ser Gly Gly Ala Pro
 305 310 315 320
 Leu Ala Lys Glu Val Gly Glu Ala Val Ala Arg Arg Phe Asn Leu Pro

325	330	335	
Gly Val Arg Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Phe Ile			
340	345	350	
Ile Thr Pro Glu Gly Asp Asp Lys Pro Gly Ala Ser Gly Lys Val Val			
355	360	365	
Pro Leu Phe Lys Val Lys Val Ile Asp Leu Asp Thr Lys Lys Thr Leu			
370	375	380	
Gly Val Asn Arg Arg Gly Glu Ile Cys Val Lys Gly Pro Ser Leu Met			
385	390	395	400
Leu Gly Tyr Ser Asn Asn Pro Glu Ala Thr Arg Glu Thr Ile Asp Glu			
405	410	415	
Glu Gly Trp Leu His Thr Gly Asp Ile Gly Tyr Tyr Asp Glu Asp Glu			
420	425	430	
His Phe Phe Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly			
435	440	445	
Tyr Gln Val Pro Pro Ala Glu Leu Glu Ser Val Leu Leu Gln His Pro			
450	455	460	
Asn Ile Phe Asp Ala Gly Val Ala Gly Val Pro Asp Pro Asp Ala Gly			
465	470	475	480
Glu Leu Pro Gly Ala Val Val Val Met Glu Lys Gly Lys Thr Met Thr			
485	490	495	
Glu Lys Glu Ile Val Asp Tyr Val Asn Ser Gln Val Val Asn His Lys			
500	505	510	
Arg Leu Arg Gly Gly Val Arg Phe Val Asp Glu Val Pro Lys Gly Leu			
515	520	525	
Thr Gly Lys Ile Asp Ala Lys Val Ile Arg Glu Ile Leu Lys Lys Pro			
530	535	540	
Gln Ala Lys Met			
545			

<210> 30
 <211> 548
 <212> PRT
 <213> Pyrocoelia miyako

<400> 30			
Met Glu Asp Asp Ser Lys His Ile Met His Gly His Arg His Ser Ile			
1	5	10	15
Leu Trp Glu Asp Gly Thr Ala Gly Glu Gln Leu His Lys Ala Met Lys			
20	25	30	
Arg Tyr Ala Gln Val Pro Gly Thr Ile Ala Phe Thr Asp Ala His Ala			
35	40	45	

Glu Val Asn Ile Thr Tyr Ser Glu Tyr Phe Glu Met Ser Cys Arg Leu
 50 55 60
 Ala Glu Thr Met Lys Arg Tyr Gly Leu Gly Leu Gln His His Ile Ala
 65 70 75 80
 Val Cys Ser Glu Thr Ser Leu Gln Phe Phe Met Pro Val Cys Gly Ala
 85 90 95
 Leu Phe Ile Gly Val Gly Val Ala Pro Thr Asn Asp Ile Tyr Asn Glu
 100 105 110
 Arg Glu Leu Tyr Asn Ser Leu Phe Ile Ser Gln Pro Thr Ile Val Phe
 115 120 125
 Cys Ser Lys Arg Ala Leu Gln Lys Ile Leu Gly Val Gln Lys Lys Leu
 130 135 140
 Pro Val Ile Gln Lys Ile Val Ile Leu Asp Ser Arg Glu Asp Tyr Met
 145 150 155 160
 Gly Lys Gln Ser Met Tyr Ser Phe Ile Glu Ser His Leu Pro Ala Gly
 165 170 175
 Phe Asn Glu Tyr Asp Tyr Ile Pro Asp Ser Phe Asp Arg Glu Thr Ala
 180 185 190
 Thr Ala Leu Ile Met Asn Ser Ser Gly Ser Thr Gly Leu Pro Lys Gly
 195 200 205
 Val Asp Leu Thr His Met Asn Val Cys Val Arg Phe Ser His Cys Arg
 210 215 220
 Asp Pro Val Phe Gly Asn Gln Ile Ile Pro Asp Thr Ala Ile Leu Thr
 225 230 235 240
 Val Ile Pro Phe His His Val Phe Gln Met Phe Thr Thr Leu Gly Tyr
 245 250 255
 Leu Thr Cys Gly Phe Arg Ile Val Leu Met Tyr Arg Phe Glu Glu Glu
 260 265 270
 Leu Phe Leu Arg Ser Leu Gln Asp Tyr Lys Ile Gln Ser Ala Leu Leu
 275 280 285
 Val Pro Thr Leu Phe Ser Phe Ala Lys Ser Thr Leu Val Asp Lys
 290 295 300
 Tyr Asp Leu Ser Asn Leu His Glu Ile Ala Ser Gly Gly Ala Pro Leu
 305 310 315 320
 Ala Lys Glu Val Gly Glu Ala Val Ala Lys Arg Phe Lys Leu Pro Gly
 325 330 335
 Ile Arg Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Ile Ile Ile
 340 345 350
 Thr Pro Glu Gly Asp Asp Lys Pro Gly Ala Cys Gly Lys Val Val Pro
 355 360 365
 Phe Phe Thr Ala Lys Ile Val Asp Leu Asp Thr Gly Lys Thr Leu Gly
 370 375 380

Val Asn Gln Arg Gly Glu Leu Cys Val Lys Gly Pro Met Ile Met Lys
 385 390 395 400
 Gly Tyr Val Asn Asn Pro Glu Ala Thr Asn Ala Leu Ile Asp Lys Asp
 405 410 415
 Gly Trp Leu His Ser Gly Asp Ile Ala Tyr Tyr Asp Lys Asp Gly His
 420 425 430
 Phe Phe Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr
 435 440 445
 Gln Val Pro Pro Ala Glu Leu Glu Ser Ile Leu Leu Gln His Pro Phe
 450 455 460
 Ile Phe Asp Ala Gly Val Ala Gly Ile Pro Asp Pro Asp Ala Gly Glu
 465 470 475 480
 Leu Pro Ala Ala Val Val Val Leu Glu Glu Gly Lys Met Met Thr Glu
 485 490 495
 Gln Glu Val Met Asp Tyr Val Ala Gly Gln Val Thr Ala Ser Lys Arg
 500 505 510
 Leu Arg Gly Gly Val Lys Phe Val Asp Glu Val Pro Lys Gly Leu Thr
 515 520 525
 Gly Lys Ile Asp Ser Arg Lys Ile Arg Glu Ile Leu Thr Met Gly Gln
 530 535 540
 Lys Ser Lys Leu
 545

<210> 31
 <211> 550
 <212> PRT
 <213> Photinus pyralis

<400> 31

Met Glu Asp Ala Lys Asn Ile Lys Lys Gly Pro Ala Pro Phe Tyr Pro
 1 5 10 15
 Leu Glu Asp Gly Thr Ala Gly Glu Gln Leu His Lys Ala Met Lys Arg
 20 25 30
 Tyr Ala Leu Val Pro Gly Thr Ile Ala Phe Thr Asp Ala His Ile Glu
 35 40 45
 Val Asn Ile Thr Tyr Ala Glu Tyr Phe Glu Met Ser Val Arg Leu Ala
 50 55 60
 Glu Ala Met Lys Arg Tyr Gly Leu Asn Thr Asn His Arg Ile Val Val
 65 70 75 80
 Cys Ser Glu Asn Ser Leu Gln Phe Phe Met Pro Val Leu Gly Ala Leu
 85 90 95
 Phe Ile Gly Val Ala Val Ala Pro Ala Asn Asp Ile Tyr Asn Glu Arg

100	105	110
Glu Leu Leu Asn Ser Met Asn Ile Ser Gln Pro Thr Val Val Phe Val		
115	120	125
Ser Lys Lys Gly Leu Gln Lys Ile Leu Asn Val Gln Lys Lys Leu Pro		
130	135	140
Ile Ile Gln Lys Ile Ile Met Asp Ser Lys Thr Asp Tyr Gln Gly		
145	150	155
Phe Gln Ser Met Tyr Thr Phe Val Thr Ser His Leu Pro Pro Gly Phe		
165	170	175
Asn Glu Tyr Asp Phe Val Pro Glu Ser Phe Asp Arg Asp Lys Thr Ile		
180	185	190
Ala Leu Ile Met Asn Ser Ser Gly Ser Thr Gly Leu Pro Lys Gly Val		
195	200	205
Ala Leu Pro His Arg Thr Ala Cys Val Arg Phe Ser His Ala Arg Asp		
210	215	220
Pro Ile Phe Gly Asn Gln Ile Ile Pro Asp Thr Ala Ile Leu Ser Val		
225	230	235
Val Pro Phe His His Gly Phe Gly Met Phe Thr Thr Leu Gly Tyr Leu		
245	250	255
Ile Cys Gly Phe Arg Val Val Leu Met Tyr Arg Phe Glu Glu Leu		
260	265	270
Phe Leu Arg Ser Leu Gln Asp Tyr Lys Ile Gln Ser Ala Leu Leu Val		
275	280	285
Pro Thr Leu Phe Ser Phe Phe Ala Lys Ser Thr Leu Ile Asp Lys Tyr		
290	295	300
Asp Leu Ser Asn Leu His Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser		
305	310	315
Lys Glu Val Gly Glu Ala Val Ala Lys Arg Phe His Leu Pro Gly Ile		
325	330	335
Arg Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Ile Leu Ile Thr		
340	345	350
Pro Glu Gly Asp Asp Lys Pro Gly Ala Val Gly Lys Val Val Pro Phe		
355	360	365
Phe Glu Ala Lys Val Val Asp Leu Asp Thr Gly Lys Thr Leu Gly Val		
370	375	380
Asn Gln Arg Gly Glu Leu Cys Val Arg Gly Pro Met Ile Met Ser Gly		
385	390	395
Tyr Val Asn Asn Pro Glu Ala Thr Asn Ala Leu Ile Asp Lys Asp Gly		
405	410	415
Trp Leu His Ser Gly Asp Ile Ala Tyr Trp Asp Glu Asp Glu His Phe		
420	425	430
Phe Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln		

435	440	445
Val Ala Pro Ala Glu Leu Glu Ser Ile Leu Leu Gln His Pro Asn Ile		
450	455	460
Phe Asp Ala Gly Val Ala Gly Leu Pro Asp Asp Asp Ala Gly Glu Leu		
465	470	475
Pro Ala Ala Val Val Val Leu Glu His Gly Lys Thr Met Thr Glu Lys		
485	490	495
Glu Ile Val Asp Tyr Val Ala Ser Gln Val Thr Thr Ala Lys Lys Leu		
500	505	510
Arg Gly Gly Val Val Phe Val Asp Glu Val Pro Lys Gly Leu Thr Gly		
515	520	525
Lys Leu Asp Ala Arg Lys Ile Arg Glu Ile Leu Ile Lys Ala Lys Lys		
530	535	540
Gly Gly Lys Ser Lys Leu		
545	550	

<210> 32
 <211> 547
 <212> PRT
 <213> Lampyris noctiluca

<400> 32		
Met Glu Asp Ala Lys Asn Ile Met His Gly Pro Ala Pro Phe Tyr Pro		
1	5	10
Leu Glu Asp Gly Thr Ala Gly Glu Gln Leu His Lys Ala Met Lys Arg		
20	25	30
Tyr Ala Gln Val Pro Gly Thr Ile Ala Phe Thr Asp Ala His Ala Glu		
35	40	45
Val Asn Ile Thr Tyr Ser Glu Tyr Phe Glu Met Ala Cys Arg Leu Ala		
50	55	60
Glu Thr Met Lys Arg Tyr Gly Leu Gly Leu Gln His His Ile Ala Val		
65	70	75
Cys Ser Glu Asn Ser Leu Gln Phe Phe Met Pro Val Cys Gly Ala Leu		
85	90	95
Phe Ile Gly Val Gly Val Ala Ser Thr Asn Asp Ile Tyr Asn Glu Arg		
100	105	110
Glu Leu Tyr Asn Ser Leu Ser Ile Ser Gln Pro Thr Ile Val Ser Cys		
115	120	125
Ser Lys Arg Ala Leu Gln Lys Ile Leu Gly Val Gln Lys Lys Leu Pro		
130	135	140
Ile Ile Gln Lys Ile Val Ile Leu Asp Ser Arg Glu Asp Tyr Met Gly		
145	150	155
		160

Lys Gln Ser Met Tyr Ser Phe Ile Glu Ser His Leu Pro Ala Gly Phe
 165 170 175
 Asn Glu Tyr Asp Tyr Ile Pro Asp Ser Phe Asp Arg Glu Thr Ala Thr
 180 185 190
 Ala Leu Ile Met Asn Ser Ser Gly Ser Thr Gly Leu Pro Lys Gly Val
 195 200 205
 Glu Leu Thr His Gln Asn Val Cys Val Arg Phe Ser His Cys Arg Asp
 210 215 220
 Pro Val Phe Gly Asn Gln Ile Ile Pro Asp Thr Ala Ile Leu Thr Val
 225 230 235 240
 Ile Pro Phe His His Gly Phe Gly Met Phe Thr Thr Leu Gly Tyr Leu
 245 250 255
 Thr Cys Gly Phe Arg Ile Val Leu Met Tyr Arg Phe Glu Glu Glu Leu
 260 265 270
 Phe Leu Arg Ser Leu Gln Asp Tyr Lys Ile Gln Ser Ala Leu Leu Val
 275 280 285
 Pro Thr Leu Phe Ser Phe Phe Ala Lys Ser Thr Leu Val Asp Lys Tyr
 290 295 300
 Asp Leu Ser Asn Leu His Glu Ile Ala Ser Gly Gly Ala Pro Leu Ala
 305 310 315 320
 Lys Glu Val Gly Glu Ala Val Ala Lys Arg Phe Lys Leu Pro Gly Ile
 325 330 335
 Arg Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Ile Ile Ile Thr
 340 345 350
 Pro Glu Gly Asp Asp Lys Pro Gly Ala Cys Gly Lys Val Val Pro Phe
 355 360 365
 Phe Ser Ala Lys Ile Val Asp Leu Asp Thr Gly Lys Thr Leu Gly Val
 370 375 380
 Asn Gln Arg Gly Glu Leu Cys Val Lys Gly Pro Met Ile Met Lys Gly
 385 390 395 400
 Tyr Val Asn Asn Pro Glu Ala Thr Ser Ala Leu Ile Asp Lys Asp Gly
 405 410 415
 Trp Leu His Ser Gly Asp Ile Ala Tyr Tyr Asp Lys Asp Gly His Phe
 420 425 430
 Phe Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln
 435 440 445
 Val Pro Pro Ala Glu Leu Glu Ser Ile Leu Leu Gln His Pro Phe Ile
 450 455 460
 Phe Asp Ala Gly Val Ala Gly Ile Pro Asp Pro Asp Ala Gly Glu Leu
 465 470 475 480
 Pro Ala Ala Val Val Val Leu Glu Glu Gly Lys Thr Met Thr Glu Gln
 485 490 495

Glu Val Met Asp Tyr Val Ala Gly Gln Val Thr Ala Ser Lys Arg Leu
500 505 510
Arg Gly Gly Val Lys Phe Val Asp Glu Val Pro Lys Gly Leu Thr Gly
515 520 525
Lys Ile Asp Gly Arg Lys Ile Arg Glu Ile Leu Met Met Gly Lys Lys
530 535 540
Ser Lys Leu
545

<210> 33
<211> 552
<212> PRT
<213> Photuris pennsylvanica

<400> 33

Met Ser Ile Glu Asn Asn Ile Leu Ile Gly Pro Pro Pro Tyr Tyr Pro
1 5 10 15
Leu Glu Glu Gly Thr Ala Gly Glu Gln Leu His Arg Ala Ile Ser Arg
20 25 30
Tyr Ala Ala Val Pro Gly Thr Leu Ala Tyr Thr Asp Val His Thr Glu
35 40 45
Leu Glu Val Thr Tyr Lys Glu Phe Leu Asp Val Thr Cys Arg Leu Ala
50 55 60
Glu Ala Met Lys Asn Tyr Gly Leu Gly Leu Gln His Thr Ile Ser Val
65 70 75 80
Cys Ser Glu Asn Cys Val Gln Phe Phe Met Pro Ile Cys Ala Ala Leu
85 90 95
Tyr Val Gly Val Ala Thr Ala Pro Thr Asn Asp Ile Tyr Asn Glu Arg
100 105 110
Glu Leu Tyr Asn Ser Leu Ser Ile Ser Gln Pro Thr Val Val Phe Thr
115 120 125
Ser Arg Asn Ser Leu Gln Lys Ile Leu Gly Val Gln Ser Arg Leu Pro
130 135 140
Ile Ile Lys Lys Ile Ile Ile Leu Asp Gly Lys Lys Asp Tyr Leu Gly
145 150 155 160
Tyr Gln Ser Met Gln Ser Phe Met Lys Glu His Val Pro Ala Asn Phe
165 170 175
Asn Val Ser Ala Phe Lys Pro Leu Ser Phe Asp Leu Asp Arg Val Ala
180 185 190
Cys Ile Met Asn Ser Ser Gly Ser Thr Gly Leu Pro Lys Gly Val Pro
195 200 205
Ile Ser His Arg Asn Thr Ile Tyr Arg Phe Ser His Cys Arg Asp Pro

210 215 220
Val Phe Gly Asn Gln Ile Ile Pro Asp Thr Thr Ile Leu Cys Ala Val
225 230 235 240
Pro Phe His His Ala Phe Gly Thr Phe Thr Asn Leu Gly Tyr Leu Ile
245 250 255
Cys Gly Phe His Val Val Leu Met Tyr Arg Phe Asn Glu His Leu Phe
260 265 270
Leu Gln Thr Leu Gln Asp Tyr Lys Cys Gln Ser Ala Leu Leu Val Pro
275 280 285
Thr Val Leu Ala Phe Leu Ala Lys Asn Pro Leu Val Asp Lys Tyr Asp
290 295 300
Leu Ser Asn Leu His Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys
305 310 315 320
Glu Ile Ser Glu Ile Ala Ala Lys Arg Phe Lys Leu Pro Gly Ile Arg
325 330 335
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Cys Ala Ile Val Ile Thr Ala
340 345 350
Glu Gly Glu Phe Lys Leu Gly Ala Val Gly Lys Val Val Pro Phe Tyr
355 ← 360 365
Ser Leu Lys Val Leu Asp Leu Asn Thr Gly Lys Lys Leu Gly Pro Asn
370 375 380
Glu Arg Gly Glu Ile Cys Phe Lys Gly Pro Met Ile Met Lys Gly Tyr
385 390 395 400
Ile Asn Asn Pro Glu Ala Thr Arg Glu Leu Ile Asp Glu Glu Gly Trp
405 410 415
Ile His Ser Gly Asp Ile Gly Tyr Phe Asp Glu Asp Gly His Val Tyr
420 425 430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val
435 440 445
Pro Pro Ala Glu Leu Glu Ala Leu Leu Leu Gln His Pro Phe Ile Glu
450 455 460
Asp Ala Gly Val Ala Gly Val Pro Asp Glu Val Ala Gly Asp Leu Pro
465 470 475 480
Gly Ala Val Val Val Leu Lys Glu Gly Lys Ser Ile Thr Glu Lys Glu
485 490 495
Ile Gln Asp Tyr Val Ala Gly Gln Val Thr Ser Ser Lys Lys Leu Arg
500 505 510
Gly Gly Val Glu Phe Val Lys Glu Val Pro Lys Gly Phe Thr Gly Lys
515 520 525
Ile Asp Thr Arg Lys Ile Lys Glu Ile Leu Ile Lys Ala Gln Lys Gly
530 535 540
Lys Ser Lys Ser Lys Ala Lys Leu

545

550

<210> 34
<211> 546
<212> PRT
<213> Phengodes sp.

<400> 34

Met Ile Lys Met Glu Glu Glu His Val Met Pro Gly Ala Met Pro Arg
1 5 10 15
Asp Leu Leu Phe Glu Gly Thr Ala Gly Gln Gln Leu His Arg Ala Leu
20 25 30
Tyr Lys His Ser Tyr Phe Pro Glu Ala Ile Val Asp Ser His Thr His
35 40 45
Glu Ile Ile Ser Tyr Ala Lys Ile Leu Asp Met Ser Cys Arg Leu Ala
50 55 60
Val Ser Phe Gln Lys Tyr Gly Leu Thr Gln Asn Asn Ile Ile Gly Ile
65 70 75 80
Cys Ser Glu Asn Asn Leu Asn Phe Phe Asn Pro Val Ile Ala Ala Phe
85 90 95
Tyr Leu Gly Ile Thr Val Ala Thr Val Asn Asp Thr Tyr Thr Asp Arg
100 105 110
Glu Leu Ser Glu Thr Leu Asn Ile Thr Lys Pro Gln Met Leu Phe Cys
115 120 125
Ser Lys Gln Ser Leu Pro Ile Val Met Lys Thr Met Lys Ile Met Pro
130 135 140
Tyr Val Gln Lys Leu Leu Ile Ile Asp Ser Met Gln Asp Ile Gly Gly
145 150 155 160
Ile Glu Cys Val His Ser Phe Val Ser Arg Tyr Thr Asp Glu His Phe
165 170 175
Asp Pro Leu Lys Phe Val Pro Leu Asp Phe Asp Pro Arg Glu Gln Val
180 185 190
Ala Leu Ile Met Thr Ser Ser Gly Thr Thr Gly Leu Pro Lys Gly Val
195 200 205
Met Leu Thr His Arg Asn Ile Cys Val Arg Phe Val His Ser Arg Asp
210 215 220
Pro Leu Phe Gly Thr Arg Phe Ile Pro Glu Thr Ser Ile Leu Ser Leu
225 230 235 240
Val Pro Phe His His Ala Phe Gly Met Phe Thr Thr Leu Ser Tyr Phe
245 250 255
Ile Val Gly Leu Lys Ile Val Met Met Lys Arg Phe Asp Gly Glu Leu
260 265 270

Phe Leu Lys Thr Ile Gln Asn Tyr Lys Ile Pro Thr Ile Val Ile Ala
275 280 285
Pro Pro Val Met Val Phe Leu Ala Lys Ser His Leu Val Asp Lys Tyr
290 295 300
Asp Leu Ser Ser Ile Lys Glu Ile Ala Thr Gly Gly Ala Pro Leu Gly
305 310 315 320
Pro Ala Leu Ala Asn Ala Val Ala Lys Arg Leu Lys Leu Gly Gly Ile
325 330 335
Ile Gln Gly Tyr Gly Leu Thr Glu Thr Cys Cys Ala Val Leu Ile Thr
340 345 350
Pro His Asn Lys Ile Lys Thr Gly Ser Thr Gly Gln Val Leu Pro Tyr
355 360 365
Val Thr Ala Lys Ile Val Asp Thr Lys Thr Gly Lys Asn Leu Gly Pro
370 375 380
Asn Gln Thr Gly Glu Leu Cys Phe Lys Ser Asp Ile Ile Met Lys Gly
385 390 395 400
Tyr Tyr Gln Asn Glu Glu Glu Thr Arg Leu Val Ile Asp Lys Asp Gly
405 410 415
Trp Leu His Ser Gly Asp Ile Gly Tyr Tyr Asp Thr Asp Gly Asn Phe
420 425 430
His Ile Val Asp Arg Leu Lys Glu Leu Ile Lys Tyr Lys Ala Tyr Gln
435 440 445
Val Ala Pro Ala Glu Leu Glu Ala Leu Leu Leu Gln His Pro Tyr Ile
450 455 460
Ala Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Glu Ala Gly Glu Leu
465 470 475 480
Pro Ala Ala Cys Val Val Leu Glu Pro Gly Lys Thr Met Thr Glu Lys
485 490 495
Glu Val Met Asp Tyr Ile Ala Glu Arg Val Thr Pro Thr Lys Arg Leu
500 505 510
Arg Gly Gly Val Leu Phe Val Asn Asn Ile Pro Lys Gly Ala Thr Gly
515 520 525
Lys Leu Val Arg Thr Glu Leu Arg Arg Leu Leu Thr Gln Arg Ala Ala
530 535 540
Lys Leu
545

<210> 35

<211> 543

<212> PRT

<213> Pyrophorus plagiophthalamus

<400> 35
 Met Met Lys Arg Glu Lys Asn Val Val Tyr Gly Pro Glu Pro Leu His
 1 5 10 15
 Pro Leu Glu Asp Leu Thr Ala Gly Glu Met Leu Phe Arg Ala Leu Arg
 20 25 30
 Lys His Ser His Leu Pro Gln Ala Leu Val Asp Val Tyr Gly Glu Glu
 35 40 45
 Trp Ile Ser Tyr Lys Glu Phe Phe Glu Thr Thr Cys Leu Leu Ala Gln
 50 55 60
 Ser Leu His Asn Cys Gly Tyr Lys Met Ser Asp Val Val Ser Ile Cys
 65 70 75 80
 Ala Glu Asn Asn Lys Arg Phe Phe Val Pro Ile Ile Ala Ala Trp Tyr
 85 90 95
 Ile Gly Met Ile Val Ala Pro Val Asn Glu Gly Tyr Ile Pro Asp Glu
 100 105 110
 Leu Cys Lys Val Met Gly Ile Ser Arg Pro Gln Leu Val Phe Cys Thr
 115 120 125
 Lys Asn Ile Leu Asn Lys Val Leu Glu Val Gln Ser Arg Thr Asp Phe
 130 135 140
 Ile Lys Arg Ile Ile Ile Leu Asp Ala Val Glu Asn Ile His Gly Cys
 145 150 155 160
 Glu Ser Leu Pro Asn Phe Ile Ser Arg Tyr Ser Asp Gly Asn Ile Ala
 165 170 175
 Asn Phe Lys Pro Leu His Tyr Asp Pro Val Glu Gln Val Ala Ala Ile
 180 185 190
 Leu Cys Ser Ser Gly Thr Thr Gly Leu Pro Lys Gly Val Met Gln Thr
 195 200 205
 His Arg Asn Val Cys Val Arg Leu Ile His Ala Leu Asp Pro Arg Val
 210 215 220
 Gly Thr Gln Leu Ile Pro Gly Val Thr Val Leu Val Tyr Leu Pro Phe
 225 230 235 240
 Phe His Ala Phe Gly Phe Ser Ile Asn Leu Gly Tyr Phe Met Val Gly
 245 250 255
 Leu Arg Val Ile Met Leu Arg Arg Phe Asp Gln Glu Ala Phe Leu Lys
 260 265 270
 Ala Ile Gln Asp Tyr Glu Val Arg Ser Val Ile Asn Val Pro Ala Ile
 275 280 285
 Ile Leu Phe Leu Ser Lys Ser Pro Leu Val Asp Lys Tyr Asp Leu Ser
 290 295 300
 Ser Leu Arg Glu Leu Cys Cys Gly Ala Ala Pro Leu Ala Lys Glu Val
 305 310 315 320
 Ala Glu Ile Ala Val Lys Arg Leu Asn Leu Pro Gly Ile Arg Cys Gly

325	330	335
Phe Gly Leu Thr Glu Ser Thr Ser Ala Asn Ile His Ser Leu Arg Asp		
340	345	350
Glu Phe Lys Ser Gly Ser Leu Gly Arg Val Thr Pro Leu Met Ala Ala		
355	360	365
Lys Ile Ala Asp Arg Glu Thr Gly Lys Ala Leu Gly Pro Asn Gln Val		
370	375	380
Gly Glu Leu Cys Ile Lys Gly Pro Met Val Ser Lys Gly Tyr Val Asn		
385	390	395
Asn Val Glu Ala Thr Lys Glu Ala Ile Asp Asp Asp Gly Trp Leu His		
405	410	415
Ser Gly Asp Phe Gly Tyr Tyr Asp Glu Asp Glu His Phe Tyr Val Val		
420	425	430
Asp Arg Tyr Lys Glu Leu Ile Lys Tyr Lys Gly Ser Gln Val Ala Pro		
435	440	445
Ala Glu Leu Glu Glu Ile Leu Leu Lys Asn Pro Cys Ile Arg Asp Val		
450	455	460
Ala Val Val Gly Ile Pro Asp Leu Glu Ala Gly Glu Leu Pro Ser Ala		
465	470	475
Phe Val Val Ile Gln Pro Gly Lys Glu Ile Thr Ala Lys Glu Val Tyr		
485	490	495
Asp Tyr Leu Ala Glu Arg Val Ser His Thr Lys Tyr Leu Arg Gly Gly		
500	505	510
Val Arg Phe Val Asp Ser Ile Pro Arg Asn Val Thr Gly Lys Ile Thr		
515	520	525
Arg Lys Glu Leu Leu Lys Gln Leu Leu Glu Lys Ser Ser Lys Leu		
530	535	540

<210> 36
 <211> 543
 <212> PRT
 <213> Pyrophorus plagiophthalmus

<400> 36			
Met Met Lys Arg Glu Lys Asn Val Ile Tyr Gly Pro Glu Pro Leu His			
1	5	10	15
Pro Leu Glu Asp Leu Thr Ala Gly Glu Met Leu Phe Arg Ala Leu Arg			
20	25	30	
Lys His Ser His Leu Pro Gln Ala Leu Val Asp Val Phe Gly Asp Glu			
35	40	45	
Ser Leu Ser Tyr Lys Glu Phe Phe Glu Ala Thr Cys Leu Leu Ala Gln			
50	55	60	

Ser Leu His Asn Cys Gly Tyr Lys Met Asn Asp Val Val Ser Ile Cys
 65 70 75 80
 Ala Glu Asn Asn Lys Arg Phe Phe Ile Pro Ile Ile Ala Ala Trp Tyr
 85 90 95
 Ile Gly Met Ile Val Ala Pro Val Asn Glu Ser Tyr Ile Pro Asp Glu
 100 105 110
 Leu Cys Lys Val Met Gly Ile Ser Lys Pro Gln Ile Val Phe Cys Thr
 115 120 125
 Lys Asn Ile Leu Asn Lys Val Leu Glu Val Gln Ser Arg Thr Asn Phe
 130 135 140
 Ile Lys Arg Ile Ile Leu Asp Thr Val Glu Asn Ile His Gly Cys
 145 150 155 160
 Glu Ser Leu Pro Asn Phe Ile Ser Arg Tyr Ser Asp Gly Asn Ile Ala
 165 170 175
 Asn Phe Lys Pro Leu His Tyr Asp Pro Val Glu Gln Val Ala Ala Ile
 180 185 190
 Leu Cys Ser Ser Gly Thr Thr Gly Leu Pro Lys Gly Val Met Gln Thr
 195 200 205
 His Gln Asn Ile Cys Val Arg Leu Ile His Ala Leu Asp Pro Arg Ala
 210 215 220
 Gly Thr Gln Leu Ile Pro Gly Val Thr Val Leu Val Tyr Leu Pro Phe
 225 230 235 240
 Phe His Ala Phe Gly Phe Ser Ile Asn Leu Gly Tyr Phe Met Val Gly
 245 250 255
 Leu Arg Val Ile Met Leu Arg Arg Phe Asp Gln Glu Ala Phe Leu Lys
 260 265 270
 Ala Ile Gln Asp Tyr Glu Val Arg Ser Val Ile Asn Val Pro Ala Ile
 275 280 285
 Ile Leu Phe Leu Ser Lys Ser Pro Leu Val Asp Lys Tyr Asp Leu Ser
 290 295 300
 Ser Leu Arg Glu Leu Cys Cys Gly Ala Ala Pro Leu Ala Lys Glu Val
 305 310 315 320
 Ala Glu Val Ala Val Lys Arg Leu Asn Leu Pro Gly Ile Arg Cys Gly
 325 330 335
 Phe Gly Leu Thr Glu Ser Thr Ser Ala Asn Ile His Ser Leu Gly Asp
 340 345 350
 Glu Phe Lys Ser Gly Ser Leu Gly Arg Val Thr Pro Leu Met Ala Ala
 355 360 365
 Lys Ile Ala Asp Arg Glu Thr Gly Lys Ala Leu Gly Pro Asn Gln Val
 370 375 380
 Gly Glu Leu Cys Val Lys Gly Pro Met Val Ser Lys Gly Tyr Val Asn
 385 390 395 400

Asn Val Glu Ala Thr Lys Glu Ala Ile Asp Asp Asp Gly Trp Leu His
 405 410 415
 Ser Gly Asp Phe Gly Tyr Tyr Asp Glu Asp Glu His Phe Tyr Val Val
 420 425 430
 Asp Arg Tyr Lys Glu Leu Ile Lys Tyr Lys Gly Ser Gln Val Ala Pro
 435 440 445
 Ala Glu Leu Glu Glu Ile Leu Leu Lys Asn Pro Cys Ile Arg Asp Val
 450 455 460
 Ala Val Val Gly Ile Pro Asp Leu Glu Ala Gly Glu Leu Pro Ser Ala
 465 470 475 480
 Phe Val Val Lys Gln Pro Gly Lys Glu Ile Thr Ala Lys Glu Val Tyr
 485 490 495
 Asp Tyr Leu Ala Glu Arg Val Ser His Thr Lys Tyr Leu Arg Gly Gly
 500 505 510
 Val Arg Phe Val Asp Ser Ile Pro Arg Asn Val Thr Gly Lys Ile Thr
 515 520 525
 Arg Lys Glu Leu Leu Lys Gln Leu Leu Glu Lys Ser Ser Lys Leu
 530 535 540

<210> 37
 <211> 545
 <212> PRT
 <213> Photuris pennsylvanica

<400> 37

Met Glu Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu
 1 5 10 15
 Ala Asp Gly Thr Ala Gly Glu Gln Met Phe Tyr Ala Leu Ser Arg Tyr
 20 25 30
 Ala Asp Ile Ser Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu
 35 40 45
 Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu
 50 55 60
 Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys
 65 70 75 80
 Ser Glu Asn Gly Leu Gln Phe Phe Leu Pro Leu Ile Ala Ser Leu Tyr
 85 90 95
 Leu Gly Ile Ile Ala Ala Pro Val Ser Asp Lys Tyr Ile Glu Arg Glu
 100 105 110
 Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Ile Phe Cys Ser
 115 120 125
 Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Tyr

130	135	140
Val Glu Thr Ile Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr		
145	150	155
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ile Asn Leu Asp		
165	170	175
Val Lys Lys Phe Lys Pro Asn Ser Phe Asn Arg Asp Asp Gln Val Ala		
180	185	190
Leu Val Met Phe Ser Ser Gly Thr Thr Gly Val Ser Lys Gly Val Met		
195	200	205
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser His Cys Lys Asp Pro		
210	215	220
Thr Phe Gly Asn Ala Ile Asn Pro Thr Thr Ala Ile Leu Thr Val Ile		
225	230	235
Pro Phe His His Gly Phe Gly Met Thr Thr Thr Leu Gly Tyr Phe Thr		
245	250	255
Cys Gly Phe Arg Val Ala Leu Met His Thr Phe Glu Glu Lys Leu Phe		
260	265	270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro		
275	280	285
Thr Leu Met Ala Phe Phe Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp		
290	295	300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys		
305	310	315
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg		
325	330	335
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro		
340	345	350
Asp Thr Asp Val Arg Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His		
355	360	365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn		
370	375	380
Glu Thr Gly Glu Leu Tyr Phe Lys Gly Asp Met Ile Met Lys Ser Tyr		
385	390	395
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asn Lys Asp Gly Trp		
405	410	415
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr		
420	425	430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val		
435	440	445
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val		
450	455	460
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro		

465 470 475 480
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile
485 490 495
Val Gln Asn Phe Val Ser Ser Gln Val Ser Thr Ala Lys Trp Leu Arg
500 505 510
Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys
515 520 525
Ile Asp Arg Lys Val Leu Arg Gln Met Phe Glu Lys His Lys Ser Lys
530 535 540
Leu
545

<210> 38
<211> 38
<212> DNA
<213> Artificial Sequence

<220>
<223> /note = "a primer"

<400> 38
gtactgagac gacgccagcc caagcttagg cctgagtg

38

<210> 39
<211> 38
<212> DNA
<213> Artificial Sequence

<220>
<223> /note = "a primer"

<400> 39
ggcatgagcg tgaactgact gaactagcgg ccgcccag

38

<210> 40
<211> 18
<212> DNA
<213> Artificial Sequence

<220>
<223> /note = "a primer"

aaaaattttg gggccaaatg aacctggaga attgtatTTT aaaggcccga tgataatgaa	1200
gggttattat aataatgaag aagctactaa agcaattatt gataatgacg gatggttgcg	1260
ctctggtgat attgcttatt atgacaatga tggccatTTT tatattgtgg acaggctgaa	1320
gtcactgatt aaatataaag gttatcaggT tgcacctgct gaaattgagg gaatactctt	1380
acaacatccg tatattgttg atgcccggcgt tactggtata ccggatgaaag ccgcgggcga	1440
gcttccagct gcagggtttg tagtacagac tggaaaatAT ctaaacgaaac aaatcgata	1500
agattatgtt gccagtcaag tttcaacagc caaatggcta cgtggtgggg tgatatTTT	1560
ggatgaaatt cccaaaggat caactggaaa aattgacaga aaagtgttaa gacaaaatgtt	1620
agaaaaaacac accaatggg	1639

<210> 43

<211> 1639

<212> DNA

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 43

ggatccaaatg gcagataaga atatTTTata tgggccccaa ccattttatc ccttggaaaga	60
tgggacggct ggagaacaga tggTTTgacgc attatctcgT tatgcagcta ttccgggctg	120
catagcattg acaaATgctc atacaaaaga aaatgttttA tatgaagagt ttctgaaact	180
gtcgtgtcgT tttagggaaa gtttAAAAA gtaggatta aaacaaaacg acacaatagc	240
ggTgtgtagc gaaaatagtc tgcaatTTT cttccTgtA attgcatcat tgtatcttgg	300
aataattgtg gcacctgttA acgataaata cattgaacgt gaattaatac acagtcttgg	360
tattgtaaaa ccacgcatac ttTTTgctc caagaatac ttcaaaaag tactgaatgt	420
aaaatctaaa ttAAAatcta ttgaaaactat tattatatta gacttAAatg aagacttgg	480
agtttatcaa tgcctcaaca acTTTatttc tcaaaattcc gatagtaatc tggacgtaaa	540
aaaatTTaaa ccctattctt ttaatcgaga cgatcaggTT gcgtcgatta tggTTTcttc	600
tggTacaact ggtctgccga agggagtcat gctaactcac aagaatattg ttgcacgatt	660
ttctattgca aaagatccta ctTTTggtaa cgcaattaat cccacgtcag caatTTtaac	720
ggtaataacct ttccaccatg gtttggTat gatgaccaca ttaggatact ttacttgg	780
atcccagtt gttctaattgc acacgTTtga agaaaaacta ttctacaat cattacaaga	840
ttataaaagtg gaaagtactt tacttggTacc aacattaatg gcatttcttgc caaaaagtgc	900
attagttgaa aagtacgatt tatcgactt aaaagaaatt gcatctggTg ggcacccTT	960
atcaaaaagaa attggggaga tggtaaaaaa acggTTaaa ttAAactttg tcaggcaagg	1020
gtatggatta acagaaacca ctTcggtgt tttattaca ccgaaaggTg acgccaAAacc	1080
gggatcaact ggtaaaatag taccattaca cgctgttaaa gttgtcgatc ctacaacagg	1140
aaaaatTTtgg gggccaaatg aacctggaga attgtatTTT aaaggcccga tgataatgaa	1200
gggttattat aataatgaag aagctactaa agcaattatt gataatgacg gatggttgcg	1260
ctctggtgat attgcttatt atgacaatga tggccatTTT tatattgtgg acaggctgaa	1320
gtcactgatt aaatataaag gttatcaggT tgcacctgct gaaattgagg gaatactctt	1380

acaacatccg tatattgttg atgccggcgt tactggtata ccggatgaag ccgcgggcga	1440
gcttccagct gcaggtgttg tagtacagac tggaaaatat ctaaacgaac aaatcgtaca	1500
agattatgtt gccagtcaag tttcaacagc caaatggcta cgtggtgggg tgaaaatttt	1560
ggatgaaatt cccaaaggat caactggaaa aattgacaga aaagtgttaa gacaaatgtt	1620
agaaaaaacac accaatggg	1639

<210> 44

<211> 544

<212> PRT

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 44

Met Ala Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu			
1	5	10	15
Glu Asp Gly Thr Ala Gly Glu Gln Met Phe Asp Ala Leu Ser Arg Tyr			
20	25	30	
Ala Asp Ile Pro Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu			
35	40	45	
Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu			
50	55	60	
Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys			
65	70	75	80
Ser Glu Asn Ser Leu Gln Phe Phe Leu Pro Val Ile Ala Ser Leu Tyr			
85	90	95	
Leu Gly Ile Ile Val Ala Pro Val Asn Asp Lys Tyr Ile Glu Arg Glu			
100	105	110	
Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Val Phe Cys Ser			
115	120	125	
Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Ser			
130	135	140	
Ile Glu Thr Ile Ile Leu Asp Leu Asn Asp Asp Leu Gly Gly Tyr			
145	150	155	160
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ser Asn Leu Asp			
165	170	175	
Val Lys Lys Phe Lys Pro Tyr Ser Phe Asn Arg Asp Asp Gln Val Ala			
180	185	190	
Leu Ile Met Phe Ser Ser Gly Thr Thr Gly Leu Pro Lys Gly Val Met			
195	200	205	
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser Ile Ala Lys Asp Pro			

210	215	220
Thr Phe Gly Asn Ala Ile Asn Pro Thr Ser Ala Ile Leu Thr Val Ile		
225	230	235
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr		
245	250	255
Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe		
260	265	270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro		
275	280	285
Thr Leu Met Ala Phe Leu Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp		
290	295	300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys		
305	310	315
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg		
325	330	335
Gln Gly Tyr Gly Leu Thr Glu Thr Ser Ala Val Leu Ile Thr Pro		
340	345	350
Lys Gly Asp Ala Lys Pro Gly Ser Thr Gly Lys Ile Val Pro Phe His		
355	360	365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn		
370	375	380
Glu Pro Gly Glu Leu Tyr Phe Lys Gly Pro Met Ile Met Lys Gly Tyr		
385	390	395
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asp Asn Asp Gly Trp		
405	410	415
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr		
420	425	430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val		
435	440	445
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val		
450	455	460
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro		
465	470	475
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile		
485	490	495
Val Gln Asp Tyr Val Ala Ser Gln Val Ser Thr Ala Lys Trp Leu Arg		
500	505	510
Gly Gly Val Ile Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys		
515	520	525
Ile Asp Arg Lys Val Leu Arg Gln Met Leu Glu Lys His Thr Asn Gly		
530	535	540

<210> 45
<211> 544
<212> PRT
<213> Artificial Sequence

<220>
<223> /note = "mutant luciferase"

<400> 45

Met Ala Asp Lys Asn Ile Leu Tyr Gly Pro Glu Pro Phe Tyr Pro Leu
1 5 10 15
Glu Asp Gly Thr Ala Gly Glu Gln Met Phe Asp Ala Leu Ser Arg Tyr
20 25 30
Ala Ala Ile Pro Gly Cys Ile Ala Leu Thr Asn Ala His Thr Lys Glu
35 40 45
Asn Val Leu Tyr Glu Glu Phe Leu Lys Leu Ser Cys Arg Leu Ala Glu
50 55 60
Ser Phe Lys Lys Tyr Gly Leu Lys Gln Asn Asp Thr Ile Ala Val Cys
65 70 75 80
Ser Glu Asn Ser Leu Gln Phe Phe Leu Pro Val Ile Ala Ser Leu Tyr
85 90 95
Leu Gly Ile Ile Val Ala Pro Val Asn Asp Lys Tyr Ile Glu Arg Glu
100 105 110
Leu Ile His Ser Leu Gly Ile Val Lys Pro Arg Ile Val Phe Cys Ser
115 120 125
Lys Asn Thr Phe Gln Lys Val Leu Asn Val Lys Ser Lys Leu Lys Ser
130 135 140
Ile Glu Thr Ile Ile Leu Asp Leu Asn Glu Asp Leu Gly Gly Tyr
145 150 155 160
Gln Cys Leu Asn Asn Phe Ile Ser Gln Asn Ser Asp Ser Asn Leu Asp
165 170 175
Val Lys Lys Phe Lys Pro Tyr Ser Phe Asn Arg Asp Asp Gln Val Ala
180 185 190
Ser Ile Met Phe Ser Ser Gly Thr Thr Gly Leu Pro Lys Gly Val Met
195 200 205
Leu Thr His Lys Asn Ile Val Ala Arg Phe Ser Ile Ala Lys Asp Pro
210 215 220
Thr Phe Gly Asn Ala Ile Asn Pro Thr Ser Ala Ile Leu Thr Val Ile
225 230 235 240
Pro Phe His His Gly Phe Gly Met Met Thr Thr Leu Gly Tyr Phe Thr
245 250 255
Cys Gly Phe Arg Val Val Leu Met His Thr Phe Glu Glu Lys Leu Phe

260	265	270
Leu Gln Ser Leu Gln Asp Tyr Lys Val Glu Ser Thr Leu Leu Val Pro		
275	280	285
Thr Leu Met Ala Phe Leu Ala Lys Ser Ala Leu Val Glu Lys Tyr Asp		
290	295	300
Leu Ser His Leu Lys Glu Ile Ala Ser Gly Gly Ala Pro Leu Ser Lys		
305	310	315
Glu Ile Gly Glu Met Val Lys Lys Arg Phe Lys Leu Asn Phe Val Arg		
325	330	335
Gln Gly Tyr Gly Leu Thr Glu Thr Thr Ser Ala Val Leu Ile Thr Pro		
340	345	350
Lys Gly Asp Ala Lys Pro Gly Ser Thr Gly Lys Ile Val Pro Leu His		
355	360	365
Ala Val Lys Val Val Asp Pro Thr Thr Gly Lys Ile Leu Gly Pro Asn		
370	375	380
Glu Pro Gly Glu Leu Tyr Phe Lys Gly Pro Met Ile Met Lys Gly Tyr		
385	390	395
Tyr Asn Asn Glu Glu Ala Thr Lys Ala Ile Ile Asp Asn Asp Gly Trp		
405	410	415
Leu Arg Ser Gly Asp Ile Ala Tyr Tyr Asp Asn Asp Gly His Phe Tyr		
420	425	430
Ile Val Asp Arg Leu Lys Ser Leu Ile Lys Tyr Lys Gly Tyr Gln Val		
435	440	445
Ala Pro Ala Glu Ile Glu Gly Ile Leu Leu Gln His Pro Tyr Ile Val		
450	455	460
Asp Ala Gly Val Thr Gly Ile Pro Asp Glu Ala Ala Gly Glu Leu Pro		
465	470	475
480		
Ala Ala Gly Val Val Val Gln Thr Gly Lys Tyr Leu Asn Glu Gln Ile		
485	490	495
Val Gln Asp Tyr Val Ala Ser Gln Val Ser Thr Ala Lys Trp Leu Arg		
500	505	510
Gly Gly Val Lys Phe Leu Asp Glu Ile Pro Lys Gly Ser Thr Gly Lys		
515	520	525
Ile Asp Arg Lys Val Leu Arg Gln Met Leu Glu Lys His Thr Asn Gly		
530	535	540

<210> 46

<211> 1633

<212> DNA

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 46

ggatccatg atgaagcgag agaaaaatgt tatatatgga cccgaacccc tacaccctt	60
ggaagactta acagctggag aaatgcttt ccgtgccctt cggaaacatt ctcatttacc	120
gcaggcttta gtagatgtgg ttggcgacga atcgcttcc tataaagagt ttttgaagc	180
gacagtcctc ctagcgaaa gtctccacaa ttgtggatac aagatgaatg atgtatgtc	240
gatctgccc gagaataata caagatttt tattcccggtt attgcagctt ggtatattgg	300
tatgattgtt gcacctgtta atgaaagtta catcccagat gaactctgta aggtcatggg	360
tatatcgaaa ccacaaatag ttttacgac aaagaacatt ttaaataagg tattggaggt	420
acagagcaga actaatttca taaaaaggat catcgactt gatactgttag aaaacataca	480
cgttgtgaa agtcttccca attttatttc tcgttattcg gatggaaata ttgccaactt	540
caaaccttta catttcgatc ctgttagagca agtggcagct atcttatgtt cgtcaggcac	600
tactggatta ccgaaagggtg taatgcaaac tcacccaaat atttgcgtcc gacttataca	660
tgccttagac cccagggcag gaacgcaact tattcctggt gtgacagtct tagtataatct	720
gccttttttc catgcttttgc ggttctctat aaccttgggta tacttcatgg tgggtcttcg	780
tgttatcatg tcaagacat ttgatccaga agcatttcta aaagctattc aggattatga	840
agttcgaagt gtaattaacg ttccatcagt aatattgttc ttatcgaaaa gtcctttgg	900
tgacaaatac gatttatcaa gtttaaggga attgtgttgc ggtgcggcac cattagcaaa	960
agaagttgct gaggttgcag caaaacgatt aaacttgcac ggaattcgct gtggatttgg	1020
tttgacagaa tctacttcag ctaatataca cagtcttagg gatgaattta aaccaggatc	1080
acttggaga gttactcctt taatggcagc taaaatagca gatagggaaa ctggtaaagc	1140
attgggacca aatcaagttt gtgaattatg cattaaaggat cccatggat cgaaaggat	1200
cgtgaacaat gtagaagcta ccaaagaagc tattgtatgat gatggttggc ttcactctgg	1260
agactttggta tactatgtatg aggtatggca tttctatgtg gtggaccgtt acaaggaatt	1320
gattaaatat aagggtcttc aggtatgcacc tgcagaacta gaagagat tattaaaaaa	1380
tccatgtatc agagatgttgc ctgtgggttgg tattcctgtat cttagaagctg gagaactgcc	1440
atctgcgttt gtggtaaac agcccgaaaa ggagattaca gctaaagaag tgtacgatta	1500
tcttgcggag agggctcccc atacaaagta ttgcgttggaa ggggttcgtat tcgttgcata	1560
cataccacgg aatgttacag gtaaaaattac aagaaaggaa cttctgaagc agttgttggaa	1620
gaaggcgggaa ggt	1633

<210> 47

<211> 542

<212> PRT

<213> Artificial Sequence

<220>

<223> /note = "mutant luciferase"

<400> 47

Met Met Lys Arg Glu Lys Asn Val Ile Tyr Gly Pro Glu Pro Leu His

1	5	10	15
Pro	Leu	Glu	Asp
Leu	Thr	Ala	Gly
Glu	Met	Leu	Phe
Asp	Leu	Phe	Arg
Leu	Arg		
20	25	30	
Lys	His	Ser	His
Leu	Pro	Gln	Ala
Val	Asp	Val	Val
Gly	Asp	Glu	
35	40	45	
Ser	Leu	Ser	Tyr
Lys	Glu	Phe	Phe
Glu	Ala	Thr	Val
Leu	Leu	Leu	Ala
Ala	Gln		
50	55	60	
Ser	Leu	His	Asn
Cys	Gly	Tyr	Lys
Met	Asn	Asp	Val
Asp	Val	Ser	Ile
Val	Ser	Ile	Cys
65	70	75	80
Ala	Glu	Asn	Asn
Thr	Arg	Phe	Phe
Ile	Pro	Val	Ile
Ala	Ala	Ala	Trp
85	90	95	
Ile	Gly	Met	Ile
Val	Ala	Pro	Val
Asn	Glu	Ser	Tyr
Ile	Pro	Asp	Glu
100	105	110	
Leu	Cys	Lys	Val
Met	Gly	Ile	Ser
Lys	Pro	Gln	Ile
Val	Asn	Ile	Val
Phe	Thr	Thr	
115	120	125	
Lys	Asn	Ile	Leu
Asn	Val	Glu	Val
Leu	Glu	Val	Gln
Asn	Ser	Arg	Thr
Thr	Asn	Phe	
130	135	140	
Ile	Lys	Arg	Ile
Ile	Val	Leu	Asp
Asp	Thr	Val	Glu
Asn	Ile	His	Gly
145	150	155	160
Glu	Ser	Leu	Pro
Asn	Phe	Ile	Ser
Arg	Tyr	Ser	Asp
Tyr	Ser	Asp	Gly
Asn	Ile	Ala	
165	170	175	
Asn	Phe	Lys	Pro
Leu	His	Phe	Asp
Asp	Pro	Val	Glu
Gln	Val	Ala	Ala
180	185	190	
Leu	Cys	Ser	Ser
Gly	Thr	Thr	Gly
Leu	Pro	Lys	Gly
Val	Gly	Val	Met
Met	Gln	Thr	
195	200	205	
His	Gln	Asn	Ile
Ile	Cys	Val	Arg
Arg	Leu	Ile	His
Leu	Asp	Ala	Leu
Asp	Pro	Arg	Ala
210	215	220	
Gly	Thr	Gln	Leu
Leu	Ile	Pro	Gly
Gly	Val	Thr	Val
Val	Leu	Val	Tyr
Leu	Val	Tyr	Leu
Tyr	Leu	Pro	Phe
225	230	235	240
Phe	His	Ala	Phe
Gly	Phe	Ser	Ile
Phe	Ser	Ile	Thr
Ile	Gly	Tyr	Phe
Gly	Tyr	Phe	Met
245	250	255	
Leu	Arg	Val	Ile
Arg	Met	Ser	Arg
Arg	Phe	Asp	Pro
Phe	Asp	Glu	Ala
Ala	Phe	Leu	Lys
260	265	270	
Ala	Ile	Gln	Asp
Asp	Tyr	Glu	Val
Glu	Val	Arg	Ser
Arg	Ser	Val	Ile
Ser	Val	Ile	Asn
Ile	Asn	Val	Pro
275	280	285	
Ile	Leu	Phe	Leu
Leu	Ser	Lys	Ser
Ser	Pro	Leu	Val
Leu	Asp	Lys	Tyr
Asp	Lys	Tyr	Asp
290	295	300	
Ser	Leu	Arg	Glu
Glu	Leu	Cys	Cys
Leu	Cys	Gly	Ala
Cys	Gly	Ala	Ala
305	310	315	320
Ala	Glu	Val	Ala
Ala	Ala	Lys	Arg
Lys	Arg	Leu	Asn
Arg	Leu	Asn	Leu
Leu	Asn	Leu	Pro
Pro	Gly	Ile	Arg
Gly	Ile	Arg	Cys
325	330	335	
Phe	Gly	Leu	Thr
Gly	Leu	Thr	Ser
Leu	Ser	Ser	Ala
Ser	Ala	Asn	Ile
Ala	Asn	Ile	His
Asp			

340	345	350
Glu Phe Lys Pro Gly Ser Leu Gly Arg Val Thr Pro Leu Met Ala Ala		
355	360	365
Lys Ile Ala Asp Arg Glu Thr Gly Lys Ala Leu Gly Pro Asn Gln Val		
370	375	380
Gly Glu Leu Cys Ile Lys Gly Pro Met Val Ser Lys Gly Tyr Val Asn		
385	390	395
Asn Val Glu Ala Thr Lys Glu Ala Ile Asp Asp Asp Gly Trp Leu His		
405	410	415
Ser Gly Asp Phe Gly Tyr Tyr Asp Glu Asp Glu His Phe Tyr Val Val		
420	425	430
Asp Arg Tyr Lys Glu Leu Ile Lys Tyr Lys Gly Ser Gln Val Ala Pro		
435	440	445
Ala Glu Leu Glu Glu Ile Leu Leu Lys Asn Pro Cys Ile Arg Asp Val		
450	455	460
Ala Val Val Gly Ile Pro Asp Leu Glu Ala Gly Glu Leu Pro Ser Ala		
465	470	475
Phe Val Val Lys Gln Pro Gly Lys Glu Ile Thr Ala Lys Glu Val Tyr		
485	490	495
Asp Tyr Leu Ala Glu Arg Val Ser His Thr Lys Tyr Leu Arg Gly Gly		
500	505	510
Val Arg Phe Val Asp Ser Ile Pro Arg Asn Val Thr Gly Lys Ile Thr		
515	520	525
Arg Lys Glu Leu Leu Lys Gln Leu Leu Glu Lys Ala Gly Gly		
530	535	540

<210> 48

<211> 23

<212> DNA

<213> Artificial Sequence

<220>

<223> /note = "a primer"

<400> 48

ttatcccttg gaagatggga cg

23

<210> 49

<211> 22

<212> DNA

<213> Artificial Sequence

<220>
<223> /note = "a primer"

<400> 49
ctggagaaca gctgttgac gc 22

<210> 50
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> /note = "a primer"

<400> 50
gttatgcaga tattccgggc tgcatacgat tg 32

<210> 51
<211> 40
<212> DNA
<213> Artificial Sequence

<220>
<223> /note = "a primer"

<400> 51
cttggaaataa ttgtggcacc tgtaacgat aaatacattg 40

<210> 52
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> /note = "a primer"

<400> 52
aaccacgcat agtttttgc tcc 23

<210> 53
<211> 31
<212> DNA

<213> Artificial Sequence

<220>

<223> /note = "a primer"

<400> 53

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INTERNATIONAL SEARCH REPORT

Internal Application No
PCT/US 99/30925A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12N15/53 C12N15/10 C12N9/02

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

WPI Data, PAJ, CAB Data, STRAND, BIOSIS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>WO 99 14336 A (HALL MARY P ;PROMEGA CORP (US); WOOD KEITH V (US)) 25 March 1999 (1999-03-25)</p> <p>Fig.46,47 page 34, line 1 -page 42, line 6; table 6 page 47, line 3 -page 61, line 2</p> <p>---</p> <p>-/-</p>	<p>1-6,8, 12,14, 16,17, 19,21, 23,24, 28-37, 47-58, 65,66, 68,73,75</p>

 Further documents are listed in the continuation of box C. Patent family members are listed in annex.

° Special categories of cited documents :

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *&* document member of the same patent family

Date of the actual completion of the international search

7 August 2000

Date of mailing of the international search report

14.08.00

Name and mailing address of the ISA

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Authorized officer

Hornig, H

INTERNATIONAL SEARCH REPORT

Intern	Int'l Application No
PCT/US 99/30925	

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP 0 524 448 A (KIKKOMAN CORP) 27 January 1993 (1993-01-27) the whole document ---	1-5,8, 12,14, 16,17, 19,21, 23,24, 28-37, 58,65, 66,73
X	WO 95 25798 A (SECR DEFENCE BRIT ;LOWE CHRISTOPHER ROBIN (GB); WHITE PETER JOHN) 28 September 1995 (1995-09-28) the whole document ---	1-5,8, 12,14, 16,17, 19,21, 23,24, 28-37, 58,65, 66,73
X	WHITE ET AL: "Improved thermostability of the north american firefly luciferase: saturation mutagenesis at position 354" BIOCHEMICAL JOURNAL, GB, PORTLAND PRESS, LONDON, vol. 319, no. 319, 1996, pages 343-350-350, XP002097112 ISSN: 0264-6021 the whole document ---	1-5,8, 12,14, 16,17, 19,21, 23,24, 28-37, 58,65, 66,73
X	WO 98 46729 A (MURRAY JAMES AUGUSTUS HENRY ;SECR DEFENCE (GB); LOWE CHRISTOPHER R) 22 October 1998 (1998-10-22) the whole document ---	1-5,8, 12,14, 16,17, 19,21, 23,24, 28-37, 58,65, 66,73
A	the whole document --- K.V. WOOD ET AL.: "Bioluminescent click beetles revisited" J. BIOLUMINESCENCE AND CHEMILUMINESCENCE, vol. 4, 1989, pages 31-39, XP000906944 JOHN WILEY & SONS, LTD, NEW YORK, US cited in the application the whole document ---	
A	WO 95 18853 A (PROMEGA CORP) 13 July 1995 (1995-07-13) the whole document ---	
		-/-

INTERNATIONAL SEARCH REPORT

Intern: PCT/US Application No 99/30925

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	K.V. WOOD ET AL.: "Complementary DNA coding click beetle luciferases can elicit bioluminescence of different colors" SCIENCE, vol. 244, 12 May 1989 (1989-05-12), pages 700-702, XP002137278 AAAS, WASHINGTON, DC, US cited in the application the whole document ---	
A	K.V. WOOD ET AL.: "Introduction to beetle luciferases and their applications" J. BIOLUMINESCENCE AND CHEMILUMINESCENCE, vol. 4, no. 1, July 1989 (1989-07), pages 289-301, XP000906968 JOHN WILEY & SONS, LTD, NEW YORK, US cited in the application the whole document ---	
A	DEMENTIEVA E I ET AL: "PHYSICOCHEMICAL PROPERTIES OF RECOMBINANT LUCIOLA MINGRELICA LUCIFERASE AND ITS MUTANT FORMS" BIOCHEMISTRY, US, AMERICAN CHEMICAL SOCIETY. EASTON, PA, vol. 1, no. 61, 1 January 1996 (1996-01-01), pages 115-119-119, XP002078631 ISSN: 0006-2960 the whole document ---	
A	L. YE ET AL.: "Cloning and sequencing of a cDNA for firefly luciferase from Photuris pennsylvanica" BIOCHIMICA BIOPHYSICA ACTA, vol. 1339, 1997, pages 39-52, XP000909154 ELSEVIER SCIENCE, AMSTERDAM, NL cited in the application the whole document ---	
X	US 5 605 793 A (STEMMER WILLEM P C) 25 February 1997 (1997-02-25) cited in the application column 16, line 1 - line 6 column 16, line 50 - line 55 ---	47-60, 70-72, 74,75
X	WO 98 13487 A (MAXYGEN INC) 2 April 1998 (1998-04-02) page 39, line 30 - line 31; claims 1-57; figures 1,3 ---	58-60, 70-72,74
		-/-

INTERNATIONAL SEARCH REPORT

Intern al Application No
PCT/US 99/30925

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	ZHANG J-H ET AL: "Directed evolution of a fucosidase from galactosidase by DNA shuffling and screening" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA,US,NATIONAL ACADEMY OF SCIENCE. WASHINGTON, vol. 94, 1 April 1997 (1997-04-01), pages 4504-4509, XP002087464 ISSN: 0027-8424 the whole document ---	47-57,75
X	STEMMER W: "DNA shuffling by random fragmentatio and reassembly: In vitro recombination for molecular evolution" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA,US,NATIONAL ACADEMY OF SCIENCE. WASHINGTON, vol. 91, 1 October 1994 (1994-10-01), pages 10747-10751, XP002087463 ISSN: 0027-8424 the whole document ---	47-57,75
A	CADWELL R AND JOYCE G ET AL: "Randomization of genes by PCR mutagenesis" PCR METHODS AND APPLICATIONS,US,COLD SPRING HARBOR, NY, vol. 2, 1992, pages 28-33, XP002087462 ISSN: 1054-9803 the whole document ---	47-57,75
A	FROMANT M ET AL: "DIRECT RANDOM MUTAGENESIS OF GENE-SIZED DNA FRAGMENTS USING POLYMERASE CHAIN REACTION" ANALYTICAL BIOCHEMISTRY,US,ACADEMIC PRESS, SAN DIEGO, CA, vol. 224, no. 1, 1995, pages 347-353, XP000486749 ISSN: 0003-2697 cited in the application the whole document -----	47-57,75

INTERNATIONAL SEARCH REPORTInt ional application No.
PCT/US 99/30925**Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)**

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:

2. Claims Nos.: 76 (11-partially) because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:

In claim 11, SEQ ID Nos. 12 and 13 do not encode any amino acid sequence of claim 9 and 10.
Claim 76 refers to itself.

3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

The additional search fees were accompanied by the applicant's protest.
 No protest accompanied the payment of additional search fees.

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: (1-10,12-46,65-68,73)-complete, (11,58)-partially

A second luciferase that has enhanced resistance to an inhibitor of the luciferase relative to a first reference beetle luciferase; said second luciferase which comprises a plurality of amino acid substitutions relative to the reference beetle luciferase; said second luciferase wherein the reference luciferase is LucpPe-2 (luciferase from *Photuris pennsylvanica*); luciferases: luc 133-1B2 and luc146-1H2 which comprise SEQ ID Nos. 44 and 45; an isolated and purified nucleic acid molecule comprising a nucleic acid sequence selected from SEQ ID Nos. 42 and 43; a vector comprising said nucleic acid molecule; a host cell comprising said vector; a fusion protein comprising said luciferase; the use of the second luciferase for detecting ATP, for labeling a molecule, as a genetic reporter, for immobilization onto a solid surface, to produce a hybrid molecule, for high temperature reactions, or for creating luminescent solutions; a method for using said vector encoding said luciferase; a kit comprising: a container comprising said second luciferase; method to prepare an luciferase that is resistant to an inhibitor, comprising: a) selecting one or more isolated polynucleotide sequences encoding luciferase which is resistant to an inhibitor from a first population of polynucleotide sequences obtained from a first isolated polynucleotide sequence encoding luciferase subjected to conditions that yield nucleotide mutations, wherein the luciferase encoded by the one or more selected isolated polynucleotide sequences has increased resistance to an inhibitor relative to the luciferase encoded by the first isolated polynucleotide sequence; b) mutating the selected isolated polynucleotide sequence to yield a second population of polynucleotide sequences; and c) repeating step a) and step b) so as to yield a further polynucleotide sequence encoding luciferase that is resistant to an inhibitor and comprises a plurality of amino acid substitutions relative to the enzyme encoded by the first polynucleotide sequence; said method wherein the first polynucleotide sequence encodes Ppe2 and/or Ppl;

1.1. Claims: (1-10,12-46,65-68,73)-complete, (11,58)-partially

Idem as Invention 1, but limited to luc49-7C6, respectively SEQ ID Nos. 1 and 14, where the reference beetle luciferase is LucPpe-2;

1.2. Claims: (1-10,12-46,65-68,73)-complete, (11,58)-partially

Idem as Invention 1, but limited to luc78-0B10, respectively SEQ ID Nos. 6 and 19, where the reference beetle luciferase is lucPpe-2;

1.3. Claims: (1-10,12-46,65-68,73)-complete, (11, 58)-partially

Idem as Invention 1, but limited to luc90-1B5 respectively SEQ ID Nos. 11 and 24, where the reference beetle luciferase is lucPpe-2;

1.4. Claims: (1-10,12-46,65-68,73)-complete, (11, 58)-partially

Idem as Invention 1, but limited to luc81-6G01 respectively SEQ ID Nos. 13 and 26, where the reference beetle luciferase is lucPpl;

1.5. Claims: (1-10,12-46,65-68,73)-complete, (11, 58)-partially

Idem as Invention 1, but limited to luc81-0B11, respectively SEQ ID Nos. 46 and 47, where the reference beetle luciferase is lucPpl;

1.6. Claims: (1-10,12-46,65-68,73)-complete, (11, 58)-partially

Idem as Invention 1, but limited to lucPpe-2[T249M], respectively SEQ ID No. 12, where the reference beetle luciferase is lucPpe-2;

2. Claims: 47-57,75

A method to prepare an enzyme which is not a beetle luciferase and which has enhanced enzymological properties, comprising: a) selecting one or more isolated polynucleotide sequences encoding an enzyme which is not a luciferase and which has at least one enhanced enzymological property from a first population of polynucleotide sequences obtained from a first isolated polynucleotide sequence encoding an enzyme which is not a luciferase, wherein the first isolated polynucleotide sequence is subjected to conditions that yield nucleotide mutations, wherein the enzyme encoded by the one or more selected isolated polynucleotide sequences has at least one enhanced enzymological property relative to the enzyme encoded by the first isolated polynucleotide sequence; b) mutating the selected isolated polynucleotide sequence to yield a second population of polynucleotide sequences, wherein the selected isolated polynucleotide

sequence is subjected to oligonucleotide mediated mutagenesis with a plurality of oligonucleotides each comprising at least one codon that encodes a consensus amino acid which is not present in the first polynucleotide sequence; and c) repeating step a) and step b) so as to yield a further polynucleotide sequence encoding an enzyme which is not a luciferase having at least one enhanced enzymological property and comprising a plurality of amino acid substitutions relative to the enzyme encoded by the first polynucleotide sequence; said method wherein the enzyme is DNA polymerase or RNA polymerase, chloramphenicol-acetyltransferase, beta-glucuronidase or beta-galactosidase; an enzyme which is encoded by the polynucleotide sequence of said method;

3. Claims: 58-partially, (59-64,69-72,74)-complete

A method to prepare an enzyme that is resistant to an inhibitor, comprising: a) selecting one or more isolated polynucleotide sequences encoding an enzyme which is resistant to an inhibitor from a first population of polynucleotide sequences obtained from a first isolated polynucleotide sequence encoding an enzyme subjected to conditions that yield nucleotide mutations, wherein the enzyme encoded by the one or more selected isolated polynucleotide sequences has increased resistance to an inhibitor relative to the enzyme encoded by the first isolated polynucleotide sequence; b) mutating the selected isolated polynucleotide sequence to yield a second population of polynucleotide sequences; and c) repeating step a) and step b) so as to yield a further polynucleotide sequence encoding an enzyme that is resistant to an inhibitor and comprises a plurality of amino acid substitutions relative to the enzyme encoded by the first polynucleotide sequence; said method wherein the further polynucleotide sequence encodes an enzyme that has increased thermostability relative to the first polynucleotide sequence, said method wherein said enzyme is DNA or RNA polymerase; a polynucleotide sequence obtained by said m

Please note that all inventions mentioned under item 1, although not necessarily linked by a common inventive concept, could be searched without effort justifying an additional fee.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 76 (11-partially)

In claim 11, SEQ ID Nos. 12 and 13 do not encode any amino acid sequence of claim 9 and 10.

Claim 76 refers to itself.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

Information on patent family members

Internal Application No

PCT/US 99/30925

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO 9914336	A 25-03-1999	AU 9492198	A	05-04-1999
		EP 1015601	A	05-07-2000
EP 0524448	A 27-01-1993	DE 69229376	D	15-07-1999
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